PART IV

SPINE

- Section 1: Prevention
 - 1.1. GENERAL PRINCIPLES
 - 1.2. Antimicrobials
 - 1.3. Bone Graft
 - 1.4. RISK FACTORS
 - 1.5. Wound Care
- Section 2: Diagnosis
 - 2.1. General Principles
 - 2.2. BIOMARKERS
 - 2.3. Imaging
- Section 3: Treatment
 - 3.1. General Principles
 - 3.2. Antibiotics
 - 3.3. Implants
 - 3.4. Wound Care

Prevention

1.1. PREVENTION: GENERAL PRINCIPLES

Authors: Steven Schmitt, Christopher Kepler

QUESTION 1: What can one do if an inadvertent contamination during instrumented spine surgery occurs?

RECOMMENDATION: There is no data to support a particular strategy in preventing infection after inadvertent contamination of spinal implants.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Left uncovered in the operating suite, spinal implants can become contaminated within 30 minutes [1]. There are no human data to support a particular algorithm for management of inadvertent contamination. In animal studies, tobramycin powder has been shown to reduce infection in contaminated spine surgery and vancomycin powder has been shown to reduce infection in contaminated knee surgery [2,3]. At least one suggests that management of inadvertent contamination should be individualized to the clinical situation and stage of surgery, and many surgeons are reluctant to proceed with implant surgery if contamination has occurred. Some experts recommend intraoperative irrigation with solutions containing antibiotics, without supporting data (personal communication).

REFERENCES

- [1] Menekse G, Kuscu F, Suntur BM, Gezercan Y, Ates T, Ozsoy KM, et al. Evaluation of the time-dependent contamination of spinal implants: prospective randomized trial. Spine. 2015;40:1247–1251. doi:10.1097/BRS.0000000000000044.
- [2] Laratta JL, Shillingford JN, Hardy N, Lombardi JM, Saifi C, Romanov A, et al. Intrawound tobramycin powder eradicates surgical wound contamination: an in vivo rabbit study. Spine. 2017;42:E1393-E1397. doi:10.1097/BRS.0000000000002187.
- [3] Edelstein Al, Weiner JA, Cook RW, Chun DS, Monroe E, Mitchell SM, et al. Intra-articular vancomycin powder eliminates methicillin-resistant S. aureus in a rat model of a contaminated intra-articular implant. J Bone Joint Surg Am. 2017;99:232–238. doi:10.2106/JBJS.16.00127.

Author: Maja Babic

QUESTION 2: How should spine surgery patients with postoperative diarrhea be managed?

RECOMMENDATION: Diarrhea can be managed in a standard approach with careful attention to the surgical site.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Postoperative diarrhea poses a risk of contaminating the surgical incision. Maintaining a clean and dry surgical site is crucial. Postoperative diarrhea is generally self-limiting but infectious etiologies, especially *C. difficile*, are particularly concerning in the inpatient setting and should be ruled out. After infectious causes are ruled out, a standard approach should be implemented to address diarrhea including discontinuing potentially inciting medication (antibiotics), increasing fiber content and using antisecretory (i.e., bismuth subsalicylate) and antimotility (i.e., loperamide) agents. A balanced

electrolyte rehydration should also be utilized. The use of probiotics and prebiotics can be used in cases of post-antibiotic-associated illness [1].

REFERENCES

[1] Riddle MS, DuPont HL, Connor BA. ACG Clinical Guideline: diagnosis, treatment, and prevention of acute diarrheal infections in adults. Am J Gastroenterol. 2016;111:602–622. doi:10.1038/ajg.2016.126.

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1.2. PREVENTION: ANTIMICROBIALS

Authors: Alexander Montgomery, Rajesh Mangattil

QUESTION 1: Is there a role for oral antibiotics in the prevention of infection in patients with draining wounds following spinal surgery?

RECOMMENDATION: There is no reliable evidence for the use of prophylactic oral antibiotic therapy in patients with draining wounds after spine surgery.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

The incidence of spinal surgical site infection (SSI) has been reported to be from 0.7–16% [1–3]. Surgical drains are used in spine surgery to avoid the risk of a hematoma formation leading to potential neurological deficit [4]. Drains retained for a longer period have been shown to have a higher rate of bacterial contamination [5]. However, not using a drain has been found to be associated with the development of late-onset SSI [6,7]. Therefore, the use of drains decreases wound drainage and consequently decreases infection rates [8,9]. Prophylactic antibiotic cover for 24 hours has now become a standard of care following orthopaedic procedures [10].

Since the first systematic review on prophylactic measures against spinal SSI was published by Brown et al. in 2004 [11], there has been a considerable increase in the preventive strategies documented in the spine literature. However, many studies are of lower methodological quality with significant heterogeneity [12].

There was only one prospective randomized study showing no significant difference in the infection rates between patients receiving prophylactic antibiotic coverage for 24 hours or for the entire duration that the drain was in place. This study was on thoracolumbar fractures. It was not clear if the antibiotic cover was administered orally or parenterally [13]. In a review of 560 cases of closed suction drainage in single level lumbar decompressions, Kanayama et al. did not report on the use of prophylactic oral antibiotics [14]. Similarly, a 2018 systematic review by Yao et al. identified 11 randomized controlled trials (RCTs), 51 case-controlled studies (CCS) and 77 case series. They reported wide variations in the surgical indications, approaches and definitions of SSI. They found strong evidence that closed-suction drainage does not affect SSI rates, but had no mention of the use of prophylactic oral antibiotic therapy [15].

There were many studies that evaluated the risk factors for wound complications following spine surgery [16–18]. Past studies are archaic in nature with very little contribution or relevance to these authors. A staged treatment algorithm for spine infections did not specify or address the indication for oral antibiotics to prevent infection in draining wounds [19]. A recent retrospective study attributed the drain volume and time to the risk factors for SSI after lumbar surgery. There was no direct reference to the impact of oral or parenteral antibiotics in their study [13,20].

A systematic evidenced-based review included 36 observational studies involving 2,439 patients. However, these were non-interventional studies to evaluate the independent risk factors for patients developing SSIs following spine surgery [17]. In their systematic review and meta-analysis of wound drains in non-instrumented

lumbar decompression surgery, Davidoff et al. included 5,327 cases who received drains. They found that the SSI rates were unaffected by the routine use of drains. However, none of these patients had prophylactic oral antibiotics [21]. Ho et al. reported a retrospective review of 70 patients who had undergone single-level lumbar discectomy. They suggested that surgical drains do not increase SSI risk and that drain tip cultures allow detection of postoperative infection at a very early stage. They found that this would lead to quicker initiation of antibiotic treatment [22].

Apart from a prospective randomized study that suggested no difference in the infection rates, there are no studies directly linking the role of oral antibiotics in the prevention of infection in patients with draining wounds following spine surgery [13]. Therefore, in the absence of reliable evidence, only a consensus recommendation can be made based on clinical opinion.

- [1] Glassman SD, Dimar JR, Puno RM, Johnson JR. Salvage of instrumental lumbar fusions complicated by surgical wound infection. Spine (Phila Pa 1976). 1996 Sep 15;21(18):2163–2169.
- 2] Stone PW. Economic burden of healthcare-associated infections: an American perspective. Expert Rev Pharmacoecon Outcomes Res. 2009;9(5):417–422.
 3] Whitehouse JD, Friedman ND, Kirkland KB, Richardson WJ, Sexton DJ.
- [3] Whitehouse JD, Friedman ND, Kirkland KB, Richardson WJ, Sexton DJ. The impact of surgical-site infections following orthopedic surgery at a community hospital and a university hospital: adverse quality of life, excess length of stay, and extra cost. Infect Control Hosp Epidemiol. 2002 ADIZ3(4):183-180.
- [4] Kou J, Fischgrund J, Biddinger A, Herkowitz H. Risk factors for spinal epidural hematoma after spinal surgery. Spine (Phila Pa 1976). 2002;27(15):1670–1673.
- [5] Drinkwater CJ, Neil MJ. Optimal timing of wound drain removal following
- total joint arthroplasty. J Arthroplasty. 1995 Apr;10(2):185–189.

 Ho C, Sucato DJ, Richards BS: Risk factors for the development of delayed infections following posterior spinal fusion and instrumentation in adolescent idiopathic scoliosis patients. Spine (Phila Pa 1976). 2007;32:2272–2277.
- [7] Sankar B, Ray P, Rai J. Suction drain tip culture in orthopaedic surgery: a prospective study of 214 clean operations. Int Orthop. 2004 Oct;28(5):311–314. Epub 2004 Aug 14.
- [8] Massie JB, Heller JG, Abitbol JJ, McPherson D, Garfin SR. Postoperative posterior spinal wound infections. Clin Orthop Relat Res. 1992 Nov;(284):99–108.
- [9] Ho C, Sucato DJ, Richards BS. Risk factors for the development of delayed infections following posterior spinal fusion and instrumentation in adolescent idiopathic scoliosis patients. Spine (Phila Pa 1976). 2007;32(20):2272-2277.
- [10] Prokuski L. Prophylactic antibiotics in orthopaedic surgery. J Am Acad Orthop Surg. 2008 May;16(5):283-293.
 [11] Brown MD, Brookfield KF. A randomized study of closed wound suction
- [11] Brown MD, Brookfield KF. A randomized study of closed wound suction drainage for extensive lumbar spine surgery. Spine (Phila Pa 1976). 2004;29(10):1066-1068
- [12] Van Middendorp JJ, Pull ter Gunne AF, Schuetz M, Habil D, Cohen DB, Hosman AJF, et al. A methodological systematic review on surgical site infections following spinal surgery. Part 2: prophylactic treatments. Spine (Phila Pa 1976). 2012;37(24):2034-2045.

- [13] Takemoto RC, Lonner B, Andres T. Appropriateness of twenty-four-hour antibiotic prophylaxis after spinal surgery in which a drain is utilized: a prospective randomized study. J Bone Joint Surg Am. 2015;97(12):979–986.
- prospective randomized study. J Bone Joint Surg Am. 2015;97(12):979–986.

 [14] Kanayama M, Oha F, Togawa D, Shigenobu K, Hashimoto T. Is closed-suction drainage necessary for single-level lumbar decompression?: review of 560 cases. Clin Orthon Relat Res. 2010;468(10):2600–2604
- cases. Člin Orthop Relat Res. 2010;468(10):2690–2694.

 [15] Yao R, Tan T, Tee JW, Street J. Prophylaxis of surgical site infection in adult spine surgery: a systematic review. J Clin Neurosci. 2018;52:5–25. doi: 10.1016/j. jocn.2018.03.023.
- [16] Piper KF, Tomlinson SB, Santangelo G, Van Galen J, DeAndrea-Lazarus I. Risk factors for wound complications following spine surgery. Surg Neurol Int. 2017;8:269. doi: 10.4103/sni.sni_306_17. eCollection 2017.
 [17] Shoji H, Hirano T, Watanabe K, Ohashi M, Mizouchi T, Endo N. Risk factors
- [17] Shoji H, Hirano T, Watanabe K, Ohashi M, Mizouchi T, Endo N. Risk factors for surgical site infection following spinal instrumentation surgery. J Orthop Sci. 2018;23(3):449–454. doi: 10.1016/j.jos.2018.02.008. Epub 2018 Mar 2.
- [18] Xing D, Ma JX, Ma XL, et al. A methodological, systematic review of evidence based independent risk factors for surgical site infections after spinal surgery. Eur Spine J. 2013;22(3):605–615.
- surgery. Eur Spine J. 2013;22(3):605–615.
 [19] Stüer C, Stoffel M, Hecker J, Ringel F, Meyer B. A staged treatment algorithm for spinal infections. J Neurol Surg A Cent Eur Neurosurg. 2013;74(2):87–95.
 doi: 10.1055/8-0032-1320022. Epub 2013 Feb 12.
- doi: 10.1055/s-0032-1320022. Epub 2013 Feb 12.

 [20] Ahmed R, Greenlee JD, Traynelis VC. Preservation of spinal instrumentation after development of postoperative bacterial infection in patients undergoing spinal arthrodesis. J Spinal Disord Tech. 2012;25(6):299–302.
- Davidoff CL, Rogers JM, Simons M, Davidson AS. A systematic review and meta-analysis of wound drains in non-instrumented lumbar decompression surgery. J Clin Neurosci. 2018;53:55-61. doi: 10.1016/j.jocn.2018.04.038.
 Kobayashi K, Imagama S, Ito Z, Ando K, Yagi H, Hida T, et al. Is a drain tip
- [22] Kobayashi K, İmagama S, Ito Z, Ando K, Yagi H, Hida T, et al. Is a drain tip culture required after spinal surgery? J Spinal Disord Tech. 2017;30(8):356– 359.

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QUESTION 2: Is there a role for the addition of gentamicin to perioperative prophylactic antibiotics in spine surgery?

RECOMMENDATION: No, we recommend AGAINST the inclusion of gentamicin for perioperative prophylaxis in spine surgery. There is no data suggesting that the addition of gentamicin to systemic perioperative prophylactic antibiotic regimens decreases the rate of postoperative infections, and strong evidence showed that it is associated with harm (namely nephrotoxicity). The question of the use of local/topical gentamicin is unresolved.

LEVEL OF EVIDENCE: Strong

DELEGATE VOTE: Agree: 62%, Disagree: 15%, Abstain: 23% (Super Majority, Weak Consensus)

RATIONALE

The use of gentamicin to expand the gram-negative activity for perioperative antimicrobial prophylaxis in spine surgery has been considered for decades, yet positive outcomes data for this practice are lacking. Pons et al. reported on a randomized, blinded study of 826 patients undergoing neurosurgical procedures, including spine surgery, and found similar surgical site infection (SSI) rates for those assigned to ceftizoxime or vancomycin and gentamicin [1]. Ramo et al. reported on a multivariate analysis of 428 posterior spinal fusion patients and found that the addition of an aminogly-coside did not lower the SSI rate [2]. In a mixed population of more than 11,000 orthopaedic surgery patients treated over 5 years in the United Kingdom, Walker et al. noted no difference in SSI rates during a period when a combination of flucloxacillin and gentamicin was given for prophylaxis compared to one where co-amoxiclav was the prophylactic regimen of choice [3].

The association of aminoglycoside prophylaxis (even singledose) for orthopaedic surgery and acute kidney injury (AKI) has now been well-documented. Dubrovskaya et al. reviewed more than 4,000 patients undergoing orthopaedic surgery, comparing those receiving a single dose of gentamicin combined with another antibiotic to those receiving non-aminoglycoside prophylaxis alone. Although for all patients the addition of gentamicin was not associated with AKI, gentamicin was associated with a statistically significantly higher rate of AKI for those undergoing spine surgery [4]. Bell et al. reported on a Scottish initiative where routine surgical prophylaxis was changed from cefuroxime to flucloxacillin and gentamicin (single-dose) between 2006 and 2010. Among 7,666 patients undergoing orthopaedic surgery, the gentamicin-containing regimen was associated with a 94% higher incidence of AKI [5]. Finally, in the previously-cited study by Walker et al., a change from routine prophylaxis with flucloxacillin and gentamicin to co-amoxiclav alone was associated with a 63% reduction in postoperative AKI [3].

Two meta-analyses on the association of gentamicin prophylaxis with nephrotoxicity have been published. Luo et al. compared the use of gentamicin and flucloxacillin to cefuroxime alone in studies of diverse surgery types. The risk of postoperative renal impairment was higher in the gentamicin group, especially for those undergoing orthopaedic surgery [6]. Srisung et al. analyzed 11 studies containing 18,354 patients comparing gentamicin versus non-gentamicin surgical prophylaxis regimens. Using random effects modeling, gentamicin prophylaxis in orthopaedic surgery was associated with a significantly higher risk of AKI (risk rate (RR) 2.99; 95% confidence interval (CI): 1.84, 4.88) [7].

Data regarding the use of topical or local wound gentamicin are limited. In a single-center study, van Herwijnen et al. reported a higher SSI rate for patients undergoing scoliosis surgery who received wound irrigation with gentamicin versus povidone-iodine [8]. On the other hand, Borkhuu et al. reported on 220 children undergoing spinal fusion and found a four-fold reduction in SSI for those treated with gentamicin-impregnated bone allograft [9]. Han et al. retrospectively analyzed data from 399 patients undergoing spine surgery. Among patients who had a gentamicin-impregnated collagen sponge applied to their wound, the SSI rate was 0.8%, versus 5% for those treated without the sponge [10]. At this time, however, given the variability in reported application methods for local gentamicin and the small number of patients studied, the routine use of topical gentamicin cannot be recommended.

REFERENCES

 Pons VG, Denlinger SL, Guglielmo BJ, Octavio J, Flaherty J, Derish PA, et al. Ceftizoxime versus vancomycin and gentamicin in neurosurgical prophylaxis: a randomized, prospective, blinded clinical study. Neurosurgery 1993;33:416-422; discussion 422-423.

- Ramo BA, Roberts DW, Tuason D, McClung A, Paraison LE, Moore HG, et al. Surgical site infections after posterior spinal fusion for neuromuscular scoliosis: a thirty-year experience at a single institution. J Bone Joint Surg
- Am. 2014;96:2038–2048. doi:10.2106/JBJS.N.00277. Walker H, Patton A, Bayne G, Marwick C, Sneddon J, Davey P, et al. Reduction in post-operative acute kidney injury following a change in antibiotic prophylaxis policy for orthopaedic surgery: an observational study. J Antimicrob Chemother. 2016;71:2598–2605. doi:10.1093/jac/dkw166.
- Dubrovskaya Y, Tejada R, Bosco J, Stachel A, Chen D, Feng M, et al. Single high dose gentamicin for perioperative prophylaxis in orthopedic surgery: evaluation of nephrotoxicity. SAGE Open Med. 2015;3:2050312115612803. doi:10.1177/2050312115612803.
- Bell S, Davey P, Nathwani D, Marwick C, Vadiveloo T, Sneddon J, et al. Risk of AKI with gentamicin as surgical prophylaxis. J Am Soc Nephrol. 2014;25:2625–2632. doi:10.1681/ASN.2014010035.
- Luo S, Lai Y, Liu C, Chen Y, Qiao X. Prophylactic use of gentamicin/flucloxacillin versus cefuroxime in surgery: a meta analysis of clinical studies. Int J Clin Exp Med. 2015;8:17856–17867.
- Srisung W, Teerakanok J, Tantrachoti P, Karukote A, Nugent K. Surgical prophylaxis with gentamicin and acute kidney injury: a systematic review and meta-analysis. Ann Transl Med. 2017;5:100. doi:10.21037/atm.2017.03.06.
- van Herwijnen B, Evans NR, Dare CJ, Davies EM. An intraoperative irrigation regimen to reduce the surgical site infection rate following adolescent idiopathic scoliosis surgery. Ann R Coll Surg Engl. 2016;98:320-323. doi:10.1308/
- Borkhuu B, Borowski A, Shah SA, Littleton AG, Dabney KW, Miller F. Antibiotic-loaded allograft decreases the rate of acute deep wound infection after spinal fusion in cerebral palsy. Spine. 2008;33:2300-2304. doi:10.1097/ BRS.obo13e31818786ff.
- Han JS, Kim SH, Jin SW, Lee SH, Kim BJ, Kim SD, et al. The use of gentamicinimpregnated collagen sponge for reducing surgical site infection after spine surgery. Korean J Spine. 2016;13:129–133. doi:10.14245/kjs.2016.13.3.129.

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QUESTION 3: Should prophylactic antibiotic prophylaxis be repeated during spine surgery? If so, when?

RECOMMENDATION: In most uncomplicated spinal procedures, a single preoperative dose of prophylactic antibiotics is sufficient. Prophylactic antibiotics should be redosed intraoperatively for procedures lasting longer than twice the half-life of the antibiotic, or if there is excessive blood loss (blood loss > 1,500 mL) in order to ensure therapeutic levels.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

There are no randomized spine studies that compare the effectiveness of redosing prophylactic antibiotics during surgery to preoperative antibiotics alone. Therefore, this review was expanded to include other surgical subspecialties. Several major guidelines including those from the North American Spine Society (NASS), Infectious Disease Society of America (IDSA) and Surgical Infection Society (SIS) have made similar recommendations supported by pharmacokinetic data and retrospective studies [1,2]. Furthermore, the Centers for Disease Control and Prevention (CDC) recently noted that there is insufficient-quality evidence to make a recommendation regarding whether or not antibiotics should be redosed intra-

In a prospective study of 57 subjects undergoing elective surgery, an analysis of intraoperative serum cefazolin concentrations at approximately 3.5 hours after receiving a preoperative dose showed that antibiotic concentrations dropped below the minimum inhibitory concentration (MIC) for methicillin-susceptible Staphylococcus aureus (MSSA) and Escherichia Coli (E. Coli) [4]. Ohge and colleagues found that cefazolin concentrations had dropped below 80% of the MIC in the adipose tissue and peritoneum for multiple bacteria three hours after the preoperative dose was administered [5]. In a prospective study of 11 elective instrumented spinal procedures with a large expected blood loss, estimated blood loss (EBL) was found to have a strong negative correlation with cefazolin tissue concentrations (r = -0.66, p = 0.5). Based on the pharmacokinetic values, the authors recommended that procedures with an EBL greater than 1,500 mL should receive an additional dose of cefazolin [6].

In a retrospective study of 1,548 patients undergoing cardiac surgery, intraoperative redosing for procedures lasting greater than 400 minutes was shown to reduce the risk of surgical site infections (SSIs) (adjusted OR 0.44, 95% CI 0.23-0.86) [7]. Similarly, Scher et al. demonstrated that for surgeries longer than three hours, patients who were redosed with cefazolin intraoperatively had a lower SSI rate than those who only received preoperative cefazolin (6.1% vs. 1.3%, p < 0.01) [8]. In another retrospective review of 4,078 patients undergoing various general surgery procedures, cases with an EBL of greater than 500 mL or those that were not re-dosed intraoperatively during longer cases were associated with a higher rate of SSI [9].

- Shaffer WO, Baisden JL, Fernand R, Matz PG, North American Spine Society.
- An evidence-based clinical guideline for antibiotic prophylaxis in spine surgery. Spine J. 2013;13(10):1387-1392. doi:10.1016/j.spinee.2013.06.030. Bratzler DW, Dellinger EP, Olsen KM, et al. Clinical practice guidelines for antimicrobial prophylaxis in surgery. Am J Health Syst Pharm. 2013;70(3):195-283. doi:10.2146/ajhp120568
- Berrios-Torres SI, Umscheid CA, Bratzler DW, et al. Centers for Disease Control and Prevention guideline for the prevention of surgical site infection, 2017. JAMA Surg. 2017;152(8):784-791. doi:10.1001/jamasurg.2017.0904. Koopman E, Nix DE, Erstad BL, et al. End-of-procedure cefazolin concentra-
- tions after administration for prevention of surgical-site infection. Am J Health-Syst Pharm. 2007;64(18):1927-1934. doi:10.2146/ajhpo70047. Ohge H, Takesue Y, Yokoyama T, et al. An additional dose of cefazolin for
- intraoperative prophylaxis. Surg Today. 1999;29(12):1233-1236. doi:10.1007/
- Swoboda SM. Does intraoperative blood loss affect antibiotic serum and tissue concentrations? Arch Surg. 1996;131(11):1165. doi:10.1001/archsurg.1996.01430230047009. Zanetti G, Giardina R, Platt R. Intraoperative redosing of cefazolin and risk
- for surgical site infection in cardiac surgery. Emerg Infect Dis. 2001;7(5):828-
- Scher KS. Studies on the duration of antibiotic administration for surgical prophylaxis. Am Surg. 1997;63(1):59-62.
- Kasatpibal N, Whitney JD, Dellinger EP, Nair BG, Pike KC. Failure to redose antibiotic prophylaxis in long surgery increases risk of surgical site infection. Surg Infect. 2017;18(4):474–484. doi:10.1089/sur.2016.164.

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QUESTION 4: Should vancomycin powder be applied to the wound in patients undergoing spinal surgeries? Are there any potential harms associated with this practice?

RECOMMENDATION: Yes. Evidence suggests that vancomycin powder applied to the wound during spinal surgery reduces the risk of infection. However, the majority of studies lack a control arm and it is not known if vancomycin powder is better than antiseptic agents. There is insufficient evidence for or against the potential harm associated with this practice.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 79%, Disagree: 14%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Surgical site infection is a known risk of spine surgery with or without instrumentation, and gram-positive organisms are the most common pathogens in such infections. Many practitioners now apply vancomycin powder intraoperatively to reduce the risk of infection. Given concern for vancomycin's adverse effects and antimicrobial resistance, it is critical to consider a risk-benefit analysis of this practice.

A number of studies addressed the efficacy of vancomycin powder use in spine surgery. These have been the subject of several systematic reviews. Xie et al. reviewed 19 retrospective cohort studies and 1 prospective case study, with results suggesting benefit in all but 2 of these with an overall infection risk of 2.83-fold higher for patients not receiving vancomycin powder compared to those receiving it [1]. The authors pointed out study heterogeneity with regard to powder, drug dosage and exposure of bone graft and instrumentation to the drug, citing these as areas for future investigation. This trend toward benefit was confirmed in five other systematic reviews [2-6].

With regard to adverse effects, Ghobrial et al. performed a systematic review of 16 studies with 6,701 patients [7]. Of these, 1 patient developed nephropathy, 2 patients experienced hearing loss, 1 patient had an elevated vancomycin level and 19 patients developed culture-negative seroma. The authors highlighted the lack of in vivo evidence regarding vancomycin resistance. There was a trend toward gram-negative and polymicrobial infections among vancomycin powder recipients in one study [8].

REFERENCES

- Xie LL, Zhu J, Yang MS, et al. Effect of intra-wound vancomycin for spinal surgery: a systematic review and meta-analysis. Orthop Surg. 2017;9(4):350-358. doi:10.1111/os.12356.
- Kang DG, Holekamp TF, Wagner SC, Lehman RA. Intrasite vancomycin powder for the prevention of surgical site infection in spine surgery: a systematic literature review. Spine J. 2015;15(4):762–770. doi:10.1016/j. spinee.2015.01.030.
- Xiong L, Pan Q, Jin G, Xu Y, Hirche C. Topical intrawound application of vancomycin powder in addition to intravenous administration of antibi-
- otics: a meta-analysis on the deep infection after spinal surgeries. Orthop Traumatol Surg Res. 2014;100(7):785-789. doi:10.1016/j.otsr.2014.05.022. Ghobrial GM, Thakkar V, Andrews E, et al. Intraoperative vancomycin use in spinal surgery: single institution experience and microbial trends. Spine (Phila Pa 1976). 2014;39(7):550-555. doi:10.1097/BRS.0000000000000241. Bakhsheshian J, Dahdaleh NS, Lam SK, Savage JW, Smith ZA. The use of
- vancomycin powder in modern spine surgery: systematic review and meta-analysis of the clinical evidence. World Neurosurg. 2015;83(5):816–823. doi:10.1016/j.wneu.2014.12.033. Khan NR, Thompson CJ, DeCuypere M, et al. A meta-analysis of spinal
- surgical site infection and vancomycin powder. J Neurosurg Spine. Ghobrial GM, Cadotte DW, Williams K, Fehlings MG, Harrop JS.
- Complications from the use of intrawound vancomycin in lumbar spinal surgery: a systematic review. Neurosurg Focus. 2015;39(4):E11. doi:10.3171/2015.7.FOCUS15258.
- Adogwa O, Elsamadicy AA, Sergesketter A, et al. Prophylactic use of intraoperative vancomycin powder and postoperative infection: an analysis of microbiological patterns in 1,200 consecutive surgical cases. J Neurosurg Spine. 2017;27(3):328-334. doi:10.3171/2017.2.SPINE161310.

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QUESTION 5: What is the optimal perioperative antibiotic prophylaxis for patients undergoing spine surgery? What considerations should be made in cases of drug allergies?

RECOMMENDATION: The optimal prophylactic antibiotic for an uncomplicated spine surgery is a first- or second-generation cephalosporin given intravenously within 60 minutes of initial incision.

In patients with a history of anaphylactic reaction after use of beta lactams or in countries with a high rate of methicillin-resistant Staphylococcal infections, vancomycin in a weight-adjusted dose (15 mg/kg) should be used. Clindamycin 600 mg intravenously is an alternative to vancomycin.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 79%, Disagree: 7%, Abstain: 14% (Super Majority, Strong Consensus)

RATIONALE

Current literature supports the use of prophylactic antibiotics for spinal procedures with or without instrumentation to decrease the risk of surgical site infections (SSI), with a first- or second-generation cephalosporin being the antibiotic of choice [1–6]. In addition, clinical guidelines set forth by the American Society of Health-System Pharmacists (ASHP), the Infectious Diseases Society of America (IDSA), the Surgical Infection Society (SIS), the Society for Healthcare Epidemiology of America (SHEA) and the North American Spine Society support the use of first-generation cephalosporins [1,7,8]. Although comparative studies to evaluate the optimal timing for preoperative antibiotic have not been conducted for spine surgery, it is well-established that intravenous cephalosporins given within 60 minutes before initial incision is effective [9,10].

In a comparative study evaluating the addition of vancomycin powder for posterior thoracic and lumbar spine surgery, Sweet et al. found that vancomycin powder reduced the rate of SSI compared to intravenous cephalexin alone (0.2% vs. 2.6%, p < 0.0001).

Regarding prophylaxis regiments combining antibiotic agents, randomized clinical trials exist which show a reduced rate of postsurgical infections if a combination of a cephalosporin and gentamicin or vancomycin and gentamicin is used, compared to placebo [11,12]. However, there are no studies available which compare combination regimens with the standard prophylaxis with cefazolin. A study by Pons et al. comparing ceftizoxime versus the combination prophylaxis with vancomycin and gentamicin found no decreased infection rate, but higher toxicity with the combination regimen [13].

There is no specific recommendation for adapted prophylaxis in obese patients in spine surgery. However, in periprosthetic joint infections, adaptation is discussed in patients with a weight more than 100 kg since infection rate was twice that in other patients [13-15].

REFERENCES

Bratzler DW, Dellinger EP, Olsen KM, Perl TM, Auwaerter PG, Bolon MK, et al. Clinical practice guidelines for antimicrobial prophylaxis in surgery. Surg Infect (Larchmt). 2013;14:73–156. doi:10.1089/sur.2013.9999.

- Pavel A, Smith RL, Ballard A, Larson IJ. Prophylactic antibiotics in elective orthopedic surgery: a prospective study of 1,591 cases. South Med J 1977;70
- Barker FG. Efficacy of prophylactic antibiotic therapy in spinal surgery: a meta-analysis. Neurosurgery. 2002;51:391-400; discussion 400-401.
- Schnöring M, Brock M. [Prophylactic antibiotics in lumbar disc surgery: analysis of 1,030 procedures]. Zentralbl Neurochir. 2003;64:24-29. doi:10.1055/s-2003-37148.
- Rubinstein E, Findler G, Amit P, Shaked I. Perioperative prophylactic cephazolin in spinal surgery. A double-blind placebo-controlled trial. J
- Bone Joint Surg Br. 1994;76:99-102. Bratzler DW, Houck PM, Surgical Infection Prevention Guidelines Writers Workgroup, American Academy of Orthopaedic Surgeons, American Association of Critical Care Nurses, American Association of Nurse Anesthetists, et al. Antimicrobial prophylaxis for surgery: an advisory statement from the National Surgical Infection Prevention Project. Clin Infect Dis. 2004;38:1706-1715. doi:10.1086/421095
- Labbé AC, Demers AM, Rodrigues R, Arlet V, Tanguay K, Moore DL. Surgicalsite infection following spinal fusion: a case-control study in a children's hospital. Infect Control Hosp Epidemiol. 2003;24:591–595. doi:10.1086/502259.
- Shaffer WO, Baisden JL, Fernand R, Matz PG, North American Spine Society. An evidence-based clinical guideline for antibiotic prophylaxis in spine
- surgery. Spine J. 2013;13:1387-1392. doi:10.1016/j.spinee.2013.06.030. Classen DC, Evans RS, Pestotnik SL, Horn SD, Menlove RL, Burke JP. The timing of prophylactic administration of antibiotics and the risk of surgical-wound infection. N Engl J Med. 1992;326:281-286. doi:10.1056/ NEJM199201303260501.
- Weber WP, Mujagic E, Zwahlen M, Bundi M, Hoffmann H, Soysal SD, et al. Timing of surgical antimicrobial prophylaxis: a phase 3 randomised controlled trial. Lancet Infect Dis. 2017;17:605-614. doi:10.1016/S1473-3099(17)30176-7.
- Young RF, Lawner PM. Perioperative antibiotic prophylaxis for prevention of postoperative neurosurgical infections. A randomized clinical trial. J Neurosurg. 1987;66:701–705. doi:10.3171/jns.1987.66.5.0701. Geraghty J, Feely M. Antibiotic prophylaxis in neurosurgery. A randomized
- controlled trial. J Neurosurg. 1984;60:724-726. doi:10.3171/jns.1984.60.4.0724.
- Pons VG, Denlinger SL, Guglielmo BJ, Octavio J, Flaherty J, Derish PA, et al. Ceftizoxime versus vancomycin and gentamicin in neurosurgical prophylaxis: a randomized, prospective, blinded clinical study. Neurosurgery. 1993;33:416–422; discussion 422-423.
- Lübbeke A, Zingg M, Vu D, Miozzari HH, Christofilopoulos P, Uçkay I, et al. Body mass and weight thresholds for increased prosthetic joint infection rates after primary total joint arthroplasty. Acta Orthop. 2016;87:132–138. doi: 10.3109/17453674.2015.1126157.
- Zingg M, Miozzari HH, Fritschy D, Hoffmeyer P, Lübbeke A. Influence of body mass index on revision rates after primary total knee arthroplasty. Int Orthop. 2016;40:723-729. doi:10.1007/s00264-015-3031-0.

Author: Dolors Rodriguez-Pardo

QUESTION 6: What are the optimal prophylactic antibiotics for patients with neurogenic bladder who are undergoing spine surgery?

RECOMMENDATION: The recommended standard perioperative antibiotic prophylaxis in spine surgery is cefazolin, but a broader-spectrum prophylaxis may be necessary in patient subpopulations more prone to acquiring surgical site infections (SSIs). In the case of neurogenic bladder, preoperative urine culture and individualized antibiotic prophylaxis are associated with a significant decrease in SSIs due to gram-negative bacilli

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 79%, Disagree: 14%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Prevention of SSI is of utmost importance in patients undergoing spine surgery, and perioperative antibiotic prophylaxis is a key measure to avoid this complication [1,2]. However, the superiority of one agent or schedule over any other has not been clearly demonstrated [1,2]. The recommended standard perioperative antibiotic prophylaxis in spinal surgery is cefazolin [1]. Isolated reports have shown that a broader-spectrum prophylaxis may be necessary in patient subpopulations more prone to acquiring poly-microbial SSI, such as those with neuromuscular deformities or spinal cord injury. In a retrospective observation study, Dessy et al. demonstrated that an enhanced antibiotic prophylaxis using intravenous (IV) cefuroxime for 24 hours plus vancomycin until drain removal in instrumented spinal surgery, and IV cefuroxime for 24 hours in noninstrumentation cases reduced the rate of SSIs in spine surgery [3].

There are no published data regarding the best antibiotic treatment to be used as prophylaxis in patients with neurogenic bladder. The North American Spine Society (NASS) evidence-based guidelines on antibiotic prophylaxis in spinal surgery have pointed out that potential subgroups of patients requiring effective prophylaxis against GNB may exist, although they have not been clearly defined [1]. In the case of patients with neurogenic bladder, they are more prone to urinary tract colorization and infection [4-5]. Although

asymptomatic bacteriuria (AB) should not be routinely treated in these patients because of rising resistance patterns, in the case of symptomatic urinary tract infection (UTI) antibiotic treatment should be administered and antibiotic selection should be based on local and patient-based resistance patterns so that the spectrum can be as narrow as possible [5]. In this line, recent Clinical Guidelines for the Diagnosis and Treatment of UTI of the Spanish Society of Infectious Diseases state that screening for, and treatment of, AB prior to performing instrumental spinal surgery is recommended for patients with neurogenic bladders or urinary incontinence in order to reduce the risk of gram-negative SSIs [6].

It was reported that up to 61% of children with myelomeningocele have neurogenic bladders [7-9]. Hatlen et al. demonstrated that the presence of positive urinary cultures before elective spine surgery for children with myelomeningocele leads to an increased risk of perioperative spine infections [10]. Olsen et al. conducted a case-control study to determine independent risk factors for SSI following orthopaedic spinal operations [11]. Among the patientlevel factors in the univariate analysis, any incontinence (bowel or bladder, or both and preoperative or postoperative) significantly increased the risk of SSIs.

Although gram-positive organisms (particularly Staphylococcus aureus) predominate as causative agents for SSIs in patients undergoing spine surgery, GNB accounted for a sizeable portion of SSIs, particularly among lower lumbar and sacral spine surgical procedures [2]. Patients with incontinence, neurogenic bladder or indwelling catheters are more prone to urinary tract colonization and infection and may therefore be at higher risk of SSIs by GNB [4]. Contamination by GNB should not occur during the operative procedure, as these microorganisms are not usually present among the patient's skin flora [12]. Previous studies have suggested that GNB contamination could be secondary to hematogenous seeding originating in the urinary tract or to local skin contamination in incontinent patients, especially those undergoing surgery at the lumbosa-

Nuñez-Pereira et al. hypothesized that detecting urinary tract colonization preoperatively and adjusting antibiotic prophylaxis according to urine culture results might lower the overall SSI rate by reducing the number of GNB infections [12]. They performed a retrospective cohort study comparing two consecutive groups of patients undergoing posterior spinal fusion and instrumentation at a single institution. Cohort A included 236 patients, operated on between January 2006 and March 2007, receiving standard preoperative antibiotic prophylaxis with cefazolin (clindamycin in allergic patients). Cohort B included 223 patients operated on between January and

December 2009, receiving individualized antibiotic prophylaxis and treatment based on preoperative urine culture. The study demonstrated that preoperative urine culture and individualized antibiotic prophylaxis are associated with a significant decrease in SSI due to GNB in high-risk patients undergoing spinal surgery.

Measures aimed at preventing UTI in patients with neurogenic bladder such as closed catheter drainage in patients with an indwelling catheter and the use of clean intermittent catheterization could reduce the risk of perioperative spine infections [4]. Intravesical Botox, bacterial interference and sacral neuromodulation show significant promise for the prevention of UTIs in neurogenic bladder patients [5].

REFERENCES

- Watters WC, Baisden J, Bono CM, Heggeness MH, Resnick DK, Shaffer WO, et al. Antibiotic prophylaxis in spine surgery: an evidence-based clinical guideline for the use of prophylactic antibiotics in spine surgery. Spine J. 2009;9:142–146. doi:10.1016/j.spinee.2008.05.008.
- Shaffer WO, Baisden JL, Fernand R, Matz PG, North American Spine Society. An evidence-based clinical guideline for antibiotic prophylaxis in spine
- surgery. Spine J. 2013;13:1387–1392. doi:10.1016/Jj.spinee.2013.06.030. Dessy AM, Yuk FJ, Maniya AY, Connolly JG, Nathanson JT, Rasouli JJ, et al. Reduced surgical site infection rates following spine surgery using an enhanced prophylaxis protocol. Cureus. 2017;9:e1139. doi:10.7759 cureus.1139
- Pigrau C, Rodríguez-Pardo MD. [Infections associated with the use of indwelling urinary catheters. Infections related to intrauterine devices]. Enferm Infecc Microbiol Clin. 2008;26:299-310.
- Matsumoto T, Takahashi K, Manabe N, Iwatsubo E, Kawakami Y. Urinary tract infection in neurogenic bladder. Int J Antimicrob Agents. 2001;17:293-297. doi:10.3978/j.issn.2223-4683.2016.01.06. de Cueto M, Aliaga L, Alós JI, Canut A, Los-Arcos I, Martínez JA, et al. Execu-
- tive summary of the diagnosis and treatment of urinary tract infection: guidelines of the Spanish Society of Clinical Microbiology and Infectious Diseases (SEIMC). Enferm Infecc Microbiol Clin. 2017;35:314-320. doi:10.1016/j.eimc.2016.11.005.
- Elliott SP, Villar R, Duncan B. Bacteriuria management and urological evaluation of patients with spina bifida and neurogenic bladder: a multicenter
- Survey. J Urol. 2005;173:217–220. doi:10.1097/01.ju.0000146551.87110.fd.
 Zickler CF, Richardson V. Achieving continence in children with neurogenic bowel and bladder. J Pediatr Health Care. 2004;18:276–283. doi:10.1016/S0891524504001232 50891524504001233
- Schlager TA, Clark M, Anderson S. Effect of a single-use sterile catheter for each void on the frequency of bacteriuria in children with neurogenic bladder on intermittent catheterization for bladder emptying. Pediatrics. 2001;108:E71
- Hatlen T, Song K, Shurtleff D, Duguay S. Contributory factors to postop erative spinal fusion complications for children with myelomeningocele. Spine. 2010;35:1294–1299. doi:10.1097/BRS.ob013e3181bf8efe
- Olsen MA, Nepple JJ, Riew KD, Lenke LG, Bridwell KH, Mayfield J, et al. Risk factors for surgical site infection following orthopaedic spinal operations. J Bone Joint Surg Am. 2008;90:62–69. doi:10.2106/JBJS.F.01515. Núñez-Pereira S, Pellisé F, Rodríguez-Pardo D, Pigrau C, Sánchez JM, Bagó J,
- et al. Individualized antibiotic prophylaxis reduces surgical site infections by gram-negative bacteria in instrumented spinal surgery. Eur Spine J. 2011;20 Suppl 3:397-402. doi:10.1007/s00586-011-1906-3.

1.3. PREVENTION: BONE GRAFT

Author: Dolors Rodriguez-Pardo

QUESTION 1: Does the use of allograft increase the risk of spinal infection?

RECOMMENDATION: The use of allograft seems to increase the risk for infection in pediatric and neuromuscular scoliosis, however there is no increased risk in the adult degenerative population.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 77%, Disagree 0%, Abstain: 23% (Super Majority, Strong Consensus)

Spine

RATIONALE

It has been postulated that infection risk from bone allograft may be caused by contamination or by the overwhelming of local host defenses [1,2]. Much of the data addressing this issue can be found in the pediatric literature. In a case-control study of 22 pediatric patients with infections after spine surgery, Croft et al. found that allograft use was strongly associated with surgical site infection (odds ratio (OR) = 10.7, p < 0.0001) [3]. Aleissa et al. showed similar results in 14 patients with SSI (risk rate (RR) 9.6, p < 0.001) [4]. Sponseller et al. were able to demonstrate a statistically significant increase in infection risk with the use of allograft versus autograft (p = 0.010)[5].

Several systematic reviews have also addressed this subject. Fei et al. performed a meta-analysis of risk factors for surgical site infection after spine surgery in 12 high-quality studies [6]. They found a relative risk for infection of 2.72% with the use of bone allograft, though there was a broad confidence interval and they failed to reach statistical significance at p = 0.244. Meng et al. [2] performed a systematic review of 13 studies of infection risk in pediatric spine surgery. The use of allograft carried an odds ratio of 8.498 with a high statistical significance at p < 0.001, though the authors cautioned about possible bias due to study heterogeneity. Glotzbecker et al. found grade C evidence of an association between allograft use and surgical site infection [7].

On the other hand, multiple studies have demonstrated that even in the pediatric literature, there is conflicting evidence. Knapp et al. studied patients with Adolescent Idiopathic Scoliosis (AIS) and found that allograft did not increase the risk for infection [8]. In a case-control study of pediatric patients undergoing spinal fusion, Shen et al. also found that there was no increased risk with allograft [9]. In the adult population, several large studies have failed to find an association between allograft use and infection. Mark et al. looked at over 1,400 patients who underwent spinal fusion, and there was no difference in infection rate when using allograft or autograft [10]. Similarly, Saeedinia et al. looked at almost 1,000 patients undergoing spinal surgery and failed to find an association between allograft and infection [11].

REFERENCES

- Hassanzadeh H, Jain A, Kebaish KM, et al. Prevalence of allograft contamination during intraoperative processing for spinal deformity correction surgery, Spine Deform. 2013;1(5):348–351.

 Meng F, Cao J, Meng X. Risk factors for surgical site infection following
- pediatric spinal deformity surgery: a systematic review and meta-analysis.
- Child's Nerv Syst. 2015;31(4):521-527. Croft LD, Pottinger JM, Chiang H-Y, Ziebold CS, Weinstein SL, Herwaldt LA.
- Risk factors for surgical site infections after pediatric spine operations. Spine (Phila Pa 1976). 2015;40(2):E112–E119.

 Aleissa S, Parsons D, Grant J, Harder J, Howard J. Deep wound infection following pediatric scoliosis surgery: incidence and analysis of risk factors.
- Can J Surg. 2011;54(4):263–269. Sponseller PD, LaPorte DM, Hungerford MW, Eck K, Bridwell KH, Lenke LG. Deep wound infections after neuromuscular scoliosis surgery: a multi-center study of risk factors and treatment outcomes. Spine (Phila Pa 1976).
- 2000;25(19):2461-2466. Fei Q, Li J, Lin J, et al. Risk factors for surgical site infection after spinal
- Surgery: a meta-analysis. World Neurosurg. 2016;95:507–515. Glotzbecker MP, Riedel MD, Vitale MG, et al. What's the evidence? Systematic literature review of risk factors and preventive strategies for surgical site infection following pediatric spine surgery. J Pediatr Orthop.
- 2013;33(5):479–487. Knapp DR Jr, Jones ET, Blanco JS, et al. Allograft bone in spinal fusion for adolescent idiopathic scoliosis. J Spinal Disord Tech. 2005;18(Suppl):S73–
- Shen J, Liang J, Yu H. Risk factors for delayed infections after spinal fusion and instrumentation in patients with scoliosis. I Neurosurg Spine.
- Mark M. Mikhael, MD, et al. Postoperative culture positive surgical site infections after the use of irradiated allograft, nonirradiated allograft, or autograft for spinal fusion. Spine. 2009;34(22):2466-2488.
- Saeedinia S, Nouri M, Azarhomayoun A, et al. The incidence and risk factors for surgical site infection after clean spinal operations: a prospective cohort study and review of the literature. Surg Neurol Int. 2015;6:154.

Authors: Steven Schmitt, Christopher Kepler, Wesley Bronson

QUESTION 2: Can allograft, synthetic bone substitute or autograft be used during revision spinal surgery in patients with prior spine infection?

RECOMMENDATION: Based on available data, it appears that allograft, autograft and synthetic cages may be used successfully along with posterior screw fixation and prolonged antibiotic therapy in the treatment of pyogenic spondylodiscitis. This data can probably be extrapolated to also confirm that allograft and autograft safe during revision spinal surgery with prior infection.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

There are several small studies suggesting that bone allograft and autograft may be used successfully with posterior screw fixation and antibiotics to treat spine infections. Dobran et al. reviewed 18 patients who underwent posterior screw fixation along with allograft and autograft for pyogenic spondylodiscitis [1]. All patients had successful fusion and normalization of C-reactive protein at a mean follow-up of 30 months. Likewise, Chung et al. reported a study of 20 patients who underwent anterior fibular allograft and posterior screw fixation for spondylodiscitis [2]. All patients had significant improvement in pain and satisfaction scores, with at least 36 months of follow-up. Only two patients had superficial wound complications. In a third study, An et al. reviewed 15 patients who underwent mixed allograft and autograft with posterior screw fusion [3]. All but one showed significant improvement in neurological deficit, functional outcome and pain, with a mean follow-up of 27 months.

Synthetic materials have also been used in the successful treatment of pyogenic spondylodiscitis. Shiban et al. reported 52 patients treated with polyetheretherketone (PEEK) cages in combination with posterior pedicle screw fixation [4]. Patients received two weeks of intravenous and three months of oral antibiotic therapy. Infection was cured in all and 16 of the 28 with some neurologic deficit improved at 12 months of follow-up. Similar results were reported with PEEK cages and posterior fixation by Schomacher et al. (51 patients, 20 months of follow-up) and Brase et al. (nine patients,

mean follow-up 13 months) [5,6]. One study compared three different types of cages (titanium mesh, titanium and PEEK) versus autologous iliac bone strut [7]. All received posterior screw fixation. There were no significant differences in clinical or radiographic outcomes, and infections were judged cured in all at a mean of 36 months for follow-up. Multiple other studies report similar findings [8–10].

REFERENCES

- Dobran M, Iacoangeli M, Nasi D, et al. Posterior titanium screw fixation without debridement of infected tissue for the treatment of thoracolumbar spontaneous pyogenic spondylodiscitis. Asian Spine J. 2016;10(3):465. doi:10.4184/asi.2016.10.3.465.
- doi:10.4184/lasj.2016.10.3.4465.

 [2] Chung TC, Yang SC, Chen HS, Kao YH, Tu YK, Chen WJ. Single-stage anterior debridement and fibular allograft implantation followed by posterior instrumentation for complicated infectious spondylitis. Medicine (Baltimore). 2014;93(27):e190. doi:10.1097/MD.000000000000190.

 [3] An KC, Kim JY, Kim TH, et al. Posterior lumbar interbody fusion using
- [3] An KC, Kim JY, Kim TH, et al. Posterior lumbar interbody fusion using compressive bone graft with allograft and autograft in the pyogenic discitis. Asian Spine J. 2012;6(1):15. doi:10.4184/asj.2012.6.1.15.

- [4] Shiban E, Janssen I, da Cunha PR, et al. Safety and efficacy of polyetheretherketone (PEEK) cages in combination with posterior pedicel screw fixation in pyogenic spinal infection. Acta Neurochir (Wien). 2016;158(10):1851-1857. doi:10.1007/s00701-016-2924-z.
- [5] Schomacher M, Finger T, Koeppen D, et al. Application of titanium and polyetheretherketone cages in the treatment of pyogenic spondylodiscitis. Clin Neurol Neurosurg. 2014;127:65–70. doi:10.1016/j.clineuro.2014.09.027.
- [6] Brase A, Ringel F, Stüer C, Meyer B, Stoffel M. Debridement and fusion with polyetheretherketone implants in purulent spondylodiscitis: a clinical experience with nine patients. Acta Neurochir (Wien). 2010;152(11):2001–2004. doi:10.1007/s00701-010-0798-z.
- [7] Pee YH, Park JD, Choi YG, Lee SH. Anterior debridement and fusion followed by posterior pedicle screw fixation in pyogenic spondylodiscitis: autologous iliac bone strut versus cage. J Neurosurg Spine. 2008;8(5):405–412. doi:10.3171/SPI/2008/8/5/405.
- [8] Kim HW, Ryu JI, Bak KH. The safety and efficacy of cadaveric allografts and titanium cage as a fusion substitutes in pyogenic osteomyelitis. J Korean Neurosurg. 2011;50:348–356.
- Neurosurg. 2011;50:348–356.
 [9] Kim SS, Kang DH, Park H, et al. Surgical treatment of pyogenic spondylitis with the use of freeze-dried structural allograft. Korean J Spine. 2014;11(3):136–144.
 [10] Schuster JM, Avellino AM, Mann FA, et al. Use of structural allografts in
- [10] Schuster JM, Avellino AM, Mann FA, et al. Use of structural allografts in spinal osteomyelitis: a review of 47 cases. J Neurosurg. 2000;93(1 Suppl):8–14.

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1.4. PREVENTION: RISK FACTORS

Authors: Koji Yamada, Yoshihirio Uchida

QUESTION 1: Does prior or active tuberculosis (TB) preclude patients from undergoing spine surgery?

RECOMMENDATION: Prior or active TB does not preclude patients from undergoing spine surgery.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

The mainstay of treating spinal TB is chemotherapy [1]. Almost all antituberculous drugs penetrate well into tuberculous lesions [2], more than the desired minimum inhibitory concentrations (MIC) [3,4]. Abscesses usually resolve with medical therapy, as antituberculosis drugs penetrate very well [5,6].

There is controversy in the literature about the necessity of using surgical intervention in addition to spinal TB treatments. A Cochrane Database Review assessing the role of routine surgery in addition to chemotherapy in spinal TB including the studies from Medical Research Council (MRC) of the United Kingdom failed to reveal any statistically significant differences in various outcomes for additional surgery including: kyphosis angle, neurological deficit (none went on to develop this), bony fusion, absence of spinal TB, death from any cause, activity level regained, change of allocated treatment or bone loss [1]. Myelopathy with or without functional impairment most often responds to chemotherapy [7]. In two MRC studies conducted in Korea, more than 80% of patients had complete resolution of myelopathy or complete functional recovery when treated medically [8,9].

Though the review of the above trials was insufficient to say routine surgery early on was beneficial, several limitations exist [1]. First, two sets of trials reviewed in the literature were performed during the 1960s and 1970s, while in recent years new medications and better operative techniques have been developed. Second, the patients included in the MRC study were limited to two-vertebra disease with or without mild neural deficit [10,11]. The results

for patients with moderate to severe motor weakness were not addressed. Moreover, the patients seen in developing countries often have a large number of vertebrae involved, accompanied with a greater chance of kyphosis progression [12] and late onset paraplegia [13,14]. Third, late onset paraplegia usually become present more than 15 years after initial spinal infection [15–17]. In MRC studies, increased progression of kyphosis was seen in the conservatively-treated group with a lower fusion rate during their follow-up period [18]. Considering the difficulties in treating severe late symptomatic post TB kyphosis, the follow-up period in these studies could be insufficient to detect the magnitude of late complications. Fourth, it is generally known that some patients do not respond well to conservative treatment and are considered nonresponders [19]. For these patients, surgery should be considered to procure adequate tissue to ascertain the diagnosis as well as to reduce the disease load.

Potential benefits of surgery include less kyphosis, immediate relief of compressed neural tissue, quicker relief of pain, a higher percentage of bony fusion, quicker bony fusion, less chance of relapse, earlier return to previous activities and less bone loss [1,2]. Early surgical intervention for prevention of deformity is relatively simple and may prevent late neurological problems due to kyphosis of the spine [15,20,21]. From a review of 124 articles, 17.1% of the procedures were performed with defined indications including: etiology, neurological deficit (severe or progressive), spinal instability with or without kyphosis (severe or progressive), multisegmental disease and paraplegia of greater than three months [19]. Surgical interven-

tion for those without neurological recovery/improvement after chemotherapy for moderate motor weakness and surgical decompression of the cord under the cover of multi-drug chemotherapy for severe motor weakness irrespective to the duration of illness or cause, are also recommended [22].

Medical treatment is generally effective for those with or without mild neural deficit. Surgical intervention may be indicated in advanced cases with marked bony involvement, abscess formation or paraplegia, regardless of prior or active tuberculosis.

REFERENCES

- Jutte PC, van Loenhout-Rooyackers JH. Routine surgery in addition to chemotherapy for treating spinal tuberculosis. Cochrane Database Syst Rev. 2006 Jan 25;(5):CD004532. doi: 10.1002/14651858.CD004532.pub2. Review.
- Garg RK, Somvanshi DS. Spinal tuberculosis: a review. J Spinal Cord Med.
- 2011;34(5):440-454. doi: 10.1179/2045772311Y.00000000023. Tuli SM, Kumar K, Sen PC (1977) Penetration of antitubercular drugs in clinical osteoarticular tubercular lesions. Acta Orthop Scand. 48(4):362-368.
- Kumar K (1992) The penetration of drugs into the lesions of spinal tuberculosis. Int Orthop (SICOT). 16:67–68.
 Bakhsh A. Medical management of spinal tuberculosis: an experience from
- Pakistan. Spine (Phila Pa 1976). 2010;35(16): E787–E791.
- Prasad R. Management of multi-drug resistant tuberculosis: practitioners
- view. Indian J Tuberc. 2007;54(1):3-11.
 Fourteenth report of the Medical Research Council Working Party on Tuberculosis of the Spine. Five-year assessment of controlled trials of short-course chemotherapy regimens of 6, 9 or 18 months' duration for spinal tuberculosis in patients ambulatory from the start or undergoing radical surgery. Int Orthop. 1999;23(2):73–81.
 Twelfth report of the Medical Research Council Working Party on Tuber-
- culosis of the Spine. Controlled trial of short-course regimens of chemotherapy in the ambulatory treatment of spinal tuberculosis: results at three years of a study in Korea. J Bone Joint Surg Br. 1993;75(2):240–248.

- Pattison PRM. Pott's paraplegia: an account of the treatment of 89 consecutive patients. Paraplegia. 1986;24(2):77-91.
- Upadhyay SS, Saji MJ, Yau AC. Duration of antituberculosis chemotherapy in conjunction with radical surgery in the management of spinal tubercu-
- losis. Spine. 1996;21:1898–1993.
 Upadhyay SS, Sell P, Saji MJ, Sell B, Yau AC, Leong JC. 17-year prospective study of surgical management of spinal tuberculosis in children: Hong Kong operation compared with debridement surgery for short and longterm outcome of deformity. Spine. 1993;18:1704–1711.
- Rajasekaran S, Shanmugasundaram K. Prediction of the angle of gibbus deformity in tuberculosis of the spine. J Bone Joint Surg Am. 1987;69:503-
- Sundararaj GD, Behera S, Ravi V, Venkatesh K, Cherian VM, Lee V. Role of posterior stabilization in the management of tuberculosis of the dorsal and lumbar spine. J Bone Joint Surg Br. 2003;85:100–106.
- Tuli SM. Severe kyphotic deformity in tuberculosis of the spine. Int Orthop.
- 1995;19:327–331. Cheung WY, Luk KD. Clinical and radiological outcomes after conservative treatment of TB spondylitis: is the 15 years' follow-up in the MRC study long enough? Eur Spine J. 2013 Jun;22 Suppl 4:594-602. doi: 10.1007/s00586-012-
- 2332-x. Epub 2012 May 8. Luk KD. Tuberculosis of the spine in the new millennium. Eur Spine J. 8:338–
- 345. Moon MS, Moon JL, Moon YW, Kim SS, Kim SS, Sun DH, Choi WT. Pott's paraplegia in patients with severely deformed dorsal or dorsolumbar spines: treatment and prognosis. Spinal Cord. 41:164–171
- Medical Research Council Working Party on Tuberculosis of the Spine. A 15-year assessment of controlled trials of the management of tuberculosis
- of the spine in Korea and Hong Kong. J Bone Joint Surg Br. 80:456–462. Jain AK, Dhammi IK. Tuberculosis of the spine: a review. Clin Orthop Relat
- Res. 2007 Jul;460:39-49. Hsu LC, Cheng CL, Leong JC. Pott's paraplegia of late onset. The cause of compression and the results of anterior decompression. J Bone Joint Surg.
- 1988;70-B(4):534-538. Leong JC. Tuberculosis of the spine. J Bone Joint Surg. 1993;75-B(2):173-174. Kumar K. Spinal tuberculosis, natural history of disease, classifications and principles of management with historical perspective. Eur J Orthop Surg Traumatol. 2016 Aug;26(6):551–558. doi: 10.1007/s00590-016-1811-x.

Author: Carles Pigrau

QUESTION 2: Should routine methicillin-resistant *Staphylococcus aureus* (MRSA) screening be in place prior to spine surgery?

RECOMMENDATION: Routine MRSA screening should not be performed prior to spine surgery. However, in hospitals with a high incidence of S. aureus spinal surgical site infection (SSI) and particularly high rates of MRSA infections, MRSA screening might be useful.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 86%, Disagree: 7%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

According to a recent review of 161 studies, the pooled average of SSI in spine surgery was 1.9% (range: 0.1 to 22.6%) [1]. Instrumented spinal fusion had the highest rate (3.8%), followed by spinal decompression (1.8%) and spinal fusion (1.6%). S. aureus contributed to almost 50% of spinal SSIs with a range of 0.02 to 10%. Among S. aureus spinal SSIs, the pooled rate of MRSA infections was 38% [1]. The 30-day mortality rate among patients with SSI was 1.06%, double that of those without SSI (0.5%), with mortality increasing with the complexity of spinal surgery or with the presence of underlying diseases [2]. Moreover, SSIs increased re-admission rates (from 20-100%), reoperation rates (with a pooled average of 67%) and doubled health-care costs [1].

Preoperative nasal carriage of S. aureus has been shown to be a risk factor for SSI, but rates have been variable between studies [3,4]. Nasal decolonization with the use of topical mupirocin is utilized in 90% of cases, however, the impact of using this strategy on the reduction of SSIs in orthopaedic surgery have reported conflicting results [5,6]. A recent meta-analysis of all published studies in cardiac and orthopaedic surgery suggested that decolonization was associated with a significant decrease in S. aureus SSIs when either the intervention was applied to all patients or only to those who were nasal carriers [7]. Another meta-analysis showed that an absolute reduction in SSIs of 1% may be cost-effective, however, universal decolonization may increase the risk of mupirocin resistance [8].

In a not-yet published retrospective study of 1,749 patients scheduled for elective instrumented neurosurgery, the MRSA colonization rate was 0.74%. After decontamination, all MRSA carriage was eliminated and none of the 13 MRSA carriers developed an SSI, while only 1 MRSA-negative case developed a MRSA SSI.

In a recent retrospective study of 4,973 consecutive spine patients who were given cefazolin as prophylactic antibiotic therapy rather than topical nasal antibiotics for decolonization, 49 (1.1%) were MRSA carriers, and 94 (2.1%) developed an SSI, 11 of which were caused by MRSA [9]. The SSI rates were similar in nasal carriers compared to non-MRSA carriers (3 of 49 vs. 91 or 4,433, p = 0.13) and nasal carriage was not a risk factor for spinal SSIs.

In conclusion, in patients undergoing spinal surgery, the low level of MRSA carriage and MRSA SSI are arguments against routine MRSA screening. In hospitals with a high incidence of *S. aureus* spinal SSI and high rates of MRSA infections, MRSA screening could be useful.

REFERENCES

- [1] Patel H, Khoury H, Girgenti D, Welner S, Yu H. Burden of surgical site infections associated with select spine operations and involvement of Staphylococcus aureus. Surg Infect (Larchmt). 2017;18:461–473. doi:10.1089/
- [2] Núñez-Pereira S, Pellisé F, Rodríguez-Pardo D, Pigrau C, Bagó J, Villanueva C, et al. Implant survival after deep infection of an instrumented spinal fusion. Bone Joint J. 2013;95-B:1121–1126. doi:10.1302/0301-620X.95B8.30784.
- fusion. Bone Joint J. 2013;95-B:1121–1126. doi:10.1302/0301-620X.95B8.30784.

 [3] Hacek DM, Robb WJ, Paule SM, Kudrna JC, Stamos VP, Peterson LR. Staphylococcus aureus nasal decolonization in joint replacement surgery reduces infection. Clin Orthop Relat Res. 2008;466:1349–1355. doi:10.1007/s11999-008-0210-y.

- [4] Kluytmans JA, Manders MJ, van Bommel E, Verbrugh H. Elimination of nasal carriage of Staphylococcus aureus in hemodialysis patients. Infect Control Hosp Epidemiol. 1996;17:793–797.
- [5] Kalmeijer MD, Coertjens H, van Nieuwland-Bollen PM, Bogaers-Hofman D, de Baere G a. J, Stuurman A, et al. Surgical site infections in orthopedic surgery: the effect of mupirocin nasal ointment in a double-blind, randomized, placebo-controlled study. Clin Infect Dis. 2002;35:353-358. doi:10.1086/341025.
- [6] Lee AS, Macedo-Vinas M, François P, Renzi G, Vernaz N, Schrenzel J, et al. Trends in mupirocin resistance in meticillin-resistant Staphylococcus aureus and mupirocin consumption at a tertiary care hospital. J Hosp Infect. 2011;77:360–362. doi:10.1016/j.jhin.2010.11.002.
- [7] Schweizer M, Perencevich E, McDanel J, Carson J, Formanek M, Hafner J, et al. Effectiveness of a bundled intervention of decolonization and prophylaxis to decrease gram positive surgical site infections after cardiac or orthopedic surgery: systematic review and meta-analysis. BMJ. 2013;346:f2743.
- [8] van Rijen MML, Bonten M, Wenzel RP, Kluytmans JAJW. Intranasal mupirocin for reduction of Staphylococcus aureus infections in surgical patients with nasal carriage: a systematic review. J Antimicrob Chemother. 2008;61:254–261. doi:10.1093/jac/dkm480.
- [9] Calfee DP. Editorial commentary: considering universal mupirocin decolonization as an option for preventing surgical site infections. Clin Infect Dis. 2016;62:637-639. doi:10.1093/cid/civ992.

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QUESTION 3: Is there a role for routine decolonization of patients undergoing spine surgery? If so, what is the optimal agent(s)?

RECOMMENDATION: There is evidence to support the use of institutionalized screening and decolonization programs in methicillin-resistant *Staphylococcus aureus* (MRSA) carriers to reduce the rate of surgical site infection (SSI), however the optimum agents for decolonization have not been determined.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 91%, Disagree: 0%, Abstain: 9% (Super Majority, Strong Consensus)

RATIONALE

There is evidence to support the use of institutionalized screening and decolonization programs to reduce the rate of SSI, however the optimum agents for decolonization have not been determined [1]. Preoperative nasal MRSA colonization is associated with increased risk postoperative spinal SSI. Thakkar et al. reported screening positive MRSA SSI rates of 12% compared with screening positive for MSSA (5.73%) and screening negative (1.82%) [2]. Furthermore, Ramos et al. found increased rates of SSI in hip and knee arthroplasty and spine fusions, reporting a 4.35% SSI rate in colonized (nasal MRSA and MSSA) patients versus a 2.39% rate in noncolonized patients [3].

While widely utilized preoperatively, there is minimal evidence specifically supporting the use of chlorhexidine gluconate (CHG) showers preoperatively. The 2015 Cochrane review written by Webster et al. reported minimal evidence supporting isolated use of CHG showers preoperatively. Four reviewed trials comparing CHG to placebo found no effect, and only one trial comparing CHG showers to controls reported an improvement in SSI rate [4].

The majority of reviewed literature bundles the use of nasal decolonization with other interventions (CHG wipes, CHG showers, etc.). Multiple reviews on the effectiveness of bundled interventions for decolonization in surgical patients (including orthopaedic surgery) report reduced SSI rates with nasal decolonization and CHG wipes [5,6]. Reported studies on nasal decolonization protocols have largely shown benefit in reducing SSIs. Mullen et al. used CHG wipes and alcohol-based nasal decolonization preoperatively and reported a mean reduction rate in SSI of 81% (1.76 per 100 to 0.33 per 100) [7].

Chen et al. reviewed 19 studies of decolonization protocols on orthopaedic procedures and found significant efficacy in reducing

SSIs, reporting reduction of *S. aureus* SSIs ranging from 40–200% and reduction of MRSA SSI from 29–149% [8]. Bode et al. performed a randomized, double blinded trial to determine if decolonization would reduce the SSI rate. Of 6,771 general, orthopaedic and neurologic surgery patients, 18.5% tested positive for *Staphylococcus* and were decolonized with 5 days of CHG showers and mupirocin nasal ointment. SSI rates significantly reduced from 7.7 to 3.4% using eradication compared with the placebo control [9]. These interventions are likely cost-effective as well, as Slover determined that the cost-efficacy threshold for their institution's screening and decolonization protocol would be met with a spine SSI reduction of only 10% [10].

It is our recommendation that patients who screen positive for nasal MSSA and MRSA should be decolonized using 2% mupirocin ointment applied intranasally and 2% chlorhexidine gluconate (CHG) showers for five days preoperatively. Additionally, in patients positive for MRSA, intravenous vancomycin 15 mg/kg should be administered preoperatively prior to skin incision and for 24 hours postoperatively.

- [1] Savage JW, Anderson PA. An update on modifiable factors to reduce the risk of surgical site infections. Spine J. 2013;13:1017–1029. doi:10.1016/j. spinee.2013.03.051.
- [2] Thakkar V, Ghobrial GM, Maulucci CM, Singhal S, Prasad SK, Harrop JS, et al. Nasal MRSA colonization: impact on surgical site infection following spine surgery. Clin Neurol Neurosurg. 2014;125:94-97. doi:10.1016/j. clineuro.2014.07.018.
- [3] Ramos N, Stachel A, Phillips M, Vigdorchik J, Slover J, Bosco JA. Prior *Staphylococcus aureus* nasal colonization: a risk factor for surgical site infections following decolonization. J Am Acad Orthop Surg. 2016;24:880–885. doi:10.5435/JAAOS-D-16-00165.

- [4] Webster J, Osborne S. Preoperative bathing or showering with skin antiseptics to prevent surgical site infection. Cochrane Database Syst Rev. 2015;CD004985. doi:10.1002/14651858.CD004985.pub5.
- [5] Schweizer M, Perencevich E, McDanel J, Carson J, Formanek M, Hafner J, et al. Effectiveness of a bundled intervention of decolonization and prophylaxis to decrease gram positive surgical site infections after cardiac or orthopedic surgery: systematic review and meta-analysis. BMJ. 2013;346:f2743.
- [6] George S, Leasure AR, Horstmanshof D. Effectiveness of decolonization with chlorhexidine and mupirocin in reducing surgical site infections: a systematic review. Dimens Crit Care Nurs. 2016;35:204–222. doi:10.1097/DCC.000000000000192.
- [7] Mullen A, Wieland HJ, Wieser ES, Spannhake EW, Marinos RS. Perioperative participation of orthopedic patients and surgical staff in a nasal decolonization intervention to reduce Staphylococcus spp surgical site infections. Am J Infect Control. 2017;45:554–556. doi:10.1016/j.ajic.2016.12.021.
- [8] Chen AF, Wessel CB, Rao N. Staphylococcus aureus screening and decolonization in orthopaedic surgery and reduction of surgical site infections. Clin Orthop Relat Res. 2013;471:2383–2399. doi:10.1007/s11999-013-2875-0.
- Orthop Relat Res. 2013;471:2383–2399. doi:10.1007/s11999-013-2875-0.

 [9] Bode LGM, Kluytmans JAJW, Wertheim HFL, Bogaers D, Vandenbroucke-Grauls CMJE, Roosendaal R, et al. Preventing surgical-site infections in nasal carriers of Staphylococcus aureus. N Engl J Med. 2010;362:9–17. doi:10.1056/NEJM0a0808939.
- [10] Slover J, Haas JP, Quirno M, Phillips MS, Bosco JA. Cost-effectiveness of a Staphylococcus aureus screening and decolonization program for highrisk orthopedic patients. J Arthroplasty. 2011;26:360–365. doi:10.1016/j. arth.2010.03.009.



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QUESTION 4: How should patients currently using disease-modifying antirheumatic drugs (DMARDs) be managed in the perioperative period?

RECOMMENDATION: Spine surgeons caring for patients with rheumatic diseases must be aware that there are specific issues involved in their perioperative management. The optimal strategy for managing DMARD medications during the perioperative period of spine surgery is unknown due to the lack of evidence and it is largely based on low-quality evidence and expert opinion. A rheumatologist should be involved in the medication management around the time of surgery.

- 1. For nonbiologic DMARDs such as methotrexate (MTX), leflunomide, hydroxychloroquine and/or sulfasalazine, continuation of the current dose throughout the perioperative period is recommended.
- 2. For biologic DMARDs such as etanercept, we recommend that physicians withhold the biologic medication and plan elective surgery at the end of the dosing cycle for that specific medication. As an example, patients taking a weekly dose should schedule the surgery in the second week after the first withheld dose. These agents should not be restarted until external wound healing is complete, which is typically around two weeks. Exception: In patients taking tofacitinib (twice daily dose), withholding of tofacitinib for at least one week prior to surgery is recommended.
- 3. For medications typically used for systemic lupus erthematosus (SLE) patients, such as mycophenolate mofetil, azathioprine, cyclosporine and tacrolimus, the decision to withhold medications prior to surgery should be made on an individual basis.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Nonbiologic DMARDs

Although a reasonable concern exists about the potential of nonbiologic DMARDs to increase the risk of infection by affecting the immune response [1,2], stopping DMARDs prior to surgery may result in a flare-up of disease activity, which may adversely affect rehabilitation. Therefore, we suggest that patients continue the current dose of nonbiologic DMARDS throughout the perioperative period, including methotrexate (MTX), leflunomide, hydroxychloroquine and/or sulfasalazine. In clinical practice, the nonbiologic DMARD dose is often missed for one day and up to three days while the patient is hospitalized. Several studies of rheumatoid arthritis (RA) patients undergoing elective orthopaedic surgery have found that continued use of MTX through the perioperative period is safe [3,4]. A systematic review including four studies with RA patients undergoing elective orthopaedic surgery evaluated the effects of continuing MTX versus stopping MTX in the perioperative period [5]. Continued MTX therapy was safe perioperatively and was associated with a reduced risk of flares. There was no evidence to suggest that stopping MTX preoperatively reduced the incidence of infection or improved wound healing. However, in all of the studies, the mean dose of MTX was less than 15 mg per week.

The limited data on the use of leflunomide during the perioperative period is conflicting [6,7]. In one study, there were signifi-

cantly more wound complications in patients taking leflunomide at the time of elective orthopaedic surgery compared with patients on MTX [7].

There are also limited data suggesting it is safe to continue hydroxychloroquine and sulfasalazine in the perioperative period. In a retrospective study of 367 orthopaedic surgeries among 204 RA patients, two-thirds of whom were receiving nonbiologic DMARDs including hydroxychloroquine and sulfasalazine, there was no increased infection associated with nonbiologic DMARD use [8].

Biologic DMARDs

We recommend that surgeons withhold biologic medication and plan the elective surgery at the end of the dosing cycle for that specific medication. As an example, patients taking weekly etanercept should aim to schedule the surgery in the second week after the first withheld dose. Patients taking adalimumab in two-week intervals should plan the surgery in the third week after the first withheld dose. In a similar manner, patients on monthly intravenous abatacept should schedule the surgery in the fifth week after the first withheld dose. Patients taking rituximab should wait until month seven after the last dose to schedule the surgery, presumably when B cells have returned to the circulation. However, nonelective procedures should not be delayed in patients who have been recently treated.

There is relatively little evidence available regarding the optimal timing for use of biologic DMARDs in the perioperative period, and our recommendation is largely based on indirect evidence suggesting an increased risk of infection associated with their use [9–11]. Many [12–16], but not all [17,18] retrospective studies suggest that use of tumor necrosis factor (TNF) inhibitors do not increase the risk of postoperative infections or impair wound healing.

The infectious risks of abatacept are similar to those of TNF inhibitors and other biologic agents, but there are no trials that have examined abatacept's safety perioperatively [9,19]. A case series described eight uncomplicated surgeries in seven RA patients on abatacept [20]. Similarly, there is no direct evidence regarding the safety of the interleukin (IL)-1 receptor inhibitor anakinra in the perioperative period. Conclusions regarding perioperative safety are largely based on trials in nonoperative patients showing that the infection rate was similar to that in patients receiving placebo [21].

These agents should not be restarted until external wound healing is complete, which is typically around two weeks. There is no evidence regarding the optimal time to restart biologic DMARDs in the perioperative setting and this approach is based on standard precautions used for biologic agents that warn against use in patients with active infection, such as an open wound.

Antirheumatic Kinase Inhibitor

In patients taking tofacitinib, we (Fang et al.) withhold the medication for at least one week prior to surgery. Tofacitinib is an orally-administered Janus kinase (JAK) inhibitor that is used in the management of patients with moderately to severely active RA. Our recommendation is based on indirect evidence from systematic reviews and meta-analyses of tofacitinib in nonsurgical patients showing there is an increased risk of infection with tofacitinib compared with placebo. Although the half-life is thought to be short for tofacitinib, there is uncertainty regarding the duration of immunosuppression after the drug is held [22].

Other SLE-specific Medications

There is uncertainty regarding the optimal perioperative medication management in patients with SLE given the lack of data. More data are needed to help guide perioperative medication management in lupus patients, including information on hydroxychloroquine, MTX, mycophenolate mofetil, azathioprine, cyclosporine and tacrolimus. Given the clinical spectrum of SLE disease severity and organ involvement, the decision to withhold medications prior to surgery should be made on an individual basis. Thus, for patients with severe SLE and multi-organ involvement in which discontinuation of the medication may result in a disease flare, it is reasonable to continue the medications through the surgical period. This is based on indirect evidence from organ transplant patients that supports continuing anti-rejection therapy during the time of surgery [23,24].

REFERENCES

[1] Jain A, Maini R, Nanchahal J. Disease modifying treatment and elective surgery in rheumatoid arthritis: the need for more data. Ann Rheum Dis. 2004;63:602-603. doi:10.1136/ard.2003.017640.

[2] Salt E, Wiggins AT, Rayens MK, Morris BJ, Mannino D, Hoellein A, et al. Moderating effects of immunosuppressive medications and risk factors for post-operative joint infection following total joint arthroplasty in patients with rheumatoid arthritis or osteoarthritis. Semin Arthritis Rheum. 2017;46:423–429. doi:10.1016/j.semarthrit.2016.08.011.

- [3] Pieringer H, Stuby U, Biesenbach G. The place of methotrexate perioperatively in elective orthopedic surgeries in patients with rheumatoid arthritis. Clin Rheumatol. 2008;27:1217–1220. doi:10.1007/s10067-008-0888-y.
- [4] Grennan DM, Gray J, Loudon J, Fear S. Methotrexate and early postoperative complications in patients with rheumatoid arthritis undergoing elective orthopaedic surgery. Ann Rheum Dis. 2001;60:214–217.
- orthopaedic surgery. Ann Rheum Dis. 2001;60:214–217.

 [5] Loza E, Martinez-Lopez JA, Carmona L. A systematic review on the optimum management of the use of methotrexate in rheumatoid arthritis patients in the perioperative period to minimize perioperative morbidity and maintain disease control. Clin Exp Rheumatol. 2009;27:856–862.
- [6] Tanaka N, Sakahashi H, Sato E, Hirose K, Ishima T, Ishii S. Examination of the risk of continuous leflunomide treatment on the incidence of infectious complications after joint arthroplasty in patients with rheumatoid arthritis. J Clin Rheumatol. 2003;9:115–118. doi:10.1097/o1.RHU.000006231.54375.bd.
 [7] Fuerst M, Möhl H, Baumgärtel K, Rüther W. Leflunomide increases the
- [7] Fuerst M, Möhl H, Baumgärtel K, Rüther W. Leflunomide increases the risk of early healing complications in patients with rheumatoid arthritis undergoing elective orthopedic surgery. Rheumatol Int. 2006;26:1138–1142. doi:10.1007/S00296-006-0138-Z.
- [8] Escalante Ä, Beardmore TD. Risk factors for early wound complications after orthopedic surgery for rheumatoid arthritis. J Rheumatol. 1995;22:1844–1851.
- [9] Mushtaq S, Goodman SM, Scanzello CR. Perioperative management of biologic agents used in treatment of rheumatoid arthritis. Am J Ther. 2011;18:426–434. doi:10.1097/MJT.ob013e3181cb4042.
- [10] Bongartz T. Elective orthopedic surgery and perioperative DMARD management: many authors, fewer answers, and some opinions. J Rheumatol. 2007;34:653-655.
- 2007;34:653-655.
 [11] Goodman SM. Rheumatoid arthritis: perioperative management of biologics and DMARDs. Semin Arthritis Rheum. 2015;44:627-632. doi:10.1016/j.semarthrit.2015.01.008.
- doi:10.1016/j.semarthrit.2015.01.008.

 [12] Bibbo C, Goldberg JW. Infectious and healing complications after elective orthopaedic foot and ankle surgery during tumor necrosis factor-alpha inhibition therapy. Foot Ankle Int. 2004;25:331–335. doi:10.117/j107110070402500510.
- [13] Talwalkar SC, Grennan DM, Gray J, Johnson P, Hayton MJ. Tumour necrosis factor alpha antagonists and early postoperative complications in patients with inflammatory joint disease undergoing elective orthopaedic surgery. Ann Rheum Dis. 2005;64:650–651. doi:10.1136/ard.2004.028365.
- Wendling D, Balblanc JC, Brousse A, Lohse A, Lehuede G, Garbuio P, et al.
 Surgery in patients receiving anti-tumour necrosis factor alpha treatment in rheumatoid arthritis: an observational study on 50 surgical procedures.
 Ann Rheum Dis. 2005;64:1378–1379. doi:10.1136/ard.2005.037762.
- [15] den Broeder AA, Creemers MCW, Fransen J, de Jong E, de Rooij DJR, Wymenga A, et al. Risk factors for surgical site infections and other complications in elective surgery in patients with rheumatoid arthritis with special attention for anti-tumor necrosis factor: a large retrospective study. J Rheumatol. 2007;34:689–695.
- [16] George MD, Baker JF, Hsu JY, Wu Q, Xie F, Chen L, et al. Perioperative timing of infliximab and the risk of serious infection after elective hip and knee arthroplasty. Arthritis Care Res (Hoboken). 2017;69:1845–1854. doi:10.1002/ acr.23209.
- [17] Momohara S, Kawakami K, Iwamoto T, Yano K, Sakuma Y, Hiroshima R, et al. Prosthetic joint infection after total hip or knee arthroplasty in rheumatoid arthritis patients treated with nonbiologic and biologic disease-modifying antirheumatic drugs. Mod Rheumatol. 2011;21:469–475. doi:10.1007/s10165-011-0423-X.
- [18] Clay M, Mazouyes A, Gilson M, Gaudin P, Baillet A. Risk of postoperative infections and the discontinuation of TNF inhibitors in patients with rheumatoid arthritis: a meta-analysis. Joint Bone Spine. 2016;83:701–705. doi:10.1016/j.jbspin.2015.10.019.
- [19] Pham T, Bachelez H, Berthelot JM, Blacher J, Claudepierre P, Constantin A, et al. Abatacept therapy and safety management. Joint Bone Spine. 2012;79 Suppl 1:3-84. doi:10.1016/S1297-319X(12)70011-8.
 [20] Nishida K, Nasu Y, Hashizume K, Nakahara R, Ozawa M, Harada R, et al.
- matol. 2014;24:544–545. doi:10.3109/14397595.2013.874758.

 [21] Nuki G, Bresnihan B, Bear MB, McCabe D, European Group of Clinical Investigators. Long-term safety and maintenance of clinical improvement following treatment with anakinra (recombinant human interleukin-1 receptor antagonist) in patients with rheumatoid arthritis: extension phase of a randomized, double-blind, placebo-controlled trial. Arthritis Rheum. 2002;46:2838–2846. doi:10.1002/art.10578.
- [22] Strand V, Ahadieh S, French J, Geier J, Krishnaswami S, Menon S, et al. Systematic review and meta-analysis of serious infections with tofacitinib and biologic disease-modifying antirheumatic drug treatment in rheumatoid arthritis clinical trials. Arthritis Res Ther. 2015;17:362. doi:10.1186/s13075-015-0880-2.
- [23] Palmisano AC, Kuhn AW, Urquhart AG, Pour AE. Post-operative medical and surgical complications after primary total joint arthroplasty in solid organ transplant recipients: a case series. Int Orthop. 2017;41:13–19. doi:10.1007/s00264-016-3265-5.
- [24] Klement MR, Penrose CT, Bala A, Wellman SS, Bolognesi MP, Seyler TM. How do previous solid organ transplant recipients fare after primary total knee arthroplasty? J Arthroplasty. 2016;31:609–615.e1. doi:10.1016/j.arth.2015.10.007.

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Spine

QUESTION 5: Is postoperative hyperglycemia a risk factor for the development of infection following spinal surgery?

RECOMMENDATION: From the limited evidence, the association between postoperative hyperglycemia and surgical site infection (SSI) remains unclear and further study is needed.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

Postoperative hyperglycemia does not only occur in patients diagnosed with diabetes mellitus (DM). Only 41% of patients with serum glucose levels greater than 200 mg/dL were identified in the medical records with the diagnosis of diabetes [1]. Langlois et al. suggested that non-diabetic patients experienced a statistical increase in blood glucose levels in the first three days following spine surgery [2]. They also pointed out the possibility of blood glucose elevation in nondiabetic patients associated with postsurgical complications. After major surgery, perioperative blood glucose elevations may be associated with previously undiagnosed DM, or occur because of the activation of the hypothalamic-pituitary axis, a physical response to severe stress in individuals at risk [3].

DM is a disease of uncontrolled hyperglycemia, which impairs the immune system. The wound healing in patients with diabetes is impaired as a result of microangiopathic changes and ischemia, impaired granulocyte function and a lack of platelet-derived growth factor function in the wound [4]. Despite the lack of multiple randomized clinical trials, various retrospective studies have found that DM is strongly associated with SSI after spinal surgery [5-16]. Moreover, DM increases the risk of not only SSI but other postoperative complications such as urinary tract infection, unplanned readmission and prolonged length of stay [17-19].

From a retrospective case-control study of patients who underwent an orthopaedic spinal operation performed at a universityaffiliated tertiary care hospital, the risk of SSI, the odds ratio for postoperative hyperglycemia (> 200 mg/dL), was 2.9 (95% confidence interval (CI): 1.2, 6.5) after univariate analyses. But, the risk did not remain significant after multivariate logistic regression analysis [11]. A retrospective case-control study evaluating 104 patients with SSI after spinal surgery matched with 104 randomly-selected control patients without SSI after spinal surgery, revealed that patients with postoperative glucose measurements greater than 126 mg/dL within 48 hours after surgery were significantly more likely to develop an SSI than patients without an elevated glucose measurement on univariate analysis (crude odds ratio: 3.2, 95% CI: 1.6, 6.3). But, it was not significant after adjusting for other variables [20]. A retrospective case-control study evaluating specific independent risk factors for SSI after laminectomy or spinal fusion at a tertiary care hospital affiliated with a university hospital, identified that high serum glucose (> 200 mg/dL) at any time during hospitalization was significantly associated with SSI in the univariate analysis (odds ratio: 3.0,

On the other hand, a retrospective study evaluating perioperative variables to determine the risk factors for SSI in a total of 2,715 patients undergoing posterior lumbar spinal surgery revealed that high preoperative serum glucose (odds ratio: 1.169, 95% CI: 1.016, 1.345) and a history of DM (odds ratio: 2.227, 95% CI: 1.100, 4.506) were associated with SSI in multivariate logistic regression analysis, although postoperative serum glucose level showed no association [21]. In another retrospective study using the Nationwide Inpatient Sample (NIS) database, uncontrolled DM revealed a higher risk of postoperative infection (odds ratio: 4.90, 95% CI = 2.84, 8.46) than controlled DM (odds ratio: 1.91, 95% CI: 1.54, 2.37) [7]. But, there was no ICD-9-CM coding standard or parameter in the clinical setting that provides standardization of "uncontrolled" or "controlled" diabetic patients. Furthermore, the NIS does not provide quantitative data on blood glucose levels or hemoglobin A1c (HbA1c) percentage, making it impossible to further stratify cohorts based on overall control of a patient's diabetic condition.

Limited evidence supports the association between perioperative HbA1c and SSI [22,23]. The cut-off values for HbA1c differ among studies and the results were originated from small retrospective studies without multivariate analyses. Larger prospective studies are needed to confirm the association.

Though DM is strongly related to SSI in spinal surgery, no observational studies were able to reveal a significant association between postoperative hyperglycemia and SSI in multivariate analyses. From the limited evidence, the association between postoperative hyperglycemia and SSI remains unclear, and further study is needed on this issue.

- Olsen MA, Mayfield J, Lauryssen C, Polish LB, Jones M, Vest J, et al. Risk factors
- for surgical site infection in spinal surgery. J Neurosurg. 2003;98:149–155. Langlois J, Bouyer B, Larroque B, Dauzac C, Guigui P. Glycemic instability of non-diabetic patients after spine surgery: a prospective cohort study. Eur Spine J. 2014;23:2455–2461. doi:10.1007/S00586-014-3489-2. Kiran RP, Turina M, Hammel J, Fazio V. The clinical significance of an elevated
- postoperative glucose value in nondiabetic patients after colorectal surgery: evidence for the need for tight glucose control? Ann Surg. 2013;258:599–604; discussion 604-605. doi:10.1097/SLA.0b013e3182a501e3.
- Satake K, Kanemura T, Matsumoto A, Yamaguchi H, Ishikawa Y. Predisposing factors for surgical site infection of spinal instrumentation surgery for diabetes patients. Eur Spine J. 2013;22:1854–1858. doi:10.1007/s00586-013-
- Golinvaux NS, Varthi AG, Bohl DD, Basques BA, Grauer JN. Complication rates following elective lumbar fusion in patients with diabetes: insulin dependence makes the difference. Spine. 2014;39:1809-1816. doi:10.1097/ BRS.000000000000506
- Guzman JZ, Skovrlj B, Shin J, Hecht AC, Qureshi SA, Iatridis JC, et al. The impact of diabetes mellitus on patients undergoing degenerative cervical spine surgery. Spine. 2014;39:1656–1665. doi:10.1097/BRS.000000000000498.
- Appaduray SP, Lo P. Effects of diabetes and smoking on lumbar spinal surgery
- outcomes. J Clin Neurosci. 2013;20:1713–1717. doi:10.1016/j.jocn.2013.01.021. Veeravagu A, Patil CG, Lad SP, Boakye M. Risk factors for postoperative spinal wound infections after spinal decompression and fusion surgeries. Spine. 2009;34:1869-1872. doi:10.1097/BRS.obo13e3181adc989.
- Demura S, Kawahara N, Murakami H, Nambu K, Kato S, Yoshioka K, et al. Surgical site infection in spinal metastasis: risk factors and countermea-
- sures. Spine. 2009;34:635–639. doi:10.1097/BRS.obo13e31819712ca. Chen S, Anderson MV, Cheng WK, Wongworawat MD. Diabetes associated with increased surgical site infections in spinal arthrodesis. Clin Orthop
- Relat Res. 2009;467:1670–1673. doi:10.1007/s11999-009-0740-y. Olsen MA, Nepple JJ, Riew KD, Lenke LG, Bridwell KH, Mayfield J, et al. Risk factors for surgical site infection following orthopaedic spinal operations. J Bone Joint Surg Am. 2008;90:62–69. doi:10.2106/JBJS.F.01515

- Friedman ND, Sexton DJ, Connelly SM, Kaye KS. Risk factors for surgical site infection complicating laminectomy. Infect Control Hosp Epidemiol. 2007;28:1060–1065. doi:10.1086/519864.
- Browne JA, Cook C, Pietrobon R, Bethel MA, Richardson WJ. Diabetes and early postoperative outcomes following lumbar fusion. Spine. 2007;32:2214–2219. doi:10.1097/BRS.0b013e31814b1bco. Fang A, Hu SS, Endres N, Bradford DS. Risk factors for infection after spinal
- surgery. Spine. 2005;30:1460–1465
- Kanafani ZA, Dakdouki GK, El-Dbouni O, Bawwab T, Kanj SS. Surgical site infections following spinal surgery at a tertiary care center in Lebanon: incidence, microbiology, and risk factors. Scand J Infect Dis. 2006;38:589-592. doi:10.1080/00365540600606440.
- Watanabe M, Sakai D, Matsuyama D, Yamamoto Y, Sato M, Mochida J. Risk factors for surgical site infection following spine surgery: efficacy of intraoperative saline irrigation. J Neurosurg Spine. 2010;12:540–546. doi:10.3171/2009.11.SPINE09308.
- Qin C, Kim JYS, Hsu WK. Impact of insulin dependence on lumbar surgery outcomes: an NSQIP analysis of 51,277 patients. Spine. 2016;41:E687–E693. doi:10.1097/BRS.0000000000001359.
- Glassman SD, Alegre G, Carreon L, Dimar JR, Johnson JR. Perioperative complications of lumbar instrumentation and fusion in patients with diabetes mellitus. Spine J. 2003;3:496-501.

- Cancienne JM, Werner BC, Hassanzadeh H, Singla A, Shen FH, Shimer AL. The association of perioperative glycemic control with deep postoperative infection after anterior cervical discectomy and fusion in patients with
- diabetes. World Neurosurg. 2017;102:13-17. doi:10.1016/j.wneu.2017.02.118. Maragakis LL, Cosgrove SE, Martinez EA, Tucker MG, Cohen DB, Perl TM. Intraoperative fraction of inspired oxygen is a modifiable risk factor for surgical site infection after spinal surgery. Anesthesiology. 2009;110:556–562. doi:10.1097/ALN.obo13e3181974be7.
- Liu JM, Deng HL, Chen XY, Zhou Y, Yang D, Duan MS, et al. Risk factors for surgical site infection after posterior lumbar spinal surgery. Spine. 2018;43:732-737. doi:10.1097/BRS.0000000000002419
- Hikata T, Iwanami A, Hosogane N, Watanabe K, Ishii K, Nakamura M, et al. High preoperative hemoglobin Aic is a risk factor for surgical site infection after posterior thoracic and lumbar spinal instrumentation surgery. J Orthop Sci. 2014;19:223–228. doi:10.1007/so07/6-013-0518-7. Arnold PM, Fehlings MG, Kopjar B, Yoon ST, Massicotte EM, Vaccaro AR, et
- al. Mild diabetes is not a contraindication for surgical decompression in cervical spondylotic myelopathy: results of the AOSpine North America multicenter prospective study (CSM). Spine J. 2014;14:65-72. doi:10.1016/j. spinee.2013.06.016.

Authors: Steven Schmitt, Christopher Kepler

QUESTION 6: Is there an association between urinary tract infection (UTI) and surgical site infection (SSI) following spinal surgery?

RECOMMENDATION: Evidence regarding an association between UTI and SSI following spine surgery is conflicting and no convincing relationship has been proven. In a like fashion, no convincing relationship has been established between asymptomatic bacteriuria and SSI following spine surgery.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 71%, Disagree: 21%, Abstain: 8% (Super Majority, Strong Consensus)

RATIONALE

The treatment of organisms isolated from urine culture in the setting of orthopaedic surgery with hardware implantation is controversial and has been often driven by anecdote. The risk of seeding of hip and knee arthroplasties from asymptomatic bacteriuria has been studied and found to be small, with no cases in two studies [1,2]. A systematic review of the topic concluded that there was no evidence to support a direct causal relationship between perioperative asymptomatic bacteriuria and arthroplasty infection [3].

Data from the American College of Surgeons National Surgical Quality Improvement Program suggests that UTIs occur in nearly 1 of 50 patients undergoing posterior lumbar fusion procedures [4]. However, there are few studies that directly address a relationship between UTI and SSI in instrumented spine surgery. Nunez-Pereira et al. studied 466 patients, of whom 89 had UTIs and 54 had SSIs, with 22 patients having both [5]. Of these 22, the same organism was isolated from the surgical site and urine in nine patients. UTI conferred an odds ratio (OR) of 3.1 for SSI, though the statistical analysis recognized all UTIs and not just infections with the same organism. Tominaga et al. studied a cohort of 825 patients with 14 patients who developed SSIs and 20 patients who developed UTIs, and found no association between SSI and UTI [6].

It seems germane as well to address the relationship of asymptomatic bacteriuria and postoperative spine infection. Lee et al. studied 355 women > 65 years of age undergoing spine surgery [7]. Of these, 42 developed asymptomatic bacteriuria, with no association with SSI. A statistically significant association was found between asymptomatic bacteriuria with a Foley catheter in place and infec-

tion in patients who had undergone instrumentation of multiple levels. However, of 15 patients with postoperative infections, only 2 had the same organism (Staphylococcus epidermidis in both cases) isolated from cultures of surgical site and urine.

- Cordero-Ampuero J, González-Fernández E, Martínez-Vélez D, Esteban J. Are antibiotics necessary in hip arthroplasty with asymptomatic bacteriuria? Seeding risk with/without treatment. Clin Orthop Relat Res. 2013;471:3822– 3829. dõi:10.1007/s[']11999-013-2868-z.
- Martínez-Vélez D, González-Fernández E, Esteban J, Cordero-Ampuero J. Prevalence of asymptomatic bacteriuria in knee arthroplasty patients and subsequent risk of prosthesis infection. Eur J Orthop Surg Ťraumatol. -214. doi:10.1007/s00590-015-1720-4.
- 2016;26:209-214. doi:10.1007/s00590-015-1720-4.

 Mayne AlW, Davies PSE, Simpson JM. Antibiotic treatment of asymptomatic bacteriuria prior to hip and knee arthroplasty; a systematic review of the literature. Surgeon. 2018;16:176-182. doi:10.1016/j.surge.2017.08.007.

 Bohl DD, Ahn J, Tabaraee E, Ahn J, Jain A, Grauer JN, et al. Urinary tract infection following posterior lumbar fusion procedures: an American College of Surgeons National Surgical Quality Improvement Program Study. Spine. :015;40:1785–1791. doi:10.1097/BRS.0000000000001003.
- Núñez-Pereira S, Rodríguez-Pardo D, Pellisé F, Pigrau C, Bagó J, Villanueva C, et al. Postoperative urinary tract infection and surgical site infection in instrumented spinal surgery: is there a link? Clin Microbiol Infect. 2014;20:768-773. doi:10.1111/1469-0691.12527.
 Tominaga H, Setoguchi T, Kawamura H, Kawamura I, Nagano S, Abematsu M,
- et al. Risk factors for unavoidable removal of instrumentation after surgical site infection of spine surgery: A retrospective case-control study. Medicine (Baltimore). 2016;95:e5118. doi:10.1097/MD.0000000000005118. Lee SE, Kim KT, Park YS, Kim YB. Association between asymptomatic urinary
- tract infection and postoperative spine infection in elderly women: a retro spective analysis study. J Korean Neurosurg Soc. 2010;47:265–270. doi:10.3340/ jkns.2010.47.4.265.

Authors: Alexander Montgomery, Daniel Tarazona

QUESTION 7: What are the risk factors predisposing a patient to surgical site infections (SSI) after spine surgery?

RECOMMENDATION: Numerous risk factors for SSIs following spine surgery have been identified, including diabetes, obesity, prior SSI, smoking, longer operative times, posterior approach to spine and the number of levels fused.

LEVEL OF EVIDENCE: Moderate

Spine

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

The relatively low incidence of postoperative SSIs after spine surgery makes it challenging for studies to evaluate the risk factors for SSI in a prospective manner [1]. Based on our literature search, a number of retrospective studies and a single prospective study were identified. The findings of prior studies have also been summarized by multiple systematic reviews. Pull ter Gunne et al. performed a systematic review of 24 studies that identified risk factors for SSI after spine surgery [2]. All 24 studies were case-control and case series. There was a total of 73 potential factors evaluated, 34 of which were found to be significant in at least 1 study. There were 11 risk factors that were found to be significant in at least 2 studies. Among all risk factors, diabetes, obesity and prior SSI were the only three that were confirmed as risk factors by a multitude of studies.

Similarly, there was another systematic review which analyzed 36 observational studies for which 46 independent factors were studied [3]. Only six risk factors had been consistently proven to show an association with SSI after spine surgery, including diabetes, obesity, longer operative time, smoking, history of SSI and type of surgical procedure (i.e. tumor resection).

More recently, a prospective multicenter surveillance study was performed which enrolled 2,736 patients who underwent posterior thoracic and/or lumbar spine surgery [4]. Of these patients, 24 (0.9%) developed postoperative deep SSI. Preoperative steroid therapy, spinal trauma, male gender and prolonged operating time (> three hours) were found to be independent risk factors for SSI after spine surgery. Several previous retrospective studies have not identified preoperative steroid use and male gender as risk factors for SSI after spine surgery [2,5,6].

An ongoing prospective study funded by Pfizer evaluating the potential role of vaccination against Staphylococcus is likely to provide valuable information regarding the most important risk factors for SSI after spine surgery.

REFERENCES

- Pawar AY, Biswas SK. Postoperative spine infections. Asian Spine J.
- 2016;10(1):176–183.
 Pull ter Gunne AF, Hosman AJF, Cohen DB, Schuetz M, Habil D, van Laarhoven CJHM, et al. A methodological systematic review on surgical site infections
- following spinal surgery: part 1: risk factors. Spine. 2012;37(24):2017–2033. Xing D, Ma JX, Ma XL, Song DH, Wang J, Chen Y, et al. A methodological, systematic review of evidence-based independent risk factors for surgical
- site infections after spinal surgery. Eur Spine J. 2013;22(3):605-615. Ogihara S, Yamazaki T, Maruyama T, Oka H, Miyoshi K, Azuma S, et al. Prospective multicenter surveillance and risk factor analysis of deep surgical site infection after posterior thoracic and/or lumbar spinal surgery
- in adults. J Orthop Sci. 2015;20(1):71–77. Fei Q, Li J, Lin J, Li D, Wang B, Meng H, et al. Risk factors for surgical site infec-
- tion after spinal surgery: a meta-analysis. World Neurosurg. 2016;95;507–515. Meng F, Cao J, Meng X. Risk factors for surgical site infections following spinal surgery. J Clin Neurosci. 2015;22(12):1862–1866.

Authors: Claus Simpfendorfer, Pouya Alijanipour, Caroline J. Granger

QUESTION 8: Should all patients with psoas abscesses be screened for both spine and hip infections?

RECOMMENDATION: Cross-sectional imaging with computed tomography (CT) and magnetic resonance imaging (MRI) will identify the source of secondary psoas abscesses in the majority of cases. If no other source is identified, consider cross-sectional imaging with CT or MRI for both the hip and spine in the setting of psoas abscess.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain 0% (Unanimous, Strongest Consensus)

RATIONALE

The iliopsoas is formed by two distinct and separate muscles - the psoas major and iliacus muscles. Each muscle is covered by its respective fascia and is typically associated with different disease entities [1]. The psoas major arises from the transverse processes of the lumbar vertebrae, exiting the pelvis beneath the inguinal ligament where it joins the iliacus (forming the iliopsoas tendon) and inserts on the lesser trochanter of the femur [2]. The iliacus muscle originates from the superior portion of the iliac fossa, superior lateral aspect of the sacrum and ventral sacroiliac and iliolumbar ligaments [2]. The medial portion of the iliacus muscle joins the psoas major tendon (forming the iliopsoas tendon) and inserts on the lesser trochanter. The lateral portion of the muscle inserts directly on the anterior and anteromedial aspect of the femur below the lesser trochanter [3].

The literature often does not delineate between the two muscles, referring to the combined muscles as the iliopsoas or simply the psoas muscle. Making a distinction between these muscles can help determine the source of infection. With regards to musculoskeletal infections, the majority of psoas muscle abscesses reflect extension from an adjacent spondylodiskitis or septic facet [4–7]. In contrast, iliacus muscle abscesses are secondary to extension of an underlying hip infection through the iliopsoas bursa or infectious sacroiliitis.

The iliopsoas bursa is the largest bursa in the body and communicates with the hip joint in 14% of the population [8]. Communication of the joint capsule with the iliopsoas bursa is likely increased following hip arthroplasty [9]. With the majority of the bursa located deep in the iliacus muscle, hip joint infections typically involve the iliacus muscle alone or less often both the iliacus and psoas muscle [1,10]. When the psoas muscle is involved, there should be visible communication with a distended iliopsoas bursa. This is in contrast to the psoas abscess associated with spondylodiscitis, which does not involve the bursa.

Both lumbar spine osteodiscitis and septic hip arthritis can be associated with psoas abscess [11]. The spine as primary source of infection for secondary psoas abscess should always be included in the differential diagnosis [12]. Studies have reported that 10–36% of secondary psoas abscess is caused by disc infection [13,14]. The anatomical proximity and communication of the psoas muscle to the hip joint capsule creates a potential transit for bacterial spread from spine to the hip joint or vice versa [15]. Screening patients with a psoas abscess for both hip and spine infection can prevent this harmful infectious spread. However, it should also be considered that the infection may simultaneously result in multiple infection sites from the same original hematogenous source of psoas abscess or spinal infection.

A non-coincidental association exists between psoas abscess and hip infection, both in the virgin hip joint and in a prosthetic hip joint. There have been multiple reports regarding the progression of the extension of psoas abscesses into the virgin or prosthetic hip joints [16–19]. In one study, the percentage of prosthetic hip infections with associated psoas abscesses has been reported to be as high as 12% [19]. Hematogenous prosthetic infection and a medical history of neoplasm have been reported as risk factors of psoas abscess in patients with an infected hip replacement [19]. Psoas abscesses may also cause relapse of prosthetic hip infection.

It is recommended that practitioners screen patients with

psoas abscesses for hip infection and spinal infection due to their anatomical communication, relationship in etiology and co-prevalence. Clinicians should be aware of the potential communication between the lumbar spine and hip joint via the psoas muscle and iliopsoas bursa. Successful treatment outcomes of psoas abscess are not only related to its early diagnosis, but also to the prompt detection of its spread to adjacent organs with potentially devastating outcomes, including the neural elements of spine and a prosthetic hip joint.

REFERENCES

- Shyam Kumar AJ, Hickerton B, Smith IC, Sinha A. Iliacus abscess: an entity to be differentiated from psoas abscess: a review of 15 cases. Eur J Orthop Surg Traumatol. 2007;17:477-478
- Gray H, Williams PL, Bannister LH. Gray's anatomy: the anatomical basis of medicine and surgery. 38th ed. New York, NY: Churchill Livingstone; 1995.
- Polster JM, Elgabaly M, Lee H, Klika A, Drake R, Barsoum W. MRI and gross anatomy of the iliopsoas tendon complex. Skeletal Radiol. 2008;37(1):55-58.
- Navarro Lopez V, Ramos JM, Meseguer V, et al. Microbiology and outcome of iliopsoas abscess in 124 patients. Medicine (Baltimore). 2009;88(2):120-
- Mückley T, Schütz T, Kirschner M, Potulski M, Hofmann G, Bühren V. Psoas abscess: the spine as a primary source of infection. Spine (Phila Pa 1976). 2003;28(6):E106-E113.
- Wong OF, Ho PL, Lam SK. Retrospective review of clinical presentations, microbiology, and outcomes of patients with psoas abscess. Hong Kong
- Med J. 2013;19(5):416–423. Dietrich A, Vaccarezza H, Vaccaro CA. Iliopsoas abscess: presentation, management, and outcomes. Surg Laparosc Endosc Percutan Tech. 2013;23(1):45-48.
- Chandler SB. Studies on the inguinal region; the inguinal canal. Anat Rec. 1950;107(1):93-102.
- Steinbach LS, Schneider R, Goldman AB, Kazam E, Ranawat CS, Ghelman B. Bursae and abscess cavities communicating with the hip. Diagnosis using arthrography and CT. Radiology. 1985;156(2):303–307.
 Dauchy FA, Dupon M, Dutronc H, et al. Association between psoas
- abscess and prosthetic hip infection: a case-control study. Acta Orthop. 2009;80(2):198-200.
- Penado S, Espina B, Francisco Campo J. [Abscess of the psoas muscle. Description of a series of 23 cases]. Enferm Infecc Microbiol Clin.
- Mückley T, et al. Psoas abscess: the spine as a primary source of infection. Spine. 2003;28:E106-E113.
- Ricci MA, Rose FB, Meyer KK. Pyogenic psoas abscess: worldwide variations in etiology. World J Surg. 1986;10:834-843
- Walsh TR, et al. Changing etiology of iliopsoas abscess. Am J Surg. 1992;163:413-416.
- Sadat-Ali M, al-Habdan I, Ahlberg A. Retrofascial nontuberculous psoas abscess. Int. Orthop. 1995;19:323-326
- Beredjiklian PK, et al. Prevertebral abscess with extension into the hip joint.
- Am J Órthop. 2001;30:572–575. Ellanti P, Moriarity A, Barry S, McCarthy T. Radiographic progression of septic arthritis of the hip. BMJ Case Rep. 2015. Kumagai K, Ushiyama T, Kawasaki T, Matsusue Y. Extension of lumbar
- spine infection into osteoarthritic hip through psoas abscess. J Orthop Sci. 2005;10:91-94.
- Navarro López V, et al. Microbiology and outcome of iliopsoas abscess in 124 patients. Medicine. 2009;88:120-130.

1.5. PREVENTION: WOUND CARE

Author: Carles Pigrau

QUESTION 1: Is negative pressure wound therapy (NPWT) safe on spinal wounds in patients with a cerebrospinal fluid (CSF) leak?

RECOMMENDATION: NPWT may be harmful in patients with a CSF leak, leading to severe neurological sequelae.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

Spine

RATIONALE

Intracranial hypotension may develop after dural puncture or spinal surgery by accidental intraoperative opening of the dura. As a complication to this, several cases of accidental drainage after spinal surgery and application of negative pressure suction devices (NPSDs) have been reported [1-4]. Secondarily, intracranial hypotension may develop leading to tonsillar herniation, subdural hemorrhage, severe neurological sequel and even death.

Recently, Sporns et al. reviewed the literature published in reference to patients diagnosed with postsurgical or post-traumatic intracranial hypotension [1,4]. In 24 relevant reports that included 27 cases, in 15 cases a NPSD (including NPWT or pleural drainage after thoracic surgery or traumatism) was applied, ten had no negative pressure devices and two could not be determined for application of a suction drain. All patients with NPSD had severe neurological symptoms, while only mild symptoms were observed in cases without such devices. They concluded that the increasing use of NPSDs causes the reported condition and that acute intracranial hypotension should be considered as an explanation of postoperative neurological symptoms or coma after cranial or spinal surgery. A precise radiological examination (preferably with magnetic resonance imaging) can help to rule out intracranial hypotension and dural laceration.

In conclusion, in patients with spinal wounds, NPSDs (including pleural drainages) may be harmful and lead to more severe neurological sequel than those cases with liquoral hypotension secondary to dural laceration without negative pressure devices.

REFERENCES

- Sporns PB, Zimmer S, Hanning U, Zoubi T, Wölfer J, Herbort M, et al. Acute tonsillar cerebellar herniation in a patient with traumatic dural tear and VAC therapy after complex trauma. Spine J. 2015;15:e13–e16. doi:10.1016/j. spinee.2015.04.025.
- Grahovac G, Vilendecic M, Chudy D, Srdoc D, Skrlin J. Nightmare complication after lumbar disc surgery: cranial nontraumatic acute epidural hematoma. Spine. 2011;36:E1761–E1764. doi:10.1097/BRS.ob013e31821cb9fd.
- Jung YY, Ju CI, Kim SW. Bilateral subdural hematoma due to an unnoticed dural tear during spine surgery. J Korean Neurosurg Soc. 2010;47:316–318. doi:10.3340/jkns.2010.47-4316. Sporns PB, Schwindt W, Cnyrim CD, Heindel W, Zoubi T, Zimmer S, et al.
- Undetected dural leaks complicated by accidental drainage of cerebrospinal fluid (CSF) can lead to severe neurological deficits. Rofo. 2016;188:451-458. doi:10.1055/s-0035-1567034.

Author: Barrett Boody

QUESTION 2: What are the risks and benefits for the use of vacuum-assisted closure (VAC) devices/PICO dressings following spine surgery?

RECOMMENDATION: The use of incisional VAC therapy (such as PICO dressings) is limited, but available literature supports its use in the prevention of dehiscence and surgical site infection (SSI) in posterior thoracolumbar deformity surgery.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 86%, Disagree: 14%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

Multiple case series and case reports have been published supporting the use of VAC therapy for staged treatment of deep/ subfascial SSI in spine surgery, with the common use being at index or second debridement, followed by multiple VAC changes until the wound is suitable for closure [1–4]. The specific VAC techniques (such as fascia open or closed, number of suction devices, suction settings, etc.) is poorly described in available studies. Ploumis reported on 73 patients undergoing VAC therapy for deep SSI, noting an average of 1.4 procedures following VAC placement (including closure) and closure of wound at an average of 7 days. They noted that methicillin-resistant Staphylococcus aureus (MRSA) and polymicrobial wound infections were more likely to require subsequent debridement after index VAC placement prior to definitive closure [2]. Similarly, Mehbod described 20 similar patients with deep SSI following spine surgery treated with VAC therapy, with an average of 2.2 procedures (including closure) following index VAC placement and resolution of infection in all patients and closed wounds by 6 months [3]. Canavese described 33 pediatric patients treated with VAC therapy for deep SSI after thoracolumbar spine surgery, with only 1 case ultimately requiring partial removal of implants [5].

Complications for VAC therapy have also been widely described, including need for reoperation and/or revision of hardware, bleeding, flap closure or skin grafting, retention of foam sponge fragments and cerebrospinal fluid (CSF) leaks resulting in neurologic complications (coma, brain herniation and intracranial hemorrhage) [1,2, 6–8]. The use of VAC therapy in the setting of CSF leak should be avoided due to risks of tonsillar herniation [7]. While VAC therapy over dura has been described in cranial surgery, no publication specifically described the application of sponges over dura in spine surgery. Multiple cranial publications describe the technique for dural application as the use of the "white" sponge (polyvinyl foam), as it is hydrophilic and less adherent, with lower suction pressures (~ 50 mmHg) [9,10].

The only available paper on the application of incision VAC therapy (such as PICO dressings) for spine surgery was published by Adogwa et al., who reviewed 160 posterior thoracolumbar deformity surgeries, of which 46 used incisional negative pressure wound therapy for 3 days. The authors reported lower rates of wound dehiscence (6.38% vs. 12.28%) and lower SSI rates (10.63% vs. 14.91%) for the incisional negative pressure wound therapy group, both reaching statistical significance (p < 0.05)[11].

REFERENCES

Jones GA, Butler J, Lieberman I, Schlenk R. Negative-pressure wound therapy in the treatment of complex postoperative spinal wound infections: complications and lessons learned using vacuum-assisted closure. J Neurosurg Spine. 2007;6(5):407-411.

- Ploumis A, Mehbod AA, Dressel TD, Dykes DC, Transfeldt EE, Lonstein JE. Therapy of spinal wound infections using vacuum-assisted wound closure: risk factors leading to resistance to treatment. I Spinal Disord Tech.
- Mehbod AA, Ogilvie JW, Pinto MR, et al. Postoperative deep wound infecwound closure. J Spinal Disord Tech. 2005;18(1):14-17.
 Watt JP, Dunn RN. The use of vacuum dressings for dead space management in deep surgical site infections allows implant and bone graft retention.
- Global Spine J. 2017;7(8):756-761. Canavese F, Krajbich JI. Use of vacuum assisted closure in instrumented spinal deformities for children with postoperative deep infections. Indian J Orthop. 2010;44(2):177–183. Dessy LA, Serratore F, Corrias F, Parisi P, Mazzocchi M, Carlesimo B. Reten-
- tion of polyurethane foam fragments during VAC therapy: a complication to be considered. Int Wound J. 2015;12(2):132–136.
- Sporns PB, Zimmer S, Hanning U, et al. Acute tonsillar cerebellar herniation in a patient with traumatic dural tear and VAC therapy after complex
- trauma. Spine J. 2015;15(7):e13–e16. Sporns PB, Schwindt W, Cnyrim CD, et al. Undetected dural leaks complicated by accidental drainage of cerebrospinal fluid (CSF) can lead to severe neurological deficits. RoFo. 2016;188(5):451-458.
 Prince N, Blackburn S, Murad G, et al. Vacuum-assisted closure therapy to
- the brain: a safe method for wound temporization in composite scalp and
- calvarial defects. Ann Plast Surg. 2015;74 Suppl 4:S218-S221. Mohammed-Ali RI, Khurram SA, Nahabedian V, Smith AT. Vacuum-assisted dressing for promoting granulation over the dura: technical note. British J
- Oral Maxillofac Surg. 2012;50(1):88-89.

 Adogwa O, Fatemi P, Perez E, et al. Negative pressure wound therapy reduces incidence of postoperative wound infection and dehiscence after long-segment thoracolumbar spinal fusion: a single institutional experience. Spine J. 2014;14(12):2911–2917.

Author: Jeffrey A. Rihn

QUESTION 3: What type of surgical dressing is most effective for lowering rates of surgical site infection (SSI) in patients undergoing spine surgery?

RECOMMENDATION: There are no randomized studies comparing the use of incisional negative pressure wound therapy (NPWT) to standard dry dressings in spine surgery. The World Health Organization (WHO) recommends the use of incisional NPWT for high risk surgical wounds to reduce the risk of SSI.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 86%, Disagree: 0%, Abstain: 14% (Super Majority, Strong Consensus)

RATIONALE

Incisional NPWT in the form of commercially available incisional suction dressings has recently gained popularity in the management of high-risk wounds in orthopaedic surgery.

These dressings are used at the time of index surgery primarily, with the aim of preventing wound complications such as SSI. Incisional NPWT protects the healing wound by preventing wound edge motion, improving of blood supply, removing of excess fluid and stimulating granulation tissue. A recent meta-analysis of all randomized and case-controlled trials comparing incisional NPWT to standard of care showed a reduction in SSI (50%), wound dehiscence and hospital length of stay [1]. In a pig spine model, Glaser showed improved early biomechanical properties as well as cosmesis in wounds dressed with incisional NPWT compared to standard dry dressings [2].

There are only two studies that have investigated incisional NPWT after spine surgery. A single-institution retrospective casecontrol study from Duke University showed a 50% decrease in wound dehiscence and a 30% decrease in SSI after a change to incisional NPWT dressing for thoracolumbar deformity wounds [3]. Similarly, a small randomized trial by Nordmeyer et al. showed a decrease in seroma and the need for nursing wound care intervention in patients who were treated with incisional NPWT [4]. The authors hypothesized that a decrease in seroma may lead to decreased SSI, but the study was underpowered to show this difference.

The 2016 WHO recommendations on intraoperative and postoperative measures for SSI prevention proposed prophylactic NPWT on primarily closed surgical incisions in high-risk wounds to reduce the incidence of SSI [5]. This recommendation drew on evidence from abdominal, thoracic and orthopaedic surgery.

In the absence of high-quality randomized trials and given the WHO recommendation, it would be reasonable to use incisional

NPWT in settings where the surgeon believes the wound is at risk of infection or breakdown. Spine wounds at high risk of infection include those in patients with diabetes, increased BMI, extended operative times and chronic steroid use [6,7]. In the pediatric spine population, risk factors for SSI include high weight centile, neuromuscular scoliosis, greater comorbidities and prolonged operative time [8].

- Strugala V, Martin R. Meta-analysis of comparative trials evaluating a prophylactic single-use negative pressure wound therapy system for the prevention of surgical site complications. Surg Infect (Larchmt).
- 2017;18:810–819. doi:10.1089/Sur.2017;156. Glaser DA, Farnsworth CL, Varley ES, Nunn TA, Sayad-Shah M, Breisch EA, et al. Negative pressure therapy for closed spine incisions: a pilot study.
- Wounds. 2012;24;308–316.
 Adogwa O, Fatemi P, Perez E, Moreno J, Gazcon GC, Gokaslan ZL, et al. Negative pressure wound therapy reduces incidence of postoperative wound infection and dehiscence after long-segment thoracolumbar spinal fusion: a single institutional experience. Spine J. 2014;14:2911–2917. doi:10.1016/j. spinee.2014.04.011.
- Nordmeyer M, Pauser J, Biber R, Jantsch J, Lehrl S, Kopschina C, et al. Negative pressure wound therapy for seroma prevention and surgical incision treatment in spinal fracture care. Int Wound J. 2016;13:1176–1179. doi:10.1111/ iwj.12436.
- Allegranzi B, Zayed B, Bischoff P, Kubilay NZ, de Jonge S, de Vries F, et al. New WHO recommendations on intraoperative and postoperative measures for surgical site infection prevention: an evidence-based global perspective. Lancet Infect Dis. 2016;16:e288–e303. doi:10.1016/S1473-3099(16)30402-9. Olsen MA, Mayfield J, Lauryssen C, Polish LB, Jones M, Vest J, et al. Risk factors
- For surgical site infection in spinal surgery. J Neurosurg. 2003;98:149–155. Sebastian A, Huddleston P, Kakar S, Habermann E, Wagie A, Nassr A. Risk factors for surgical site infection after posterior cervical spine surgery: an analysis of 5,441 patients from the ACS NSQIP 2005-2012. Spine J. 2016;16:504–
- 509. doi:10.1016/j.spinee.2015.12.009. Croft LD, Pottinger JM, Chiang HY, Ziebold CS, Weinstein SL, Herwaldt LA. Risk factors for surgical site infections after pediatric spine operations. Spine. 2015;40:E112-E119. doi:10.1097/BRS.0000000000000693.

Diagnosis

2.1. DIAGNOSIS: GENERAL PRINCIPLES

Authors: Robert Sawyer, Joseph K. Weistroffer, Anna White

QUESTION 1: What is the definition of surgical site infection (SSI) in spinal surgery?

RECOMMENDATION: We recommend utilizing the definition provided by the Centers for Disease Control and Prevention (CDC), National Health-care Safety Network (NHSN) Patient Safety Component Manual, Chapter 9: Surgical Site Infection (SSI) Event.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

The most persuasive argument for adopting the CDC's definition for an SSI lies in utilizing search protocols to map International Classification of Disease, 10th revision, Procedure Classification System (ICD-10-PCS) and Current Procedural Terminology (CPT) codes when querying databases.

The CDC definition is the accumulation of multiple years of planning/tracking and modifying this instrument via annual reviews and input from professionals worldwide. The description includes such categorical sub-elements as the definition of an operative procedure and the definition of an operating room. It includes criteria for the sub-classifications of a superficial incisional SSI, deep incisional SSI and organ/space SSI [1]. The CDC's definition delineates the exclusion of such events as cellulitis, stitch abscesses, as well as stab wound or pin site infections. It also defines such infections about primary or secondary wounds and the surveillance periods for SSI following operative procedures. Furthermore, numerous spine-related studies have utilized the same definition put forth by the CDC [2–5].

Adopting a thorough and uniform definition for SSI is imperative, as studies have shown that the rate of SSI following spine

surgery varies based on the definition used [6]. In addition, having a standardized definition will improve surveillance, provide consistency among studies and improve overall patient care.

REFERENCES

- Surgical Site Infection (SSI) Event. https://www.cdc.gov/nhsn/pdfs/ pscmanual/9pscssicurrent.pdf. 2018:32.
- [2] Petignat C, Francioli P, Harbarth S, Regli L, Porchet F, Reverdin A, et al. Cefuroxime prophylaxis is effective in noninstrumented spine surgery: a double-blind, placebo-controlled study. Spine. 2008;33:1919–1924. doi:10.1097/BRS.0b013e31817d97cf.
- [3] Anderson PA, Savage JW, Vaccaro AR, Radcliff K, Arnold PM, Lawrence BD, et al. Prevention of surgical site infection in spine surgery. Neurosurgery. 2017;80:S114–S123. doi:10.1093/neuros/nyw066.
- [4] Olsen MA, Nepple JJ, Riew KD, Lenke LG, Bridwell KH, Mayfield J, et al. Risk factors for surgical site infection following orthopaedic spinal operations. J Bone Joint Surg Am. 2008;90:62–69, doi:10.2106/JBJS.F.01515.
- Bone Joint Surg Am. 2008;90:62-69. doi:10.2106/JBJS.F.01515.

 Butler JS, Wagner SC, Morrissey PB, Kaye ID, Sebastian AS, Schroeder GD, et al. Strategies for the prevention and treatment of surgical site infection in the lumbar spine. Clin Spine Surg. 2018. doi:10.1097/BSD.00000000000000535.
- [6] Nota SPFT, Braun Y, Ring D, Schwab JH. Incidence of surgical site infection after spine surgery: what is the impact of the definition of infection? Clin Orthop Relat Res. 2015;473:1612–1619. doi:10.1007/s11999-014-3933-y.

Author: Claus Simpfendorfer

QUESTION 2: What defines delay in the diagnosis of a spine infection?

RECOMMENDATION: There is no clear or established definition of delayed diagnosis for spine infection.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

The diagnosis of spinal infections is often delayed from one to three months from the onset of symptoms [1,2]. Delay in diagnosis

is frequently secondary to nonspecific symptoms including back and neck pain. A couple of studies have used delayed diagnosis

of greater than eight weeks as a predictor of lower recovery rates, neurologic deficits and long-term disability [2–4]. A recent study by Issa et al. demonstrated that the percent of positive cultures from blood and/or biopsy decreases as the delay in diagnosis increases [2–5].

Jean et al. looked at predictors of delayed diagnosis and found that X-rays resulted in an increased delay from 14 days to 34.7 days [6]. It is presumed that, although delaying diagnosis, X-ray findings (either normal or demonstrating degenerative changes) provide the physician with reassurance. Alternatively, Jean et al. found that fever at initial presentation, elevated C-reactive protein (CRP) and blood cultures shortened the time to diagnosis [6]. The most significant impact was the elevated CRP which shortened the diagnostic delay from 73 days to 17 days [6]. It is therefore suggested that CRP be routinely checked in cases of new onset or sudden increased back pain [6,7]. Furthermore, if CRP is elevated or if there is clinical suspicion for spine infection, MRI with gadolinium should be performed [8].

REFERENCES

- [1] Cottle L, Riordan T. Infectious spondylodiscitis. J Infect. 2008;56:401–412.
- doi:10.1016/j.jinf.2008.02.005.

 [2] McHenry MC, Easley KA, Locker GA. Vertebral osteomyelitis: long-term outcome for 253 patients from 7 Cleveland-area hospitals. Clin Infect Dis. 2002;34:1342-1350. doi:10.1086/340102.
- O'Daly BJ, Morris SF, O'Rourke SK. Long-term functional outcome in pyogenic spinal infection. Spine. 2008;33:E246–E253. doi:10.1097/ BRS Oborge18668872
- BRS.0b013e31816b8872.

 [4] Solis Garcia del Pozo J, Vives Soto M, Solera J. Vertebral osteomyelitis: long-term disability assessment and prognostic factors. J Infect 2007;54:129–134. doi:10.1016/j.iinf.2006.0.013.
- [5] Issa K, Pourtaheri S, Vijapura A, Stewart T, Sinha K, Hwang K, et al. Delay in diagnosis of vertebral osteomyelitis affects the utility of cultures. Surg Technol Int. 2016;29:379–383.
- [6] Jean M, Irisson J-O, Gras G, Bouchand F, Simo D, Duran C, et al. Diagnostic delay of pyogenic vertebral osteomyelitis and its associated factors. Scand J
- Rheumatól. 2017;46:64–68. doi:10.3109/03009742.2016.1158314.
 [7] Siemionow K, Steinmetz M, Bell G, Ilaslan H, McLain RF. Identifying serious causes of back pain: cancer, infection, fracture. Cleve Clin J Med. 2008;75:557–
- [8] Dunbar JA, Sandoe JA, Rao AS, Crimmins DW, Baig W, Rankine JJ. The MRI appearances of early vertebral osteomyelitis and discitis. Clin Radiol. 2010;65:974–981. doi:10.1016/j.crad.2010.03.015.

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Authors: John Koerner, David Kaye

QUESTION 3: Is there an optimal window for diagnosis of an early spine infection?

RECOMMENDATION: There is no defined window, but early diagnosis of a postoperative spine infection (up to three months from time of surgery) treated with surgical debridement and antibiotics often allows for retention of instrumentation.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

Although the evidence regarding this topic is from low-quality studies, the findings and recommendations are consistent. Most postoperative spinal infections in adults present early, typically within the first three months [1]. Early diagnosis and debridement typically allows for retention of implants when present [1]. Implant removal due to infection can result in satisfactory results and eradicate infection, but can lead to malalignment and pseudarthrosis [2].

Early spine infections (< three months after surgery) treated with irrigation and debridement have improved outcomes compared to before surgery, but cause increased back pain and a lower probability of achieving a minimal clinically important difference [3].

In a cohort study of 51 patients who developed a postoperative spinal implant infection, prompt treatment (< 3 months) with debridement allowed for implant preservation in 41 patients, versus 10 patients in which treatment was delayed and implants were removed [4]. Another case series identified 26 postoperative infections, of which 24 were able to be treated without removal of implants by aggressive debridement and secondary closure [5]. Early identification and treatment can often allow for implant retention compared to delayed presentation, when implants may need to be removed [6–8].

Late spine infections are, however, seen more commonly in idiopathic scoliosis cases [9]. In a case-controlled series of 236 patients, seven developed an infection [10]. One was early and the other six were diagnosed at an average of 34.2 months postoperatively.

It is typical for patients to have symptoms of low back pain for 4 to 10 weeks prior to diagnosis of spondylodiscitis [11,12]. Although

most studies recommend early treatment, no specific timeframe could be identified that definitely leads to better outcomes.

- [1] Mok JM, Guillaume TJ, Talu U, Berven SH, Deviren V, Kroeber M, et al. Clinical outcome of deep wound infection after instrumented posterior spinal fusion: a matched cohort analysis. Spine. 2009;34:578–583. doi:10.1097/BRS.0b019e31819a827c.
- [2] Kim J Il, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infection after instrumented Spinal Fusion. J Spinal Disord Tech. 2010;23:258–265. doi:10.1097/BSD.0b013e3181a9452c.
- [3] Petilon JM, Glassman SD, Dimar JR, Carreon LY. Clinical outcomes after lumbar fusion complicated by deep wound infection: a case-control study. Spine. 2012;37:1370–1374. doi:10.1097/BRS.ob013e31824a4d93.
- [4] Chen SH, Lee CH, Huang KC, Hsieh PH, Tsai SY. Postoperative wound infection after posterior spinal instrumentation: analysis of long-term treatment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014-3636-9.
 [5] Picada R, Winter RB, Lonstein JE, Denis F, Pinto MR, Smith MD, et al. Post-
- [5] Picada R, Winter RB, Lonstein JE, Denis F, Pinto MR, Smith MD, et al. Post-operative deep wound infection in adults after posterior lumbosacral spine fusion with instrumentation: incidence and management. J Spinal Disord. 2000;13:42–45. doi:10.1097/00002517-200002000-00009.
- [6] Glassman SD, Dimar JR, Puno RM, Johnson JR. Salvage of instrumental lumbar fusions complicated by surgical wound infection. Spine (Phila Pa 1976). 1996;21:2163-2169. doi:10.1097/00007632-199609150-00021.
- [7] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913–920. doi:10.1086/512194.
- [8] Bose B. Delayed infection after instrumented spine surgery: case reports and review of the literature. Spine J. 2003;3:394–399. doi:10.1016/\$1529-0420(02)000228
- [9] Richards BS, Emara KM. Delayed infections after posterior TSRH spinal instrumentation for idiopathic scoliosis: revisited. Spine. 2001;26:1990-1996. doi:10.1097/00007632-200109150-00009.
- [10] Rihn JA, Lee JY, Ward WT. Infection after the surgical treatment of adolescent idiopathic scoliosis: evaluation of the diagnosis, treatment, and impact on clinical outcomes. Spine. 2008;33:289–294. doi:10.1097/ BRS.0b013e318162016e.

- [11] Bettini N, Girardo M, Dema E, Cervellati S. Evaluation of conservative treatment of non specific spondylodiscitis. Eur Spine J. 2009;18. doi:10.1007/ s00586-009-0979-8.
- [12] Valancius K, Hansen ES, Hoy K, Helmig P, Niedermann B, Bünger C. Failure modes in conservative and surgical management of infectious spondylodiscitis. Eur Spine J. 2013;22:1837–1844. doi:10.1007/s00586-012-2614-3.

Author: Gregory Schroeder

QUESTION 4: How do early and late infectious complications differ following spine surgery?

RECOMMENDATION: Early infections, defined as occurring within 30 days of surgery, often present with local signs of infection such as increased surgical site pain, erythema, warmth and wound drainage. Conversely, late infections (> 90 days after surgery) commonly present with an insidious onset of chronic pain and implant failure/ pseudarthrosis if following a fusion.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 87%, Disagree: 0%, Abstain: 13% (Super Majority, Strong Consensus)

RATIONALE

Postoperative spine infection occurs at a rate of 0.7–16% depending on the procedure; the lumbar spine is the site of 51% of infections [1].

A postoperative infection is classified as early when it occurs within 30 days of the initial surgery. Early infections typically present with increasing back pain (83–100%) as the primary symptom [2,3]. Fever, weight loss, erythema, swelling, warmth, tenderness and elevated white blood cell (WBC) count may also be present, with fever having an incidence of 16–65% [2–4]. One of the most reliable and specific signs of early infection is increased wound drainage (67%) as it can occur in both deep and superficial infections [4].

A postoperative infection occurring three to nine months following surgery can be classified as a late infection. As opposed to early infections, late infections typically present with delayed symptoms such as lack of adequate fusion, chronic pain or implant failure months after surgery [5]. Local symptoms may also occur, including increased pain and tenderness at the incision site. Wound drainage may occur but is less common than in early infections [5].

Complications of postoperative spine infection include impairment of function, significant morbidity and increased health care costs approximating up to \$200,000 per patient [1,3]. Increase in hospital stay and increased rates of repeat surgery have also been observed.

Gram-positive bacteria, specifically *Staphylococcus aureus*, are responsible for approximately 45% of spine infections [6]. Other

gram-positives such as *Staphylococcus epidermis* and *Enterococcus* as well as gram-negatives *Pseudomonas aeruginosa* and *Escheria coli* have been observed at lower incidences [1,2,6]. There is no clear association between type of surgical procedure and bacteria strain. However, gram-negatives tend to present more commonly in sacral and lumbar regions [6]. Fungal infections may occur in immunocompromised patients. *C. acnes* has recently been identified as another potential causative organism [2]. No significant difference has been observed in the type of organism present in early and late infections.

REFERENCES

- [1] Pawar A, Biswas S. Postoperative spine infections. Asian Spine J. 2016;10:176–183.
- [2] Chahoud J, Kanafani Z, Kanj S. Surgical site infections following spine surgery: eliminating the controversies in the diagnosis. Front Med (Lausanne). 2014;1:17.
- [3] Parchi P, Evangelisti G, Andreani L. Postoperative spine infections. Orthop
- Rev. 2015;7(3):5900.
 [4] Kasliwal MK, Tan LA, Traynelis VC. Infection with spinal instrumentation: review of pathogenesis, diagnosis, prevention, and management. Surg Neurol Int. 2013;4(Suppl 5):S392.
 [5] Radcliff K, et al. What is new in the diagnosis and prevention of spine
- [5] Radcliff K, et al. What is new in the diagnosis and prevention of spine surgical site infections? Spine J. 2015;15:336–347.
 [6] Abdul-Jabbar A, et al. Surgical site infections in spine surgery: identifi-
- [6] Abdul-Jabbar A, et al. Surgical site infections in spine surgery: identification of microbiologic and surgical characteristics in 239 cases. Spine. 2013;38:E1425–E1431.

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Author: Bryan Alexander

QUESTION 5: Are there patients with degenerative pathology, such as disc herniations, who are actually infected with a low-grade infection (e.g., *Propionibacterium acnes*)?

RECOMMENDATION: The association between the *Cutibacterium acnes* (*C. acnes*) (formerly *P. acnes*) and degenerative spinal disease is inconclusive.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 86%, Disagree: 14%, Abstain: 0% (Super Majority, Strong Consensus)

Spine

RATIONALE

The initial connection between potential low-level infection and degenerative spinal pathology was drawn when a group identified over half of discectomies performed for disc herniation as culture positive for *C. acnes* or coagulase-negative *Staphylococcus spp* [1]. A large number of predominantly small studies have since come to opposite conclusions on the connection between these bacteria and degenerative spinal disease, most commonly evaluated radiographically by the presence of Modic changes (examples of those finding no relationship [2–7] versus those finding a correlation [8–12]). One controversial placebo-controlled, double-blinded trial administered extended-duration antibiotic therapy to those patients with Modic type 1 changes and demonstrated better pain resolution in those receiving antibiotics [8].

Recent systematic reviews, each published in 2015, independently concluded that while there was strong evidence from multiple studies that patients undergoing spinal surgery have increased rates of bacteria at the site of degenerative disease of spine, causation between that finding and the pathologic changes resulting in back pain were unclear [1,13,14].

One important cause for heterogeneity in the data is the possibility that microbiologic sampling could be more readily contaminated with bacteria based on differences in surgical and collection technique [3,15]. However, this does not fully explain the fact that in clinical studies, C. acnes is consistently the most common, if not only, organism isolated. Recent studies, including control groups of patients not anticipated to have infectious etiologies for their spinal condition, have also noted increased rates of bacterial presence in degenerative disease compared to patients without degenerative disease [2,16]. Methods attempting to disrupt biofilm-encapsulated bacteria have attempted to explain negative culture results from earlier studies [10,17]. Similarly, molecular subtyping of C. acnes allows for better characterization of these isolates into those more likely to be routine skin contamination from those more likely to be pathogenic [2,17-19]. These studies have demonstrated a mixture of these subtypes present, with those generally not representing skin flora predominating. Recent studies have additionally investigated histologic methods [20], inflammatory cytokine responses [16,21] and proteomic analysis [22] in addition to bacterial presence as a marker for true infection. Finally, some groups have recently used animal models to attempt to support a connection between bacterial inoculation and symptomatic spinal pathology [23,24].

Though still unverified, there is an enlarging body of evidence using modern techniques and accounting for technical limitations in earlier studies for the role of infection in at least some types of degenerative spinal pathology. A well-designed, multicenter trial effort, which successfully confirms this connection would allow for reasonable consideration of further studies utilizing antibiotic therapy as a non-invasive therapy option for degenerative disc disease.

- Stirling A, Worthington T, Rafiq M, Lambert PA, Elliott TS. Association between sciatica and Propionibacterium acnes. Lancet. 2001;357(9273):2024-
- Coscia MF, Denys GA, Wack MF. Propionibacterium acnes, coagulase-negative Staphylococcus, and the "biofilm-like" intervertebral disc. Spine (Phila Pa 1976). 2016;41(24):1860-1865.

- Rigal J, Thelen T, Byrne F, Cogniet A, Boissière L, Aunoble S, Le Huec JC. Prospective study using anterior approach did not show association between Modic 1 changes and low grade infection in lumbar spine. Eur Spine J. 2016;25(4):1000-1005. Arndt J, Charles YP, Koebel C, Bogorin I, Steib JP. Bacteriology of degenerated
- lumbar intervertebral disks. J Spinal Disord Tech. 2012;25(7):E211-E216. Wedderkopp N, Thomsen K, Manniche C, Kolmos HJ, Secher Jensen T,
- Leboeuf Yde C. No evidence for presence of bacteria in modic type I changes. Acta Radiol. 2009;50(1):65–70.
 Ben-Galim P, Rand N, Giladi M, Schwartz D, Ashkenazi E, Millgram M, Dekel
- S, Floman Y. Association between sciatica and microbial infection: true infection or culture contamination? Spine (Phila Pa 1976). 2006;31(21):2507-
- Alamin TF, Munoz M, Zagel A, Ith A, Carragee E, Cheng I, Scuderi G, Budvytiene I, Banei N. Ribosomal PCR assay of excised intervertebral discs from patients undergoing single-level primary lumbar microdiscectomy. Eur Spine J. 2017;26(8):2038–2044.
- Albert HB, Sorensen JS, Christensen BS, Manniche C. Antibiotic treatment in patients with chronic low back pain and vertebral bone edema (Modic type 1 changes): a double-blind randomized clinical controlled trial of efficacy. Eur Spine J. 2013;22(4):697–707. Albert HB, Lambert P, Rollason J, Sorensen JS, Worthington T, Pedersen MB,
- Nørgaard HS, Vernallis A, Busch F, Manniche C, Elliott T. Does nuclear tissue infected with bacteria following disc herniations lead to Modic changes in
- the adjacent vertebrae? Eur Spine J. 2013;22(4):690–696.
 Capoor MN, Ruzicka F, Machackova T, Jancalek R, Smrcka M, Schmitz JE, Hermanova M, Sana J, Michu E, Baird JC, Ahmed FS, Maca K, Lipina R, Alamin TF, Coscia MF, Stonemetz JL, Witham T, Ehrlich GD, Gokaslan ZL, Mavrommatis K, Birkenmaier C, Fischetti VA, Slaby O. Prevalence of Propionibacterium acnes in intervertebral discs of patients undergoing lumbar microdiscectomy: a prospective cross-sectional study. PLoS One. 2016;11(8):e0161676.
- Zhou Z, Chen Z, Zheng Y, Cao P, Liang Y, Zhang X, Wu W, Xiao J, Qiu S. Relationship between annular tear and presence of Propionibacterium acnes in
- lumbar intervertebral disc. Eur Spine J. 2015;24(11):2496–2502. Aghazadeh J, Salehpour F, Ziaeii E, Javanshir N, Samadi A, Sadeghi J, Mirzaei F, Naseri Alavi SA. Modic changes in the adjacent vertebrae due to disc material infection with Propionibacterium acnes in patients with lumbar disc herniation. Eur Spine J. 2017;26(12):3129–3134. Ganko R, Rao PJ, Phan K, Mobbs RJ. Can bacterial infection by low virulent
- organisms be a plausible cause for symptomatic disc degeneration? A systematic review. Spine (Phila Pa 1976). 2015;40(10):E587–E592.
 Urquhart DM, Zheng Y, Cheng AC, Rosenfeld JV, Chan P, Liew S, Hussain SM, Cicuttini FM. Could low grade bacterial infection contribute to low back pain? A systematic review. BMC Med. 2015;13:13.
- . Javanshir N, Salehpour F, Aghazadeh J, Mirzaei F, Naseri Alavi SA. The distribution of infection with Propionibacterium acnes is equal in patients with
- cervical and lumbar disc herniation. Eur Spine J. 2017;26(12):3135-3140. Yuan Y, Chen Y, Zhou Z, Jiao Y, Li C, Zheng Y, Lin Y, Xiao J, Chen Z, Cao P. Association between chronic inflammation and latent infection of Propionibacterium acnes in non-pyogenic degenerated intervertebral discs: a pilot study. Eur Spine J. 2017 Oct 31. [Epub ahead of print].
- Capoor MN, Ruzicka F, Schmitz JE, James GA, Machackova T, Jancalek R, Smrcka M, et al. Propionibacterium acnes biofilm is present in intervertebral discs of patients undergoing microdiscectomy. PLoS One. 2017;12(4):e0174518.
- Rollason J, McDowell A, Albert HB, Barnard E, Worthington T, Hilton AC, Vernallis Å, et al. Genotypic and antimicrobial characterisation of Propioni-bacterium acnes isolates from surgically excised lumbar disc herniations. Biomed Res Int. 2013;2013:530382.
- Achermann Y, Goldstein EJ, Coenye T, Shirtliff ME. Propionibacterium acnes: from commensal to opportunistic biofilm-associated implant pathogen. Clin Microbiol Rev. 2014 Jul;27(3):419–440. Yuan Y, Zhou Z, Jiao Y, Li C, Zheng Y, Lin Y, Xiao J, Chen Z, Cao P. Histological
- identification of Propionibacterium acnes in nonpyogenic degenerated intervertebral discs. Biomed Res Int. 2017;2017:6192935
- Dudli S, Miller S, Demir-Deviren S, Lotz JC. Inflammatory response of disc cells against Propionibacterium acnes depends on the presence of lumbar
- Modic changes. Eur Spine J. 2017 Sep 7. [Epub ahead of print]. Rajasekaran S, Tangavel C, Aiyer SN, Nayagam SM, Raveendran M, Demonte NL, Subbaiah P, et al. ISSLS Prize in Clinical Science 2017: Is infection the possible initiator of disc disease? An insight from proteomic analysis. Eur
- Spine J. 2017;26(5):1384–1400. Dudli S, Liebenberg E, Magnitsky S, Miller S, Demir-Deviren S, Lotz JC. Propionibacterium acnes infected intervertebral discs cause vertebral bone marrow lesions consistent with Modic changes. J Orthop Res. 2016
- Aug;34(8):1447-1455. Chen Z, Zheng Y, Yuan Y, Jiao Y, Xiao J, Zhou Z, Cao P. Modic changes and disc degeneration caused by inoculation of Propionibacterium acnes inside intervertebral discs of rabbits: a pilot study. Biomed Res Int. 2016;2016;9612437.

Authors: Barrett Woods, Maja Babic

QUESTION 6: What is the diagnostic algorithm of patients with suspected hematogenous vertebral osteomyelitis? Is the algorithm different for patients with tuberculosis (TB)?

RECOMMENDATION: We support the diagnostic algorithm for suspected hematogenous vertebral osteomyelitis per Infectious Disease Society of America (IDSA) Clinical Practice Guidelines, 2015. Diagnostic algorithm is not different for patients with TB.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 87%, Disagree: 6%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Vertebral osteomyelitis typically occurs due to hematogenous seeding of the adjacent avascular disc from a distant foci [1]. Appropriate management is contingent upon timely diagnosis. Patients with vertebral osteomyelitis are commonly misdiagnosed and treated for degenerative pathology [2]. This often leads to a delay in treatment on average from two to four months [3]. The diagnosis of vertebral osteomyelitis is not challenging in patients with acute onset low back pain and fever. In this circumstance the diagnosis can be confirmed with a serologic test and imaging studies. However, fever and leukocytosis occur in approximately 45% of patients with bacterial vertebral osteomyelitis and very rarely in those with fungal, brucellar or mycobacterial infections [4,5]. Vertebral osteomyelitis should be suspected in patients who have recalcitrant back pain in the setting of elevated inflammatory markers. In 2015, the IDSA published Clinical Guidelines for the diagnosis and treatment of native vertebral osteomyelitis (NVO) in adults [6,7]. These guidelines provide an algorithmic approach to the diagnosis of NVO based on a systematic review of the literature.

Obtaining a detailed history is a critical portion of the diagnostic algorithm and should include any recent travel, infections, open wounds, recent antibiotic treatment and intravenous drug use. Patients who have back pain and a history of bacteremia, particularly *Staphylococcus aureus*, should be suspected of having vertebral osteomyelitis; therefore, further work up is warranted in these scenarios [8–10]. Patients with vertebral osteomyelitis typically present with back pain exacerbated by physical activity. Pain may not be isolated to the affected area and can radiate to the abdomen, hip, leg, scrotum, groin or perineum [11]. A full physical examination should be performed and include assessment of motor and sensory function. It takes three to six weeks after the onset of symptoms for bone destruction to be evident on plain radiographs. Thus, normal images do not exclude diagnosis.

Magnetic resonance imaging (MRI) should be obtained in patients with suspected vertebral osteomyelitis, as it has a sensitivity of 97%, specificity of 93% and an accuracy of 94% in diagnosing vertebral osteomyelitis [12,13]. Gadolinium enhancement is critical to appreciate paravertebral or epidural involvement [14]. A repeat MRI should be considered in two to four weeks in a patient suspected of vertebral osteomyelitis whose initial imaging study failed to show features consist with the diagnosis [15]. Imaging features consistent with TB infections include destruction of two or more contiguous vertebrae, extension along the anterior longitudinal ligament and disc infection, with or without a paraspinal mass or mixed soft tissue fluid collection [16]. In patients for whom MRI is not possible, a spine gallium/Tc99 bone scan is an alternative with a sensitivity and specificity of around 90% for diagnosing vertebral osteomyelitis [17,18].

Positron emission tomographic scanning is also highly sensitive for detecting osteomyelitis [19].

Serologic testing is important in the diagnostic algorithm of vertebral osteomyelitis. A minimum of two blood cultures should be obtained for patients with suspected vertebral osteomyelitis [20]. Blood cultures should be incubated for up to two weeks and should include aerobic, anaerobic and fungal. Leukocytosis has low sensitivity and specificity in the diagnosis with approximately 40% of patients with osteomyelitis having a normal white blood cell (WBC) count [21]. However, an elevated erythrocyte sedimentation rate (ESR) or C-reactive protein (CRP) in patients with back pain, though not specific, has a sensitivity that can range from 94% to 100% [22].

In patients with suspected vertebral osteomyelitis who reside in or have traveled to areas endemic for TB, a purified protein derivative (PPD) skin test can be performed; however, this test has a low sensitivity and specificity for diagnosis. An interferon-γ release assay has been shown to have a higher sensitivity than PPD, especially in immunocompromised patients with immune compromise [23]. Enzyme-linked immunospot assay has some diagnostic utility for TB and has been proven superior to PPD alone (sensitivity 82.8% vs. 58.6% and specificity, 81.3% vs. 59.4%, respectively) [24].

Empiric antibiotic therapy should not be initiated in aseptic patients without neurologic deficit until an image-guided biopsy can be obtained, especially if microbiologic diagnosis for a known associated organism has not been established by blood cultures or serologic tests [6]. Biopsy increases the likelihood of microbiologic diagnosis, improving the chance of successful medical management through targeted antibiotic therapy [25]. *S. aureus* bacteremia eliminates the need for biopsy, and antibiotics should not be delayed [8,22]. If biopsy is non-diagnostic, a repeat biopsy, image-guided or open biopsy, should be considered.

- Zimmerli W. Clinical practice. Vertebral osteomyelitis. N Engl J Med. 2010;362:1022-1029. doi:10.1056/NEJMcp0910753.
 Gupta A, Kowalski TJ, Osmon DR, Enzler M, Steckelberg JM, Huddleston
- [2] Gupta A, Kowalski TJ, Osmon DR, Enzler M, Steckelberg JM, Huddleston PM, et al. Long-term outcome of pyogenic vertebral osteomyelitis: a cohort study of 260 patients. Open Forum Infect Dis. 2014;1:ofu107. doi:10.1093/ofid/ofu107.
- [3] Gasbarrini AL, Bertoldi E, Mazzetti M, Fini L, Terzi S, Gonella F, et al. Clinical features, diagnostic and therapeutic approaches to haematogenous vertebral osteomyelitis. Eur Rev Med Pharmacol Sci. 2005;9:53–66.
- [4] Oztekin O, Calli C, Adibelli Z, Kitis O, Eren C, Altinok T. Brucellar spondylodiscitis: magnetic resonance imaging features with conventional sequences and diffusion-weighted imaging. Radiol Med. 2010;115:794–803. doi:10.1007/s11547-010-0530-3.
- [5] Song KJ, Yoon SJ, Lee KB. Cervical spinal brucellosis with epidural abscess causing neurologic deficit with negative serologic tests. World Neurosurg. 2012;78:375.e15-e19. doi:10.1016/j.wneu.2011.12.081.

- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. Executive summary: 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native verte bral osteomyelitis in adults. Clin Infect Dis. 2015;61:859-863. doi:10.1093/cid/
- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native vertebral osteomyelitis in adults. Clin Infect Dis. 2015;61:e26-e46. doi:10.1093/cid/civ482.
- Corrah TW, Enoch DA, Aliyu SH, Lever AM. Bacteraemia and subsequent vertebral osteomyelitis: a retrospective review of 125 patients. QIM. 2011;104:201–207. doi:10.1093/qjmed/hcq178.
- Mylona E, Samarkos M, Kakalou E, Fanourgiakis P, Skoutelis A. Pyogenic vertebral osteomyelitis: a systematic review of clinical characteristics. Semin Arthritis Rheum. 2009;39:10–17. doi:10.1016/j.semarthrit.2008.03.002.
- [10] Priest DH, Peacock JE. Hematogenous vertebral osteomyelitis due to Staphylococcus aureus in the adult: clinical features and therapeutic outcomes. 6outh Med J. 2005;98:854–862. doi:10.1097/01.smj.0000168666.98129.33.
- Wong-Chung JK, Naseeb SA, Kaneker SG, Aradi AJ. Anterior disc protrusion as a cause for abdominal symptoms in childhood discitis. A case report. Spine. 1999;24:918–920.
- Dagirmanjian A, Schils J, McHenry M, Modic MT. MR imaging of vertebral osteomyelitis revisited. Am J Roentgenol. 1996;167:1539-1543. doi:10.2214/ ajr.167.6.8956593
- Ledermann HP, Schweitzer ME, Morrison WB, Carrino JA. MR imaging findings in spinal infections: rules or myths? Radiology. 2003;228:506-514. doi:10.1148/radiol.228202075
- Schininà V, Rizzi EB, Rovighi L, de Carli G, David V, Bibbolino C. Infectious spondylodiscitis: magnetic resonance imaging in HIV-infected and HIV-uninfected patients. Clin Imaging. 2001;25:362–367.

- Dunbar JA, Sandoe JA, Rao AS, Crimmins DW, Baig W, Rankine JJ. The MRI appearances of early vertebral osteomyelitis and discitis. Clin Radiol. 2010;65:974–981. doi:10.1016/j.crad.2010.03.015.
- De Backer Al, Mortelé KJ, Vanschoubroeck JJ, Deeren D, Vanhoenacker FM, De Keulenaer BL, et al. Tuberculosis of the spine: CT and MR imaging
- Features. JBR-BTR. 2005;88:92–97. Love C, Patel M, Lonner BS, Tomas MB, Palestro CJ. Diagnosing spinal osteomyelitis: a comparison of bone and Ga-67 scintigraphy and magnetic resonance imaging. Clin Nucl Med. 2000;25:963-977.
- Palestro CJ, Torres MA. Radionuclide imaging in orthopedic infections.
- Semin Nucl Med. 1997;27:334-345. Ohtori S, Suzuki M, Koshi T, Yamashita M, Yamauchi K, Inoue G, et al. 18F-fluorodeoxyglucose-PET for patients with suspected spondylitis showing Modic change. Spine. 2010;35:E1599-E1603.
- Lee A, Mirrett S, Reller LB, Weinstein MP. Detection of bloodstream infections in adults: how many blood cultures are needed? J Clin Microbiol.
- 2007;45:3546-3548. doi:10.1128/JCM.01555-07. Siemionow K, Steinmetz M, Bell G, Ilaslan H, McLain RF. Identifying serious causes of back pain: cancer, infection, fracture. Cleve Clin J Med. 2008;75:557-
- Jensen AG, Espersen F, Skinhøj P, Frimodt-Møller N. Bacteremic Staphylo-
- coccus aureus spondylitis. Arch Intern Med. 1998;158:509–517. Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent tuberculosis infection: areas of uncertainty and recommendations for research. Ann Intern Med. 2007;146:340-354
- Yuan K, Zhong Z, Zhang Q, Xu S, Chen J. Evaluation of an enzyme-linked immunospot assay for the immunodiagnosis of atypical spinal tuberculosis (atypical clinical presentation/atypical radiographic presentation) in
- China. Braz J Infect Dis. 2013;17:529–537. doi:10.1016/j.bjid.2013.01.013. de Lucas EM, González Mandly A, Gutiérrez A, Pellón R, Martín-Cuesta L, laquierdo J, et al. CT-guided fine-needle aspiration in vertebral osteomy-elitis: true usefulness of a common practice. Clin Rheumatol. 2009;28:315– 320. doi:10.1007/s10067-008-1051-5.

Author: Taolin Fang

QUESTION 7: Should antibiotics be held prior to image-guided biopsy/aspiration for a suspected spine infection?

RECOMMENDATION: We recommend that prior to image-guided biopsy/aspiration for a suspected spine infection, all antibiotics should be withheld until after appropriate culture samples are obtained. Antibiotic administration, without aspiration/biopsy may be justified in patients who are critically ill and cannot withstand intervention or in patients with deteriorating neurological conditions.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

The definitive diagnosis of spinal osteomyelitis can be made only with isolation of the organism from a positive blood culture or biopsy and culture of the tissues from the region of the infection. Spinal biopsies may be performed using computed tomography (CT) or fluoroscopy for guidance in localizing the site of the suspected infection. The identification of the infecting organism is useful in directing antibiotic therapy. In suspected infection of the spine, biopsy and culture of the tissues from the affected site has been reported to be successful in the identification of the infecting organism in 46–91% of cases [1–5].

In real practice, there are some instances where antibiotic treatment is empirically instituted before the patient has been biopsied. Such cases may include patients who have been on antibiotics for other infections such as pneumonia or patients with surgical implants and prior deep wound infections who are on chronic antibiotic therapy. Theoretically, retrieval of a pathogen from the disc space or vertebral body may be compromised by previous or ongoing antibiotic treatment. However, we were unable to identify any high-quality randomized clinical trial comparing the culture results of the image-guided biopsy between patients who received empirical antibiotic treatment versus those who did not have any antibiotic treatment prior to biopsy.

There has been a general consensus of opinion that antibiotics should be withheld prior to biopsy of the site of suspected infection in an effort to improve the yield of culture [6,7]. A study by Rankine et al. found that the yield of biopsy in isolating the infecting organism was lower at 25% in patients who had received antibiotics compared to 50% yield in patients who had not received antibiotics [8]. It is important to note that not all studies agree with the notion of withholding antibiotics prior to biopsy of the infected site. A recent study by Sehn et al. [9] reported that four of 14 patients with a high suspicion for infection, who were confirmed to have been treated with antibiotics within 3 days of their biopsy, had positive cultures. The yield of culture was not different from the cohort of 92 patients who had not received antibiotics (28.6% vs. 30.4%, p = 0.86). Both of the reports were retrospective non-randomized studies with a relatively small sample size.

In the absence of randomized prospective data, and using the logic drawn from other fields of orthopaedic study related to this issue, we recommend that empirical treatment with antibiotics be withheld in patients with suspected infection of the spine until biopsy of site of suspected infection can be carried out. There are, however, circumstances (such as situations involving critically ill patients and those with deteriorating neurological status) in whom antibiotics may be started prior to the performance of biopsy.

REFERENCES

- Carragee EJ. Pyogenic vertebral osteomyelitis. J Bone Joint Surg Am. 1997;79:874-880.
- Patzakis MJ, Rao S, Wilkins J, et al. Analysis of 61 cases of vertebral osteomyelitis. Clin Orthop Relat Res. 1991;264:178-183.

- Tehranzadeh J, Tao C, Browning A. Percutaneous needle biopsy of the spine. Acta Radiol, 2007;48:860-868.
- Hadjipavlou AG, Kontakis GM, Gaitanis JN, et al. Effectiveness and pitfalls of percutaneous transpedicle biopsy of the spine. Clin Orthop Relat Res. 2003;411:54-60.
- Chew FS, Kline MJ. Diagnostic yield of CT-guided percutaneous aspiration procedures in suspected spontaneous infectious diskitis. Radiology, 2001;218:211-214
- Paluska SA. Osteomyelitis. Clin Fam Pract. 2004;6:127-156.
- An HS, Seldomridge JA. Spinal infections: diagnostic tests and imaging studies. Clin Orthop Relat Res. 2006;444:27-33.
 Rankine JJ, Barron DA, Robinson P, et al. Therapeutic impact of percuta-
- neous spinal biopsy in spinal infection. Post Grad Med J. 2004;80:607–609. Sehn JK, Gilula LA. Percutaneous needle biopsy in diagnosis and identification of causative organisms in cases of suspected vertebral osteomyelitis. Eur J Radiol. 2012;8:940-946.



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QUESTION 8: What is the incidence of infectious bacterial meningitis (PBM) following spinal surgery? Does the use of instrumentation affect this?

RECOMMENDATION: The incidence of PBM following spinal surgery varies from 0.1-0.4%. There is insufficient evidence to make any observations as to whether the use of instrumentation affects the incidence of PBM following spinal surgery.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

PBM is a potentially devastating complication following spinal surgery. It could occur after any primary elective spinal surgery with or without instrumentation, traumatic fracture-dislocation or surgical site infection after spinal instrumented surgery [1–3]. This also presents as a delayed complication after scoliosis surgery and through a dural tear with cerebrospinal fluid (CSF) leakage [4,5].

The early diagnostic differentiation from PBM and postoperative aseptic meningitis (PAM) is difficult and depends on CSF culture results [6-7]. The success in the treatment of patients with PBM depends on the stage of diagnosis, speed of diagnostic evaluation and appropriate anti-microbial and adjunctive therapy [8–9].

PBM is a potentially life-threatening infection with higher rates of mortality and significant disabling morbidity [9]. Pneumococcal meningitis is the most prevalent and is associated with a mortality of 30% [10]. PBM can also be caused by staphylococci [11], aerobic gram-negative bacilli (including P. aeruginosa) [12] and methicillinresistant Staphylococcus aureus (MRSA) [13].

The incidence of PBM is rare after spinal surgery and is considered to be related to incidental durotomy [14]. Patients who have the triad of fever, neck stiffness and consciousness disturbance during postoperative period should be suspected and subjected to further evaluations [14]. In a large retrospective study, Lin et al. reviewed 20,178 lumbar spinal surgeries and reported a PBM rate of 0.10% [14]. Another retrospective study by Twyman et al. reported the incidence of PBM to be 0.18% after spinal operations with and without instrumentation [15]. The incidence could be as high as 0.4% after spinal surgery, when epidural abscess, subdural empyema, brain abscess, bone-flap infections and wound infections are combined [16].

In their sub-analysis, Lin et al. found that dural tears, pseudomeningocele and poor wound healing contributed to the majority of the complications [14]. The optimal management of PBM required reoperation to repair dural tears and administration of parenteral antibiotics [17]. The occurrence of pseudomeningocele is a sequela of dural tear, imperfect suture of the dura or fascia and inappropriate administration of antibiotics [14,18,19]. Zhang et al. reported surgical intervention to be an effective method of treating PBM where initial conservative measures failed. They proposed the idea that it is important to consider the possibility of PBM in any patient with CSF leakage after spinal surgery. They recommended early diagnostic imaging and CSF cultures to ensure prompt diagnosis and treatment [20].

Spinal instrumentation surgery usually involves longer operative time, greater blood loss and a higher incidence of subsequent SSI compared to decompression surgery alone. These features of spinal instrumentation surgery could influence the incidence of PBM. There is little literature examining the potential association of instrumentation with PBM with no supporting evidence linking the use of instrumentation to the incidence of infectious meningitis after spinal surgery [14,15,20]. Therefore, based on available evidence, it is not possible to link the use of instrumentation during spine surgery with PBM.

- Cummings RJ. Recurrent meningitis secondary to infection after spinal arthrodesis with instrumentation. A case report. I Bone Joint Surg Am.
- Morris BJ, Fletcher N, Davis RA, Mencio GA. Bacterial meningitis after traumatic thoracic fracture-dislocation: two case reports and review of the litera-
- ture. J Orthop Trauma. 2010;24(5):e49–e53. doi:10.1097/BOT.0b013e3181b7f708. Schmidt M, Maxime V, Pareire F, Carlier R, Lawrence C, Clair B, Annane D. A lethal case of meningitis due to Lactobacillus rhamnosus as a late complication of anterior cervical spine surgery. J Infect. 2011;62(4):309-310. doi: 10.1016/j.jinf.2011.02.006.
- da Costa LB, Ahn H, Montanera W, Ginsberg H. Repeated meningitis as a delayed complication of scoliosis surgery. J Spinal Disord Tech. 2007;20(4):333-336.

- [5] Leblanc W, Heagarty MC. Posttraumatic meningitis due to Hemophilus
- influenzae type Å. J Natl Med Assoc. 1983;75:995–996. 999–1000. Kneen R, Solomon T, Appleton R. The role of lumbar puncture in suspected
- CNS infection a disappearing skill? Arch Dis Child. 2002;87:181–183.

 Zarrouk V, Vassor I, Bert F, et al. Evaluation of the management of postoperative aseptic meningitis. Clin Infect Dis. 2007;44:1555–1559.

 Tunkel AR, Hartman BJ, Kaplan SL, et al. Practice guidelines for the manage-
- ment of bacterial meningitis. Clin Infect Dis. 2004;39:1267–1284. Brouwer MC, Tunkel AR, van de Beek D, et al. diagnosis and antimicrobial
- treatment of acute bacterial meningitis. Clin Microbiol Rev. 2010;23:467-
- van de Beek D, de Gans J, Spanjaard L, et al. Clinical features and prognostic
- factors in adults with bacterial meningitis. N Engl J Med. 2004;351:1849–1859. Huy NT, Thao NT, Diep DT, et al. Cerebrospinal fluid lactate concentration to distinguish bacterial from aseptic meningitis: a systemic review and meta-analysis. Crit Care. 2010;14:R240
- Weisfelt M, van de Beek D, Spanjaard L, et al. Nosocomial bacterial meningitis in adults: a prospective series of 50 cases. J Hosp Infect. 2007;66:71-78.
- Balouch MA, Bajwa RJ, Hassoun A. Successful use of ceftaroline for the treatment of MRSA meningitis secondary to an infectious complica-

- tion of lumbar spine surgery. J Antimicrob Chemother. 2015;70(2):624-625. doi:10.1093/jac/dku392.
- Lin TY, ChenWJ, Hsieh MK, et al. Postoperative meningitis after spinal surgery: a review of 21 cases from 20,178 patients. BMC Infect Dis. 2014;14:220. Twyman RS, Robertson P, Thomas MG. Meningitis complicating spinal
- surgery. Spine (Phila Pa 1976). 1996;21:763-765.
 McClelland S 3rd, Hall WA. Postoperative central nervous system infection: incidence and associated factors in 2,111 neurosurgical procedures. Clin
- Infect Dis. 2007;45(1):55–59. Epub 2007 May 21.

 De Freitas DJ, McCabe JP. Acinetobacter baumanii meningitis: a rare compli-
- cation of incidental durotomy. J Spinal Disord Tech. 2004;17:115–116. Morgan-Hough CV, Jones PW, Eisenstein SM. Primary and revision lumbar discectomy. A 16-year review from one centre. J Bone Joint Surg Br. 2003;85:871-874.
- Couture D, Branch CJ. Spinal pseudomeningoceles and cerebrospinal fluid
- fistulas. Neurosurg Focus. 2003;13:E6. Zhang LM, Ren L, Zhao ZQ, Zhao YR, Zheng YF, Zhou JL. Surgical treatment for bacterial meningitis after spinal surgery: a case report. Medicine (Baltimore). 2017;96(11):e6099. doi:10.1097/MD.0000000000006099.

Author: Taolin Fang

QUESTION 9: What are the early infectious complications after operations on the spine following the use of instrumentation?

RECOMMENDATION: Early infections are traditionally defined as those occurring within a month after surgery, typically becoming evident within two to three weeks of surgery. Recently, the definition has been broadened to include infection within 90 days of surgery.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 60%, Disagree: 20%, Abstain: 20% (Super Majority, Weak Consensus)

RATIONALE

Early infections are traditionally defined as those occurring within a month of surgery, typically becoming evident within two to three weeks of surgery. Recently the definition of early infection has been broadened to include infection within 90 days of surgery [1]. Surgical site infections (SSIs) and wound dehiscence are among the most common complications following spine surgery. It has been reported that the incidence of SSIs after adult spine surgery varies from 2–20% following instrumented procedures [2].

A study based on the American College of Surgeons' National Surgical Quality Improvement Program database reported that in a total of 99,152 spine surgery cases between 2012 and 2014, the overall wound complication rate was 2.2% with superficial SSI, 0.9% with deep SSI, o.8% organ space SSI and o.4% dehiscence: 0.3%. Of all the patients who experienced wound dehiscence, 46% had concomitant SSI. The average postoperative day of occurrence was 14 days with a standard deviation of 9 days (superficial SSI: 16 ± 8 , deep SSI: 13 ± 10 , organ/space SSI: 11 \pm 10, dehiscence: 17 \pm 8) [3].

Similar to other SSIs, early infections after spine surgery may present as pain, fever, erythema, swelling, warmth, tenderness and wound drainage. Local pain may herald the development of infection, particularly when it is escalating in nature. Wound drainage is common for both superficial or deep SSIs and may be present in up to 90% of patients [4].

Early postoperative spinal infections are most frequently due to relatively virulent pathogens such as Staphylococcus aureus, betahemolytic streptococci and aerobic gram-negative bacilli. Staphylococcus aureus is the most common bacteria responsible for early postoperative infection after spinal surgery [5–7]. The majority of the cases are due to methicillin-sensitive Staphylococcus aureus (MSSA), however the incidence of methicillin-resistant Staphylococcus aureus (MRSA) is escalating [8]. The majority of early infections are due to a single pathogen [9]. There has been an increase in the frequency of infections caused by gram-negative bacteria and other organisms such as Pseudomonas aeruginosa, Escherichia coli, Enterobacter and Acinetobacter [10-12].

Utilization of posterior instrumentation is well-recognized as a risk factor for the development of postoperative spinal wound infections. However, this finding is largely based on suboptimal retrospective analyses. Multiple factors increase the rates of infection following instrumented spinal surgery, such as increased wound exposure to air due to longer surgical time, greater soft tissue dissection, increased muscle/skin retraction, greater blood loss and potentially larger dead spaces [13–15].

However, anterior spinal exposures were reportedly correlated with a reduced risk of infection as they typically traverse relatively avascular tissue planes and avoid significant muscle dissection [16–19]. It is yet to be determined whether minimally invasive spine surgery is associated with lower infection rates versus open surgery following the use instrumentation [20–21], although a recent study involving 108,419 procedures reported that the use of a minimally invasive approach was associated with a lower rate of infection for lumbar discectomy (0.4% vs. 1.1%, p < 0.001) and for transforaminal lumbar interbody fusion (1.3% vs. 2.9%, p = 0.005)[22].

- Sebaaly A, Shedid D, Boubez G, Zairi F, Kanhonou M, Yuh SJ, Wang Z. Surgical site infection in spinal metastasis: incidence and risk factors, Spine J. 2018;18(8):1382-1387. Kasliwal MK, Tan LA, Traynelis VC. Infection with spinal instrumentation:
- review of pathogenesis, diagnosis, prevention, and management. Surg Neurol Int. 2013;4(Suppl 5):S392-S403.

657

- Piper KF, Tomlinson SB, Santangelo G, et al. Risk factors for wound compli-[3]
- cations following spine surgery. Surg Neurol Int. 2017;8:269. Pull ter Gunne AF, Mohamed AS, Skolasky RL, van Laarhoven CJ, Cohen DB. The presentation, incidence, etiology, and treatment of surgical site infections after spinal surgery. Spine (Phila Pa 1976). 2010;35:1323–1328. Beiner JM, Grauer J, Kwon BK, Vaccaro AR. Postoperative wound infections
- of the spine. Neurosurg Focus. 2003;15:E14. Bemer P, Corvec S, Tariel S, Asseray N, Boutoille D, Langlois C, et al. Significance of Propionibacterium acnes-positive samples in spinal instrumentation. Spine (Phila Pa 1976). 2008;33:E971-E976.
- Blam OG, Vaccaro AR, Vanichkachorn JS, Albert TJ, Hilibrand AS, Minnich JM, et al. Risk factors for surgical site infection in the patient with spinal
- injury. Spine (Phila Pa 1976). 2003;28:1475-1480.
 Kim JI, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infection after instrumented spinal fusion. J Spinal Disord Tech. 2010;23:258-265.
 Rohmiller MT, Akbarnia BA, Raiszadeh K, Raiszadeh K, Canale S. Closed
- suction irrigation for the treatment of postoperative wound infections following posterior spinal fusion and instrumentation. Spine. 2010;35:642-
- Fang A, Hu SS, Endres N, Bradford DS. Risk factors for infection after spinal
- surgery. Spine (Phila Pa 1976). 2005;30:1460–1465. Ido K, Shimizu K, Nakayama Y, Shikata J, Matsushita M, Nakamura T. Suction/irrigation for deep wound infection after spinal instrumentation:
- a case study. Eur Spine J. 1996;5:345–349.
 Jones GA, Butler J, Lieberman I, Schlenk R. Negative-pressure wound therapy in the treatment of complex postoperative spinal wound infections: complications and lessons learned using vacuum-assisted closure. J Neurosurg Spine. 2007;6:407–411. Ho C, Sucato DJ, Richards BS. Risk factors for the development of
- delayed infections following posterior spinal fusion and instrumen-

- tation in adolescent idiopathic scoliosis patients. Spine (Phila Pa 1976). 2007;32:2272-2277
- Pull ter Gunne AF, Cohen DB. Incidence, prevalence, and analysis of risk factors for surgical site infection following adult spinal surgery. Spine
- (Phila Pa 1976). 2009;34:1422–1428. Rathjen K, Wood M, McClung A, Vest Z. Clinical and radiographic results after implant removal in idiopathic scoliosis. Spine (Phila Pa 1976). 2007;32:2184-2188
- Ghanayem AJ, Zdeblick TA. Cervical spine infections. Orthop Clin North
- Am. 1996;27:53-67. Maragakis LL, Cosgrove SE, Martinez EA, Tucker MG, Cohen DB, Perl TM. Intraoperative fraction of inspired oxygen is a modifiable risk factor for
- surgical site infection after spinal surgery. Anesthesiology. 2009;110:556–562. Olsen MA, Mayfield J, Lauryssen C, Polish LB, Jones M, Vest J, et al. Risk factors for surgical site infection in spinal surgery. J Neurosurg. 2003;98(2
- Suppl):149-155.
 Wimmer C, Gluch H, Franzreb M, Ogon M. Predisposing factors for infection in spine surgery: a survey of 850 spinal procedures. J Spinal Disord. 1998;11:124-128.
- O'Toole JE, Eichholz KM, Fessler RG. Surgical site infection rates after mini-
- mally invasive spinal surgery. J Neurosurg Spine. 2009;11:471–476. Parker SL, Adogwa O, Witham TF, Aaronson OS, Cheng J, McGirt MJ. Post-operative infection after minimally invasive versus open transforaminal lumbar interbody fusion (TLIF): literature review and cost analysis. Minim
- Invasive Neurosurg. 2011;54:33–37. Smith JS, Shaffrey CI, Sansur CA, Berven SH, Fu KM, Broadstone PA, et al. Rates of infection after spine surgery based on 108,419 procedures: a report from the Scoliosis Research Society Morbidity and Mortality Committee. Spine (Phila Pa 1976). 2011;36:556–563.

2.2. DIAGNOSIS: BIOMARKERS

Author: Maja Babic

QUESTION 1: Are there any diagnostic tools that are useful for early surgical site infection (SSI) detection following spinal surgery? Does this differ whether or not there was instrumentation?

RECOMMENDATION: C-reactive protein (CRP) can be used to diagnose early SSI following spinal surgery.

A failure of CRP to decline or a second rise on postoperative days four to seven is a sensitive marker for infection following spine surgery, including both instrumented and non-instrumented spine surgery.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 86%, Disagree: 7%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

In a prospective study of 73 consecutive patients undergoing spinal decompression with and without instrumentation, inflammatory markers were assessed. They showed that following uncomplicated spinal surgery, CRP levels rise sharply, peaking on the second postoperative day [1]. Peak CRP values after instrumented lumbar surgery are significantly higher than those after non-instrumented spine surgery, but decline with the same half-life [1]. CRP was superior to erythrocyte sedimentation rate (ESR) in early detection of infections after cervical spine surgery, as shown in a prospective study of 51 cases [2]. In another large, prospective trial including 400 elective discectomy cases, CRP was shown to be a reliable, simple and economical screening test for infectious complications after lumbar microdiscectomy, superior to classical laboratory parameters. The sensitivity of serial CRP testing was calculated to be 100% with 95.8% specificity. ESR and white blood cell measurements fail to reach distinctive significance in diagnosing early SSI [3].

- Takahashi J, Ebara S, Kamimura M, et al. Early-phase enhanced inflamma-
- Rosahl SK, Gharabaghi A, Zink PM, Samii M. Monitoring of blood parameters following anterior cervical fusion. J Neurosurg Spine. 2001;26:1698–1704.

 Meyer B, Schaller K, Rohde V, Hassler W. The C-reactive protein for detec-
- tion for early infections after lumbar microdiscectomy. Acta Neurochir. 1995;136:145-150.

Author: Maja Babic

QUESTION 2: When do common blood biomarkers such as C-reactive protein (CRP), erythrocyte sedimentation rate (ESR) or Procalcitonin normalize after spine surgery?

RECOMMENDATION: Following spinal surgery with or without instrumentation, CRP values peak on days 2-3 postoperatively and normalize within 14 days. ESR also normalizes within 14 days.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 50%, Disagree: 29%, Abstain: 21% (NO Consensus)

RATIONALE

Multiple prospective studies suggest that CRP values peak within 2-3 days postoperatively (peak levels depend on extent of surgery, levels involved, etc.) and decrease back to baseline within 14 days. A rapid decline of CRP postoperatively is interrupted if postoperative infection sets in and a secondary rise occurs [1,2]. Prospective studies have shown that ESR peaks by day four following spinal surgery and in the majority of cases normalizes by two weeks postoperatively [3]. However, monitoring of CRP level was found to be superior to that of ESR for early detection of infections after cervical spine surgery in a series of 51 cases of anterior cervical fusion [4]. A second rise of CRP and ESR or failure to decline is an indicator of potential surgical site infection [5,6]. Limited data is available on the value of Procalcitonin [7].

REFERENCES

- Thelander U, Larsson S. Quantitation of C-reactive protein levels and eryth-
- rocyte sedimentation rate after spinal surgery. Spine. 1992;17:400–404. Takahashi J, Ebara S, Kamimura M, Kinoshita T, Itoh H, Yuzawa Y, et al. Early-phase enhanced inflammatory reaction after spinal instrumentation
- surgery. Spine. 2001;26:1698–1704. Jönsson B, Söderholm R, Strömqvist B. Erythrocyte sedimentation rate after
- lumbar spine surgery. Spine. 1991;16:1049-1050.
 Rosahl SK, Gharabaghi A, Zink PM, Samii M. Monitoring of blood parameters following anterior cervical fusion. J Neurosurg. 2000;92:169-174.
 Kong CG, Kim YY, Park JB. Postoperative changes of early-phase inflamma-
- tory indices after uncomplicated anterior cervical discectomy and fusion using allograft and demineralised bone matrix. Int Orthop. 2012;36:2293-
- tusing anografi and definitefatised bone matrix. Int Orthop. 2012;36:2293-2297. doi:10.1007/s00264-012-1645-z.
 Mok JM, Pekmezci M, Piper SL, Boyd E, Berven SH, Burch S, et al. Use of C-reactive protein after spinal surgery: comparison with erythrocyte sedimentation rate as predictor of early postoperative infectious complications. Spine. 2008;33:415-421. doi:10.1097/BRS.0b013e318163fgee.

 Syvänen J, Peltola V, Pajulo O, Ruuskanen O, Mertsola J, Helenius I. Normal behavior of placement prodiction in adolescents undergoing surgery for
- behavior of plasma procalcitonin in adolescents undergoing surgery for scoliosis. Scand J Surg. 2014;103:60-65. doi:10.1177/1457496913504910.

Author: Maja Babic

QUESTION 3: Is there a role for the use of serum biomarker for the diagnosis of spinal surgical site infection (SSI)?

RECOMMENDATION: Yes, C-reactive protein (CRP) is a predictable, reliable and economical screening tool for early infectious complications following spine surgery. Erythrocyte sedimentation rate and white blood cell count have nonspecific kinetics that are less helpful in identifying early SSI.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 67%, Disagree: 25%, Abstain: 8% (Super Majority, Weak Consensus)

RATIONALE

In a prospective study involving 348 patients who underwent decompression laminectomy, postoperative CRP was helpful in detecting early infectious complications following surgery. As a predictor for early wound infection, the sensitivity, specificity, positive predictive value and negative predictive value for abnormal CRP responses were calculated as 100%, 96.8%, 31.3% and 100%, respectively. Close observation of the surgical site is recommended in patients with abnormal CRP values at day five or seven postoperatively, namely for failure to decline or a secondary rise [1].

Of 149 patients undergoing elective spine surgery, 20 developed infectious SSI complications. Postoperative CRP kinetics were predictable and indicative of early infection where a secondary rise or lack of CRP decrease had a sensitivity, specificity, positive predictive value and negative predictive value of 82%, 48%, 41%, and 86% for infectious complications, respectively [2].

Out of 400 patients undergoing lumbar micro-discectomy over a 15-month period, 9 developed infectious complications related to surgery. CRP values were shown to be a reliable and economic screening tool in identifying the patients at risk with a sensitivity for serial CRP testing (day one and five postoperatively) calculated as 100% with a specificity of 95.8% [3].

REFERENCES

Kang BU, Lee SH, Ahn Y, Choi WC, Choi YG. Surgical site infection in spinal surgery: detection and management based on serial

- C-reactive protein measurements. J Neurosurg Spine. 2010;13:158–164. doi:10.3171/2010.3.SPINE09403.
- [2] Mok JM, Pekmezci M, Piper SL, Boyd E, Berven SH, Burch S, et al. Use of C-reactive protein after spinal surgery: comparison with erythrocyte sedi-
- mentation rate as predictor of early postoperative infectious complications. Spine. 2008;33:415-421. doi:10.1097/BRS.ob013e318163f9ee.
- Meyer B, Schaller K, Rohde V, Hassler W. The C-reactive protein for detection of early infections after lumbar microdiscectomy. Acta Neurochir (Wien). 1995;136:145-150.



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QUESTION 4: Is there a role for molecular techniques such as polymerase chain reaction (PCR) or next-generation sequencing (NGS) for the diagnosis of spinal surgery infection? If so, in which group of patients should this be done?

RECOMMENDATION: It is reasonable to selectively incorporate these diagnostic modalities as an adjunct to standard methodologies where there is a history or high pre-test probability for culture negative infection.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 71%, Disagree: 14%, Abstain: 15% (Super Majority, Strong Consensus)

RATIONALE

Successful management of periprosthetic joint infections (PJI) is significantly enhanced with a prompt and accurate microbiological diagnosis. Conventional culture methods for diagnosis of PJI can be compromised and complicated by early antibiotic treatment, heterogeneity of surgical sampling, fastidious microorganisms difficult to grow in culture and non-planktonic pathogens utilizing biofilms. Therefore, modern molecular microbiologic methods have naturally been seen as very promising for increasing diagnostic yield in these circumstances. Technologies that have more recently been applied to PJI generally include ribosomal RNA sequencing, species-specific and multiplex PCR and matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS).

Specifically, with respect to spinal and vertebral infections, these varied technologies have demonstrated success in leading to an etiologic diagnosis. These methods have been used to identify a variety of pathogens, including *Staphylococcus spp.* [1–3], *Streptococcus spp* [3,4], *Enterococcus spp.* [4], Enterobacteriaceae [3–5], *Brucella spp.* [6], *Mycobacterium spp.* [2], atypical bacteria (*T. whipplei*) [7], *Mycoplasma spp.*) [8], anaerobes (*Clostridium spp.*) [3], *Fusobacterium spp.*) [4,9] and fungi (*Aspergillus spp.*) [10].

By far, the most experience with these techniques for spinal infections is in the diagnosis of Pott's disease (*Mycobacterium tuberculosis*) [2,6,11–15]. These reports generally demonstrate a high sensitivity and specificity of PCR modalities, though many of these studies have been completed in tuberculosis endemic geographic areas with likely higher inoculum infections and a well-defined pretest probability.

False positive results from dead or colonizing/contaminating bacteria is a concern with these tests, and studies evaluating the appropriate number of samples to optimize sensitivity and specificity specific to these molecular methods are limited and not specific to spinal infections [16]. Another important concern with molecular techniques for PJI diagnostics is that they do not commonly allow for susceptibility testing to appropriately target antimicrobial therapy. Certain resistance mechanisms, such as methicillin resistance in *S. aureus* [1,17,18] or rifampin resistance in *M. tuberculosis* [12], are reliably expressed if genetically detected. This is not the norm, however, as resistance expression is generally a complex phenotype determined by multiple factors. Care should be taken not to overly rely on non-susceptibility-generating techniques, as they can just as easily

lead to long courses of overly-broad therapy, as can no etiologic diagnosis at all, undermining patient safety and important principles of antimicrobial stewardship. In addition, it has been noted that utilizing molecular methods as an adjunct to and in combination with standard culture methodologies often serves to improve overall diagnostic yield [3].

A few studies have attempted to establish test sensitivity and specificity data when compared to routine culture for bone and joint specimens in general [4,15,19–23], however these efforts are limited by lack of a true gold standard diagnostic method for comparison, the variety of testing methodologies clinically employed and non-standardized clinical criteria for utilization of these methods. Predictably, results vary widely, with sensitivities reported between 50-92% and specificities between 65-94% [20]. No studies investigating sensitivity and specificity of these techniques specific only to spinal postsurgical infections have yet been reported. Therefore, an evidencebased evaluation of the appropriate clinical criteria for utilization of these techniques in spinal surgery patients is not currently possible. One study proposed a strategy for routine collection and potential use of molecular diagnostics in PJI [24]. There is no data investigating the cost effectiveness for any diagnostic schema incorporating molecular methods, however given their positive proof-of-concept and the significant clinical impact of spinal post-surgical infections, it seems reasonable to selectively incorporate the use of molecular methods into situations where there is a high pre-test probability for indolent or culture-negative infection as further studies are done to standardize their use.

- [1] Tsuru A, Setoguchi T, Kawabata N, Hirotsu M, Yamamoto T, Nagano S, et al. Enrichment of bacteria samples by centrifugation improves the diagnosis of orthopaedics-related infections via real-time PCR amplification of the bacterial methicillin-resistance gene. BMC Res Notes. 2015;8:288. doi:10.1186/s13104-015-1180-2.
- [2] Sheikh AF, Khosravi AD, Goodarzi H, Nashibi R, Teimouri A, Motamedfar A, et al. Pathogen identification in suspected cases of pyogenic spondylodiscitis. Front Cell Infect Microbiol. 2017;7:60. doi:10.3389/frip.horg.co.60
- fcimb.2017.00060.

 [3] Fuursted K, Arpi M, Lindblad BE, Pedersen LN. Broad-range PCR as a supplement to culture for detection of bacterial pathogens in patients with a clinically diagnosed spinal infection. Scand J Infect Dis. 2008;40:772–777. doi:10.1080/00365540802119994.

- [4] Fihman V, Hannouche D, Bousson V, Bardin T, Lioté F, Raskine L, et al. Improved diagnosis specificity in bone and joint infections using molecular techniques. J Infect. 2007;55:510–517. doi:10.1016/j.jinf.2007.09.001.
- [5] Shibata S, Tanizaƙi R, Watanabe K, Makabe K, Shoda N, Kutsuna S, et al. Escherichia coli vertebral osteomyelitis diagnosed according to broad-range 16S rRNA gene polymerase chain reaction (PCR). Intern Med. 2015;54:3237–3240. doi:10.2169/internalmedicine.54.5066.
 [6] Colmenero JD, Morata P, Ruiz-Mesa JD, Bautista D, Bermúdez P, Bravo MJ, et
- [6] Colmenero JD, Morata P, Ruiz-Mesa JD, Bautista D, Bermúdez P, Bravo MJ, et al. Multiplex real-time polymerase chain reaction: a practical approach for rapid diagnosis of tuberculous and brucellar vertebral osteomyelitis. Spine. 2010;35;E1392-E1396. doi:10.1097/BRS.ob013e3181e8eeaf.
- [7] Altwegg M, Fleisch-Marx A, Goldenberger D, Hailemariam S, Schaffner A, Kissling R. Spondylodiscitis caused by Tropheryma whippelii. Schweiz Med Wochenschr. 1006;136:1405-1406.
- Wochenschr. 1996;126:1495–1499.

 [8] Flouzat-Lachaniette C-H, Guidon J, Allain J, Poignard A. An uncommon case of Mycoplasma hominis infection after total disc replacement. Eur Spine J. 2013;22 Suppl 3:S394–S398. doi:10.1007/s00586-012-2511-9.
- [9] Sanmillán JL, Pelegrín I, Rodríguez D, Ardanuy C, Cabellos C. Primary lumbar epidural abscess without spondylodiscitis caused by Fusobacterium necrophorum diagnosed by 16S rRNA PCR. Anaerobe. 2013;23:45-47. doi:10.1016/j.anaerobe.2013.06.014.
- doi:10.1016/j.anaerobe.2013.06.014.
 [10] Dayan I., Sprecher H, Hananni A, Rosenbaum H, Milloul V, Oren I. Aspergillus vertebral osteomyelitis in chronic leukocyte leukemia patient diagnosed by a novel panfungal polymerase chain reaction method. Spine J. 2007;7:615-617. doi:10.1016/j.spinee.2006.08.005.
- [11] Sharma K, Meena RK, Aggarwal A, Chhabra R. Multiplex PCR as a novel method in the diagnosis of spinal tuberculosis-a pilot study. Acta Neuro-chir (Wien). 2017;159:503-507. doi:10.1007/s00701-016-3065-0.
- chir (Wien). 2017;159:503-507. doi:10.1007/s00701-016-3065-0.

 [12] Held M, Laubscher M, Zar HJ, Dunn RN. GeneXpert polymerase chain reaction for spinal tuberculosis: an accurate and rapid diagnostic test. Bone Joint J. 2014;96-B:1366-1369. doi:10.1302/0301-620X.96B10.34048.

 [13] Pandey V, Chawla K, Acharya K, Rao S, Rao S. The role of polymerase chain
- [13] Pandéy V, Chawla K, Acharya K, Rao S, Rao S. The role of polymerase chain reaction in the management of osteoarticular tuberculosis. Int Orthop. 2009;33:801–805. doi:10.1007/s00264-007-0485-8.
- [14] Sun Y, Zhang Y, Lu Z. [Clinical study of polymerase chain reaction technique in the diagnosis of bone tuberculosis]. Zhonghua Jie He He Hu Xi Za Zhi. 1997;20:145–148.
- [15] Van der Spoel van Dijk A, MCleod A, Botha PL, Shipley JA, Kapnoudhis MA, Beukes CA. The diagnosis of skeletal tuberculosis by polymerase chain reaction. Cent Afr J Med. 2000;46:144–149.

- [16] Marín M, Garcia-Lechuz JM, Alonso P, Villanueva M, Alcalá L, Gimeno M, et al. Role of universal 16S rRNA gene PCR and sequencing in diagnosis of prosthetic joint infection. J Clin Microbiol. 2012;50:583–589. doi:10.1128/JCM.00170-11.
- [17] Choe H, Aota Y, Kobayashi N, Nakamura Y, Wakayama Y, Inaba Y, et al. Rapid sensitive molecular diagnosis of pyogenic spinal infections using methicillin-resistant Staphylococcus-specific polymerase chain reaction and 16S ribosomal RNA gene-based universal polymerase chain reaction. Spine J. 2014;14:255-262. doi:10.1016/j.spinee.2013.10.044.
- [18] Dubouix-Bourandy A, de Ladoucette A, Pietri V, Mehdi N, Benzaquen D, Guinand R, et al. Direct detection of Staphylococcus osteoarticular infections by use of Xpert MRSA/SA SSTI real-time PCR. J Clin Microbiol.
- tions by use of Xpert MRSA/SA SSTI real-time PCR. J Clin Microbiol. 2011;49:4225–4230. doi:10.1128/JCM.00334-11.

 [19] Borde JP, Häcker GA, Guschl S, Serr A, Danner T, Hübner J, et al. Diagnosis of prosthetic joint infections using UMD-Universal Kit and the automated multiplex-PCR University of 160 ITI(®) cartridge system: a pilot study. Infection. 2015;42:551–560. doi:10.1007/s15010-015-0706-4
- 2015;43:551–560. doi:10.1007/s15010-015-0796-4.

 [20] Malandain D, Bémer P, Leroy AG, Léger J, Plouzeau C, Valentin AS, et al. Assessment of the automated multiplex-PCR Unyvero i60 ITI® cartridge system to diagnose prosthetic joint infection: a multicentre study. Clin Microbiol Infect. 2018;24:83-e1–83.e6. doi:10.1016/j.cmj.2017.05.017.
- Microbiol Infect. 2018;24:83.e1-e83.e6. doi:10.1016/j.cmi.2017.05.017.

 [21] Bémer P, Plouzeau C, Tande D, Léger J, Giraudeau B, Valentin AS, et al. Evaluation of 16S rRNA gene PCR sensitivity and specificity for diagnosis of prosthetic joint infection: a prospective multicenter cross-sectional study. J Clin Microbiol. 2014;52:3282-3280. doi:10.1128/JCM.01450-14
- Microbiol. 2014;52:3583-3589. doi:10.1128/JCM.01459-14.

 [22] Grif K, Heller I, Prodinger WM, Lechleitner K, Lass-Flörl C, Orth D. Improvement of detection of bacterial pathogens in normally sterile body sites with a focus on orthopedic samples by use of a commercial 16S rRNA broad-range PCR and sequence analysis. J Clin Microbiol. 2012;50:2250-2254. doi:10.1128/JCM.00362-12.
- [23] Fenollar F, Roux V, Stein A, Drancourt M, Raoult D. Analysis of 525 samples to determine the usefulness of PCR amplification and sequencing of the 16S rRNA gene for diagnosis of bone and joint infections. J Clin Microbiol. 2006;44:1018–1028. doi:10.1128/JCM.44.3.1018-1028.2006.
- [24] Lévy P-Y, Fenollar F. The role of molecular diagnostics in implant-associated bone and joint infection. Clin Microbiol Infect. 2012;18:1168–1175. doi:10.1111/1469-0691.12020.

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QUESTION 5: For which investigations should samples obtained by image-guided biopsy be sent?

RECOMMENDATION: A priority should be placed on obtaining bacterial cultures and pathohistology. In the appropriate epidemiological setting, mycobacterial, fungal and brucellar cultures can be considered.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RESPONSE

There is limited data available in the literature to help establish clear evidence-based parameters for treatment. However, there are society-based clinical guidelines such as the 2015 Infectious Diseases Society of America (IDSA) clinical practice guidelines for the diagnosis and treatment of native vertebral osteomyelitis in adults, which provide assistance in decision-making. Highlights from this statement recommend the acquisition of image-guided aspiration biopsy in patients with suspected vertebral osteomyelitis when a microbiologic diagnosis for a known associated organism has not been established by blood cultures or serologic tests. Further, they recommend for the addition of fungal, mycobacterial or brucellar cultures on image-guided biopsy and aspiration specimens in patients with suspected vertebral osteomyelitis if epidemiologic, host risk factors or characteristic radiologic clues are present, or if testing to appropriately stored bacterial specimens reveal no growth [1].

There is some data to suggest that standard samples should be sent for both microbiology and pathohistology. Pathologic evaluation is meaningful, particularly in culture negative cases where the presence of leukocytes can indicate pyogenic osteomyelitis, or visualization of granulomas can suggest mycobacterial infection or brucellosis [2]. Pathology can also support ruling out diagnoses like ankylosing spondylitis, hemodialysis-associated spondyloarthropathy or neuropathic Charcot joint deformities [3]. Furthermore, crystal deposits can aid in the diagnosis of pseudogout [4].

- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native vertebral osteomyelitis in adults. Clin Infect Dis. 2015;61:e26-e46. doi:10.1093/cid/civ482.
- adults. Clin Infect Dis. 2015;61:e26–e46. doi:10.1093/cid/civ482.

 [2] Zimmerli W. Clinical practice. Vertebral osteomyelitis. N Engl J Med. 2010;362:1022–1029. doi:10.1056/NEJMcp0910753.

- [3] Hong SH, Choi JY, Lee JW, Kim NR, Choi JA, Kang HS. MR imaging assessment of the spine: infection or an imitation? Radiographics. 2009;29:599–612. doi:10.1148/rg.292085137.
- [4] Godfrin-Valnet M, Godfrin G, Godard J, Prati C, Toussirot E, Michel F, et al. Eighteen cases of crowned dens syndrome: presentation and diagnosis. Neurochirurgie. 2013;59:115–120. doi:10.1016/j.neuchi.2013.03.003.

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QUESTION 6: How many intraoperative tissue samples should be sent for culture in suspected spinal surgery infection?

RECOMMENDATION: With the currently-available evidence, it is recommended that at least three to five tissue samples be sent for culture in cases of suspected spinal infection. In the setting of instrumentation, we recommend additional techniques, such as vortexing and sonification, to increase the yield of culture samples.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 80%, Disagree: 13%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Turnbull described surgical site infections (SSIs) in spinal surgery in 1953, noting three cases of deep infection of the disc after discectomy as well as significant morbidity that followed [1]. While clinically these cases presented as infection, Turnbull reported them as "presumed" infection because culture of the causative organism was not obtained. Since his work, the incidence of SSI following spine surgery has been studied extensively, with reported incidences ranging from 0.2-15%, varying widely based on underlying pathology and procedure type, with revision procedure, fusion, implantation, and traumatic injury carrying the greatest risk [2–6]. The most common causative culprits are Staphylococcus species, including methicillin-resistant S. aureus (MRSA) [3,6-9], although less virulent organisms such as Propionibacterium acnes can also occur, particularly in revision cases without a definitive preoperative diagnosis of infection [10–12]. Prior to obtaining intraoperative cultures, some suggest computed tomography-guided aspiration, although this practice has been shown to have low sensitivity [13,14].

The evidence for the optimal number of specimens to obtain in cases of suspected spinal infection is sparse. In their study of patients undergoing Cotrel-Dubousset instrumentation, Bemer et al. evaluated cases of *Cutibacterium acnes (C. acnes)* infection, noting that in earlier years of the study one to two samples for culture were obtained, whereas later in their series they had increased that number to four to six samples. Based on their experience and the difficulty in diagnosis of *C. acnes*, these studies recommend that at least four deep tissue samples be taken to facilitate interpretation of the cultures [11].

In the setting of implantation, one major challenge is that peri-implant cultures may not be accurate and it can be difficult for biofilm cultures to grow [15]. In their study of removed spinal implants in the setting of infection, Sampedro et al. report using a technique of vortexing and sonification followed by culturing, which resulted in significantly increased sensitivity compared to simply taking two to nine peri-implant tissue samples [12]. Finally, in a study assessing specimens taken from orthopaedic device revision surgery (5.1% spine cases), the standard procedure was to obtain three to six (mean: four) samples per case, including specimens from the inflammatory membrane around the implant, joint capsule (as applicable) and any macroscopically suspect tissues [16]. In this study, a threshold of at least three positive samples with identical microorganisms was used for diagnosis. The authors

note that this definition is strict compared to other studies that use two identical culture-positive specimens for diagnosis and report that their findings would not have differed had they used a lower threshold of two.

It is important to remember that positive cultures may not represent true infection and should be interpreted in the overall context of the individual patient and clinical picture. Gelalis et al. studied bacterial contamination during simple and complex spinal procedures in 40 patients, taking culture swabs during each case, first from the sterile transparent sheet over the operative site at the start of the case, followed by hourly samples from the surgical wound. The authors reported that none of the patients with positive cultures developed clinical signs of infection or required antibiotics, whereas three patients with negative cultures developed postoperative infection [17].

Though there is little guidance in the spine literature, the data in arthroplasty may help to guide future practices. In a study looking at revision hip and knee arthroplasty, Atkins et al. found that the presence of three or more culture-positive specimens was highly predictive of infection (likelihood ratio, 169; sensitivity, 66%; specificity, 99.6%), whereas a single culture-positive specimen was found to have low diagnostic value (likelihood ratio, 0.7; post-test probability of infection, 10.6%) [15]. In their study, the authors determined that five or six samples are required to produce excellent sensitivity and specificity. Similarly, in a study of periprosthetic joint infection caused by MRSA, Parvizi et al. took five cultures in each case [18]. In accordance with the evidence, the Workgroup of the Musculoskeletal Infection Society recommends that three to five culture samples be taken and incubated in an aerobic and anaerobic environment [19].

There is little evidence regarding the optimal number of samples to obtain in the setting of suspected spinal surgery infection. Given the limited data that is available in the spine literature, we conclude that taking at least three to five tissue samples represents current best practice. In the setting of instrumentation, we recommend additional techniques, such as vortexing and sonification, to increase the yield of culture samples.

REFERENCES

[1] Turnbull F. Postoperative inflammatory disease of lumbar discs. J Neurosurg. 1953;10:469-473. doi:10.3171/jns.1953.10.5.0469.

- [2] Silber JS, Anderson DG, Vaccaro AR, Anderson PA, McCormick P, NASS. Management of postprocedural discitis. Spine J. 2002;2:279–287.
- [3] Chen SH, Lee CH, Huang KC, Hsieh PH, Tsai SY. Postoperative wound infection after posterior spinal instrumentation: analysis of long-term treatment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014-3636-9.
- ment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014-3636-9.

 [4] Beiner JM, Grauer J, Kwon BK, Vaccaro AR. Postoperative wound infections of the spine. Neurosurg Focus. 2003;15:E14.
- [5] Smith JS, Shaffrey CJ, Sansur CA, Berven SH, Fu K-MG, Broadstone PA, et al. Rates of infection after spine surgery based on 108,419 procedures: a report from the Scoliosis Research Society Morbidity and Mortality Committee. Spine. 2011;36:556–563. doi:10.1097/BRS.0b013e3181eadd41.
- [6] Patel H, Khoury H, Girgenti D, Welner S, Yu H. Burden of surgical site infections associated with select spine operations and involvement of Staphylococcus aureus. Surg Infect (Larchmt). 2017;18:461-473. doi:10.1089/ SUIZ-2016.186.
- [7] Ee WWG, Lau WLJ, Yeo W, Von Bing Y, Yue WM. Does minimally invasive surgery have a lower risk of surgical site infections compared with open spinal surgery? Clin Orthop Relat Res. 2014;472:1718–1724. doi:10.1007/s11999-012-2158-5
- [8] Koutsoumbelis S, Hughes AP, Girardi FP, Cammisa FP, Finerty EA, Nguyen JT, et al. Risk factors for postoperative infection following posterior lumbar instrumented arthrodesis. J Bone Joint Surg Am. 2011;93:1627–1633. doi:10.2106/JBIS.l.00039.
- doi:10.2106/JBJS.J.00039.

 [9] Meredith DS, Kepler CK, Huang RC, Brause BD, Boachie-Adjei O. Postoperative infections of the lumbar spine: presentation and management. Int Orthop. 2012;36:439-444. doi:10.1007/s00264-011-1427-z.
- [10] Shifflett GD, Bjerke-Kroll BT, Nwachukwu BU, Kueper J, Burket J, Sama AA, et al. Microbiologic profile of infections in presumed aseptic revision spine surgery. Fur Spine L 2016;25:2002-2007. doi:10.1007/s00586-016-4520-8
- surgery. Eur Spine J. 2016;25:3902–3907. doi:10.1007/s00586-016-4539-8. [11] Bémer P, Corvec S, Tariel S, Asseray N, Boutoille D, Langlois C, et al. Significance of Propionibacterium acnes-positive samples in spinal instrumentation. Spine. 2008;33:E971–E976. doi:10.1097/BRS.0b013e31818e28dc.

- [12] Sampedro MF, Huddleston PM, Piper KE, Karau MJ, Dekutoski MB, Yaszemski MJ, et al. A biofilm approach to detect bacteria on removed spinal implants. Spine. 2010;35:1218–1224. doi:10.1097/BRS.0b013e3181c3b2f3.
- [13] Jo JE, Miller AO, Cohn MR, Nemani VM, Schneider R, Lebi DR. Evaluating the diagnostic yield of computed tomography-guided aspirations in suspected post-operative spine infections. HSS J. 2016;12:119–124. doi:10.1007/s11420-016-9490-z.
- [14] Enoch DA, Cargill JS, Laing R, Herbert S, Corrah TW, Brown NM. Value of CT-guided biopsy in the diagnosis of septic discitis. J Clin Pathol. 2008;61:750–753. doi:10.1136/jcp.2007.054296.
- 2008;61:750–753. doi:10.1136/jcp.2007.054296.

 [15] Atkins BL, Athanasou N, Deeks JJ, Crook DW, Simpson H, Peto TE, et al. Prospective evaluation of criteria for microbiological diagnosis of prosthetic-joint infection at revision arthroplasty. The OSIRIS Collaborative Study Group. J Clin Microbiol. 1998;36:2932–2939.

 [16] Schwotzer N, Wahl P, Fracheboud D, Gautier E, Chuard C. Optimal culture
- [16] Schwotzer N, Wahl P, Fracheboud D, Gautier E, Chuard C. Optimal culture incubation time in orthopedic device-associated infections: a retrospective analysis of prolonged 14-day incubation. J Clin Microbiol. 2014;52:61-66. doi:10.1128/JCM.01766-13.
- [17] Gelalis ID, Arnaoutoglou CM, Politis AN, Batzaleksis NA, Katonis PG, Xenakis TA. Bacterial wound contamination during simple and complex spinal procedures. A prospective clinical study. Spine J. 2011;11:1042–1048. doi:10.1016/j.spinee.2011.10.015.
 [18] Parvizi J, Azzam K, Ghanem E, Austin MS, Rothman RH. Periprosthetic infections.
- [18] Parvizi J, Azzam K, Ghanem E, Austin MS, Rothman RH. Periprosthetic infection due to resistant staphylococci: serious problems on the horizon. Clin Orthop Relat Res. 2009;467:1732–1739. doi:10.1007/s11999-009-0857-z.
- [19] Parvizi J, Zmistowski B, Berbari EF, Bauer TW, Springer BD, Della Valle CJ, et al. New definition for periprosthetic joint infection: from the Workgroup of the Musculoskeletal Infection Society. Clin Orthop Relat Res. 2011;469:2992– 2994. doi:10.1007/s11999-011-2102-9.

2.3. DIAGNOSIS: IMAGING

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QUESTION 1: What is the optimal mode of imaging in the diagnosis of spine infections? If magnetic resonance imaging (MRI) is contraindicated, what imaging modality should be used?

RECOMMENDATION: MRI remains the gold standard for the diagnosis of spinal infection, with sensitivity and specificity above 90%. In the presence of MRI contraindications, consider a combination of modalities, such as computed tomography (CT), positron emission tomography-CT (PET-CT), and single photon emission CT (SPECT)+67Gallium or Gallium-67.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Plain radiography should be the initial exam performed for all patients with non-specific spine or back complaints. In patients with spinal infections, early radiographic findings will occur two weeks to three months after the onset of symptoms. Plain radiographic findings characteristic of a spinal infection include disc space narrowing, end plate irregularity, loss of end plate contour, subchondral defects and/or hypertrophic or sclerotic bone formation. Disc space narrowing has been reported as the most consistent plain radiographic finding occurring in 74% of cases [1]. Late plain radiographic findings include vertebral body collapse, pathologic fractures, segmental kyphotic collapse and/or bony ankylosis. Plain radiography has reported sensitivity of 82% and specificity of 57 to 59% in subjects with pyogenic spondylodiscitis [2,3]. While this modality may not provide the highest level of diagnostic quality, it can give clinicians an understanding of global and focal alignment,

deformities associated with infectious processes and mechanical stability [4]. Plain radiographs may also be used for post-treatment surveillance and/or monitoring for potential late deformity or instability associated with these diseases.

CT is an advanced imaging technique that can be utilized for diagnosing spinal infections. It provides higher resolution and multiplanar imaging of the bony architecture. CT findings characteristic of spinal infections can include cystic bony changes, gas within vertebral discs, endplate osteolysis surrounding the vertebral disc and/or paravertebral soft tissue swelling or abscess formation [5–7]. The addition of contrast media during computed tomography can help better delineate the edges of paravertebral abscesses and edematous musculature [5–7]. In cases with neurological deficits or new onset radiculopathy, post-myelogram CT scan can provide excellent detail of the spinal canal and poten-

tial epidural and/or subdural abscesses [8]. In cases where myelogram is performed, it is recommended to analyze the cerebral spinal fluid to rule out meningitis [9]. SPECT is a scintigraphic CT modality that has increased bone contrast resolution, and when combined with technetium or gallium, has high sensitivity and diagnostic accuracy for spinal infections. SPECT with gallium has been shown to be superior to SPECT + technetium and is now the recommended imaging modality for patients with MRI contraindications [10].

MRI remains the gold standard for early and accurate diagnosis of spinal infections [11–20]. MRI has a reported sensitivity of 96%, specificity of 93% and diagnostic accuracy of 94% [18]. MRI has higher accuracy for differentiating degenerative and neoplastic conditions from infections in patients presenting with severe back pain of unknown etiology [11]. While MRI may provide the most detailed information for diagnosing possible infections, it does not reduce the need for tissue biopsy for histological analysis. T1-weighted and T2-weighted sequences should be obtained. The most common MRI findings consistent with spinal infections show decreased vertebral body intensity with poor differentiation between the disc and body on T1-weighted images and increased disc space intensity with marked decreased vertebral body intensity on T2-weighted images [16,18,20]. Utilizing gadolinium contrast can enhance MRI ability to detect and delineate epidural abscesses [21]. All publications consider MRI the gold standard imaging modality for spinal infections and recommend it should be used in all patients without MRIspecific contraindications.

Radionucleotide studies are another modality that is useful for diagnosing spinal infections. These include technetium-99m bone scans, gallium-67 scans, and indium-111 labeled leukocyte scans. Pathologic changes have been shown to appear sooner on radionucleotide studies compared to plain radiography [22-27]. Gallium scans have demonstrated earlier diagnosis of disc-space infections compared to technetium scans and have a reported sensitivity of 89%, specificity of 85% and accuracy of 86% [22,23,28]. Technetium-99m scans have a reported sensitivity of 90%, specificity of 78% and accuracy of 86%.18 When both gallium and technetium scans are performed together, the sensitivity is increased to 90%, specificity 100% and accuracy is 94%.18

Indium-111 scans are known to be sensitive for appendicular skeletal infections, however sensitivity is low in the spine [29–32]. In patients with low-virulence chronic infections, indium-111 scans can provide false-negative results due to white blood cell pooling with any inflammatory process [31]. Indium scans may also result in false-positive results in neoplastic conditions. One important advantage of indium-111 scans is the ability to differentiate noninfectious conditions such as hematoma or seroma in patients with unclear soft tissue etiology. This may be a valuable diagnostic step when investigating possible postoperative infections. Overall, most publications endorsed less utility for radionucleotide studies versus MRI. However, in patients with MRI contraindications, technetium-99m combined with gallium-67 studies is another method that has high sensitivity, specificity and diagnostic accuracy similar to MRI.

There is no single diagnostic test with 100% accuracy for these devastating diseases. A full diagnostic workup includes laboratory studies, blood cultures, imaging and tissue histological analyses. It is generally accepted that plain radiography should be the first imaging study obtained, however, diagnostic sensitivity is low. MRI remains the gold standard with the highest sensitivity, specificity and accuracy compared to other modalities. In the presence of MRI contraindications, clinicians should utilize SPECT+gallium-67 or gallium-67 and technetium-99 combined scans to achieve similar diagnostic accuracy as MRI.

REFERENCES

- Sapico FL, Montgomerie JZ. Pyogenic vertebral osteomyelitis: report of nine
- cases and review of the literature. Rev Infect Dis. 1979;1:754-776. Grados F, Lescure FX, Senneville E, Flipo RM, Schmit JL, Fardellone P. Suggestions for managing pyogenic (non-tuberculous) discitis in adults. Joint Bone Spine. 2007;74:133–139. doi:10.1016/j.jbspin.2006.11.002.
- Jevtic V. Vertebral infection. Eur Radiol. 2004;14 Suppl 3:E43–E52. doi:10.1007/ soo330-003-2046-x.
- Cornett CA, Vincent SA, Crow J, Hewlett A. Bacterial spine infections in adults: evaluation and management. J Am Acad Orthop Surg. 2016;24:11-18.
- doi:10.5435/JAAOS-D-13-00102. Golimbu C, Firooznia H, Rafii M. CT of osteomyelitis of the spine. AJR Am J Roentgenol. 1984;142:159-163. doi:10.2214/ajr.142.1.159.
- Kattapuram SV, Phillips WC, Boyd R. CT in pyogenic osteomyelitis of the spine. AJR Am J Roentgenol. 1983;140:1199–1201. doi:10.2214/ajr.140.6.1199.
- Lardé D, Mathieu D, Frija J, Gaston A, Vasile N. Vertebral osteomyelitis: disk hypodensity on CT. AJR Am J Roentgenol. 1982;139:963–967. doi:10.2214/ ajr.139.5.963
- Brant-Zawadzki M, Burke VD, Jeffrey RB. CT in the evaluation of spine infection. Spine. 1983;8:358–364. Fraser RA, Ratzan K, Wolpert SM, Weinstein L. Spinal subdural empyema.
- Arch Neurol. 1973;28:235–238. Love C, Patel M, Lonner BS, Tomas MB, Palestro CJ. Diagnosing spinal osteomyelitis: a comparison of bone and Ga-67 scintigraphy and magnetic resonance imaging. Clin Nucl Med. 2000;25:963–977.
 Abram SR, Tedeschi AA, Partain CL, Blumenkopf B. Differential diagnosis of
- severe back pain using MRI. South Med J. 1988;81:1487-1492.
- Bruns J, Maas R. Advantages of diagnosing bacterial spondylitis with magnetic resonance imaging. Arch Orthop Trauma Surg. 1989;108:30-35.
- Carragee EJ. Pyogenic vertebral osteomyelitis. J Bone Joint Surg Am. 1997;79:874-880.
- Carragee EJ. The clinical use of magnetic resonance imaging in pyogenic vertebral osteomyelitis. Spine. 1997;22:780-78
- Carragee EJ, Kim DH. A prospective analysis of magnetic resonance imaging findings in patients with sciatica and lumbar disc herniation. Correlation of outcomes with disc fragment and canal morphology. Spine. 1997;22:1650-
- Dagirmanjian A, Schils J, McHenry M, Modic MT. MR imaging of vertebral osteomyelitis revisited. AJR Am J Roentgenol. 1996;167:1539–1543. doi:10.2214/ ajr.167.6.8956593
- Ledermann HP, Schweitzer ME, Morrison WB, Carrino JA. MR imaging findings in spinal infections: rules or myths? Radiology. 2003;228:506-514. doi:10.1148/radiol.2282020752.
- Modic MT, Feiglin DH, Piraino DW, Boumphrey F, Weinstein MA, Duchesneau PM, et al. Vertebral osteomyelitis: assessment using MR. Radiology.
- 1985;157:157-166. doi:10.1148/radiology.157.1.3875878. Post MJ, Quencer RM, Montalvo BM, Katz BH, Eismont FJ, Green BA. Spinal infection: evaluation with MR imaging and intraoperative US. Radiology. 1988;169:765-771. doi:10.1148/radiology.169.3.3055039. Shih TT, Huang KM, Hou SM. Early diagnosis of single segment vertebral
- osteomyelitis-MR pattern and its characteristics. Clin Imaging. 1999;23:159-
- Post MJ, Sze G, Quencer RM, Eismont FJ, Green BA, Gahbauer H. Gadolinium-
- enhanced MR in spinal infection. J Comput Assist Tomogr. 1990;14:721–729. Bruschwein DA, Brown MI, McLeod RA. Gallium scintigraphy in the evaluation of disk-space infections: concise communication. J Nucl Med. 1980;21:925-927
- Dux S, Halevi J, Pitlik S, Rosenfeld JB. Early diagnosis of infective spondylitis with Gallium-67. Isr J Med Sci. 1981;17:451-452. Adatepe MH, Powell OM, Isaacs GH, Nichols K, Cefola R. Hematogenous
- pyogenic vertebral osteomyelitis: diagnostic value of radionuclide bone imaging, J Nucl Med. 1986;27:1680–1685. Haase D, Martin R, Marrie T. Radionuclide imaging in pyogenic vertebral
- osteomyelitis. Clin Nucl Med 1980;5:533–537.
 Osenbach RK, Hitchon PW, Menezes AH. Diagnosis and management of
- pyogenic vertebral osteomyelitis in adults. Surg Neurol. 1990;33:266-275
- Waldvogel FA, Papageorgiou PS. Osteomyelitis: the past decade. N Engl J Med. 1980;303:360-70. doi:10.1056/NEJM198008143030703. Norris S, Ehrlich MG, McKusick K. Early diagnosis of disk space infection
- with 67Ga in an experimental model. Clin Orthop Relat Res. 1979:293–298
- Merkel KD, Fitzgerald RH, Brown ML. Scintigraphic evaluation in musculo-skeletal sepsis. Orthop Clin North Am. 1984;15:401–416. Fernandez-Ulloa M, Vasavada PJ, Hanslits ML, Volarich DT, Elgazzar AH.
- Diagnosis of vertebral osteomyelitis: clinical, radiological and scintigraphic features. Orthopedics. 1985;8:1144-1150
- Whalen JL, Brown ML, McLeod R, Fitzgerald RH. Limitations of indium leukocyte imaging for the diagnosis of spine infections. Spine. 1991;16:193-
- Wukich DK, Van Dam BE, Abreu SH. Preoperative indium-labeled white blood cell scintigraphy in suspected osteomyelitis of the axial skeleton. Spine. 1988;13:1168-1170.

Authors: John Koerner, Christopher Kepler, Anand Segar

QUESTION 2: Is there a role for computed tomography (CT) scan with contrast in the diagnosis of spinal infections in patients who cannot undergo magnetic resonance imaging (MRI)?

RECOMMENDATION: Although evidence is limited for the routine use of CT scan with contrast, there is a role for it to be used in the presence of spine infection where MRI is contraindicated or when other advanced imaging is not available

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Although there is growing evidence of the safety of MRI in the presence of implanted metallic devices [1], obtaining such a study may not always be possible. CT with either extradural or intravenous contrast can be used to identify spine infections.

Prior to the wide adoption of MRI, CT myelography was commonly used to diagnose extradural compressive pathology such as epidural abscesses [2]. The use of this invasive investigation in the setting of postoperative spine epidural abscess has not been studied. However, it can be assumed that the accuracy will be lower due to metal artefact [3].

The role of CT with intravenous contrast in the postoperative setting is unclear and has not been directly studied. CT is most useful in identifying implant and bony related complications such as implant loosening, endplate erosion and destruction. The addition of contrast provides information on paraspinal soft tissue involvement, phlegmon or abscesses albeit with lower sensitivity and specificity when compared to MRI [4].

REFERENCES

- Russo RJ, Costa HS, Silva PD, Anderson JL, Arshad A, Biederman RWW, et al. Assessing the risks associated with MRI in patients with a pacemaker or defibrillator. N Engl J Med. 2017;376:755–764. doi:10.1056/NEJM0a1603265.

 Tyrrell PN, Cassar-Pullicino VN, McCall IW. Spinal infection. Eur Radiol.
- 1999;9:1066-1077. doi:10.1007/s003300050793. Chaudhary SB, Vives MJ, Basra SK, Reiter MF. Postoperative spinal wound
- infections and postprocedural diskitis. J Spinal Cord Med. 2007;30:441–451.
- Sundaram VK, Doshi M. Infections of the spine: a review of clinical and imaging findings. Appl Radiol. 2016;45(8):10-20.

Author: Glenn S. Russo

QUESTION 3: Is there a role for nuclear imaging (e.g., positron emission tomography scan (PET)) in the diagnosis of spinal infections?

RECOMMENDATION: PET scan, preferably PET-computed tomography (PET-CT), can be used as an adjunct to magnetic resonance imaging (MRI) to diagnose spinal infections when an MRI cannot be performed or is inconclusive.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

At the present time, MRI is the imaging test of choice for diagnosing spondylodiscitis (SD). This study should be performed when SD is suspected to avoid the morbidity and mortality associated with a delay in diagnosis. MRI is a favored choice as part of an infectious work up due to its lack of ionizing radiation, multi-planar capability, superior soft tissue contrast and ability to evaluate the neural structures. It has a sensitivity and specificity of 97% and 93% respectively. Ultimately, its accuracy in diagnosing SD is 94% [1-3]. A typical protocol should include T1- and T2-weighted sequences with gadolinium. T2 and post-gadolinium T1-weighted sequences should also be performed with fat suppression to increase the sensitivity of identifying pathology [4,5]. Furthermore, MRI allows for the evaluation of bone marrow edema and disc space inflammation, as well as paraspinal and epidural soft tissue involvement. Gadolinium is helpful in differentiating phlegmonous changes versus abscess

Fluorine-18-fluorodeoxyglucose (18F-FDG) is the radionuclideimaging test that can be a useful compliment to MRI. The role of 18F-FDG in the diagnosis of SD has been extensively investigated [6–13]. It has shown acceptable levels of sensitivity and specificity and is useful when MRI cannot be performed or is inconclusive. In addition to its value for diagnosing spondylodiscitis, 18F-FDG can be utilized to monitor response to treatment. Gallium-67-SPECT/CT is an acceptable alternative when 18F-FDG is not available [14].

REFERENCES

- Dagirmanjian A, Schils J, McHenry M, Modic MT. MR imaging of vertebral osteomyelitis revisited. AJR Am J Roentgenol. 1996;167:1539–1543. doi:10.2214/ air.167.6.8956593
- Ledermann HP, Schweitzer ME, Morrison WB, Carrino JA. MR imaging findings in spinal infections: rules or myths? Radiology. 2003;228:506-514. doi:10.1148/radiol.2282020752
- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native vertebral osteomyelitis in adults. Clin Infect Dis. 2015;61:e26-e46. doi:10.1093/cid/civ482. Longo M, Granata F, Ricciardi K, Gaeta M, Blandino A. Contrast-enhanced
- MR imaging with fat suppression in adult-onset septic spondylodiscitis. Eur Radiol. 2003;13:626-637. doi:10.1007/s00330-002-1411-5.
- Diehn FE. Imaging of spine infection. Radiol Clin North Am. 2012;50:777-
- 798. doi:10.1016/j.rcl.2012.04.001. Guhlmann A, Brecht-Krauss D, Suger G, Glatting G, Kotzerke J, Kinzl L, et al. Fluorine-18-FDG PET and technetium-99m antigranulocyte antibody scintigraphy in chronic osteomyelitis. J Nucl Med. 1998;39:2145–2152.
- Zhuang H, Duarte PS, Pourdehand M, Shnier D, Alavi A. Exclusion of chronic osteomyelitis with F-18 fluorodeoxyglucose positron emission tomographic imaging. Clin Nucl Med. 2000;25:281-284.

- Guhlmann A, Brecht-Krauss D, Suger G, Glatting G, Kotzerke J, Kinzl L, et al. Chronic osteomyelitis: detection with FDG PET and correlation with histopathologic findings. Radiology. 1998;206:749-754. doi:10.1148/radiology.206.3.9494496.
- Schiesser M, Stumpe KDM, Trentz O, Kossmann T, Von Schulthess GK. Detec $tion\,of\,metallic\,implant-associated\,in fections\,with\,FDG\,PET\,in\,patients\,with$ trauma: correlation with microbiologic results. Radiology. 2003;226:391–398. doi:10.1148/radiol.2262011939
- Kälicke T, Schmitz A, Risse JH, Arens S, Keller E, Hansis M, et al. Fluorine-18 fluorodeoxyglucose PET in infectious bone diseases: results of histologically confirmed cases. Eur J Nucl Med. 2000;27:524-528.
- Schmitz A, Risse JH, Grünwald F, Gassel F, Biersack HJ, Schmitt O. Fluorine-18 fluorodeoxyglucose positron emission tomography findings in spondylo-
- discitis: preliminary results. Eur Spine J. 2001;10:554–539.

 Meller J, Köster G, Liersch T, Siefker U, Lehmann K, Meyer I, et al. Chronic bacterial osteomyelitis: prospective comparison of (18)F-FDG imaging with a dual-head coincidence camera and (111)In-labelled autologous leucocyte scintigraphy. Eur J Nucl Med Mol Imaging. 2002;29:53-60. doi:10.1007/ 00259-001-0661-9.
- Hungenbach S, Delank K-S, Dietlein M, Eysel P, Drzezga A, Schmidt MC. 18F-fluorodeoxyglucose uptake pattern in patients with suspected spondylodiscitis. Nucl Med Commun. 2013;34:1068–1074. doi:10.1097/MNM.obo13e328365abec.
- Bagrosky BM, Hayes KL, Koo PJ, Fenton LZ. 18F-FDG PET/CT evaluation of children and young adults with suspected spinal fusion hardware infection. Pediatr Radiol. 2013;43:991-1000. doi:10.1007/s00247-013-2654-9.

Author: Susana Núñez-Pereira

QUESTION 4: How can postoperative infections be distinguished from normal postoperative changes on magnetic resonance imaging (MRI)?

RECOMMENDATION: The presence of an abscess in the back muscles or posterior site, confirmed by gadolinium enhancement, is the most frequently-reported change on MRI of patients with surgical site infection (SSI). The presence of a collection of fluid outside the head of the pedicle screws is another sign of SSI.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 71%, Disagree: 8%, Abstain: 21% (Super Majority, Strong Consensus)

RATIONALE

A search was conducted using the MeSH terms "spine AND MRI AND surgical site infection." The initial search yielded 149 references, and after screening, 13 abstracts remained. However, only three studies assessed the use of MRI for postoperative spine infections and were found eligible.

Kanayama et al. retrospectively used MRI in 20 patients with surgical site infections after instrumented spinal surgery [1]. In their series they considered two markers for diagnosing SSI: (1) the presence or absence of osteomyelitis at the instrumented vertebra and (2) the presence or absence of intervertebral abscess. All 20 patients had a confirmed SSI, but in 7 MRIs it was considered negative. The study mainly aimed to assess the utility of MRI to confirm the severity of the infection. Using the above-mentioned criteria, they tried to predict the need for implant removal. However, MRI was not evaluated as a diagnostic tool for assessing the presence or absence of infection.

Kim et al. reviewed 43 patients with MRI after SSI [2]. First, they divided their infections on an anatomical basis, assessing if it affected only the posterior region (31 cases), only the anterior area or both posterior and anterior regions [2]. In addition, they looked for abscess in different spinal locations (posterior epidural space, laminectomy site, back muscles, subcutaneous fat layer, paravertebral space, psoas muscle and anterior epidural space). They also evaluated the presence of osteomyelitis of the vertebral body and discitis. The most frequent findings were abscesses in the back muscles in 40 patients (93%), abscesses in the laminectomy site in 29 (67.4%) and in the subcutaneous fat layer in 27 (62.8%). All abscesses were identified by the presence of peripheral rim or diffused enhancement of adjacent soft tissue after administration of intravenous gadolinium.

They did not compare their findings with those of patients without confirmed SSI. The authors concluded that for diagnosing infection, the posterior surgical field was more important than the vertebral body or the disc area. This conclusion supports the findings of the previous study by Kanayama, in which seven patients with SSIs did not show involvement of the vertebral body or the disc area.

Finally, Kimura et al. published a comparative study on postoperative MRI including 17 patients with a deep SSI and 64 non-SSI controls who had an MRI examination within 4 weeks after surgery [3]. Their investigation focused on the "pedicle screw fluid sign" (PS fluid sign) and did not search for other signs of infection. The PS fluid sign refers to the collection of fluid outside the head of a pedicle screw, suggesting the presence of an abscess on axial MRI scans. The authors observed that fluid collections medial to the pedicle screw head are not infrequent. They considered that when the collection expands outside the head of the screw into the paravertebral

muscles, it is likely to be an abscess. In their view, artifacts have little effect on the area outside the screw head, compared with the inside. In their study, this sign was positive in 15 of 17 deep SSI infections and only in 7 of 64 patients without infection. Sensitivity was 88.2%, specificity 89.1%, positive predictive value 68.1% and negative predictive value 96.6%.

In conclusion, abscesses in the back muscles, laminectomy site and subcutaneous fat layer, after administration of gadolinium were the most common findings related with surgical site infection. In addition, the PS fluid sign had a sensitivity of 88.2% and specificity of 89.1%.

REFERENCES

- Kanayama M, Hashimoto T, Shigenobu K, Oha F, Iwata A, Tanaka M. MRI-based decision making of implant removal in deep wound infection after instrumented lumbar fusion. Clin Spine Surg. 2017;30(2):
- Kim SJ, Lee SH, Chung HW, Lee MH, Shin MJ, Park SW. Magnetic resonance imaging patterns of post-operative spinal infection: relationship between the clinical onset of infection and the infection site. J Korean Neurosurg
- Soc. 2017;60 (4):448-455.
 Kimura H, Shikata J, Odate S, Soeda T. Pedicle screw fluid sign: an indication on magnetic resonance imaging of a deep infection after posterior spinal instrumentation. Clin Spine Surg. 2017;30(4):169-175.

Treatment

3.1. TREATMENT: GENERAL PRINCIPLES

Author: Claus Simpfendorfer

QUESTION 1: Can a non-surgical approach be used to treat postoperative spine infections? If so, what factors predict a successful outcome?

RECOMMENDATION: Postoperative spine infections should be treated with irrigation and debridement (with or without implant removal) followed by appropriate antibiotic therapy. Antibiotic suppression without surgical intervention should be attempted in cases where the patient is not a surgical candidate, or in attempt to achieve spinal fusion prior to implant removal.

LEVEL OF EVIDENCE: Strong

DELEGATE VOTE: Agree: 80%, Disagree: 7%, Abstain: 13% (Super Majority, Strong Consensus)

RATIONALE

Postoperative surgical site infections are a major complication that occur between 1 and 12% of all spinal surgeries [1–3]. Treatment varies based on general location in relation to superficial, or deep to the muscular fascia, and the time from initial surgery, with early infections occurring before 90 days and late infections occurring after 90

In the case of superficial wound infections, local debridement, healing by secondary intention and a short course of antibiotics is usually sufficient [4]. Deep surgical site infections, on the other hand, require aggressive irrigation and debridement with or without implant removal. The retention of hardware predominantly depends on if the infection is early or late. Several studies have demonstrated that hardware can be retained successfully following aggressive irrigation and debridement in the setting of early infection, except in cases where the implants are loose or there is bony involvement [5-9]. Optimal treatment of delayed infections is aggressive irrigation and debridement with implant removal [10–12]. In the cases where spinal fusion has been achieved, implant removal is routinely performed. However, in cases of fusion failure or pseudoarthrosis, surgical options include aggressive debridement and irrigation with attempted implant retention, implant removal with primary or delayed reimplantation or implant removal without reimplantation [6,13-16].

Antibiotic suppression without surgical intervention is attempted in cases where the patient is not a surgical candidate, or in an attempt to achieve spinal fusion prior to implant removal.

REFERENCES

Pull ter Gunne AF, Hosman AJ, Cohen DB, et al. A methodological systematic review on surgical site infections following spinal surgery: part 1: risk factors. Spine (Phila Pa 1976). 2012;37(24):2017–2033.

- Schuster JM, Rechtine G, Norvell DC, Dettori JR. The influence of periopera-
- tive risk factors and therapeutic interventions on infection rates after spine surgery: a systematic review. Spine (Phila Pa 1976). 2010;35(9 Suppl):S125–137. Smith JS, Shaffrey CI, Sansur CA, et al. Rates of infection after spine surgery based on 108,419 procedures: a report from the Scoliosis Research Society Morbidity and Mortality Committee. Spine (Phila Pa 1976). 2011;36(7):556–
- 563. Pull ter Gunne AF, Mohamed AS, Skolasky RL, van Laarhoven CJ, Cohen DB.
- The presentation, incidence, etiology, and treatment of surgical site infections after spinal surgery. Spine (Phila Pa 1976). 2010;35(13):1323-1328. Wang TY, Back AG, Hompe E, Wall K, Gottfried ON. Impact of surgical site infection and surgical debridement on lumbar arthrodesis: a single-institution and surgical debridement on lumbar arthrodesis: tion analysis of incidence and risk factors. J Clin Neurosci. 2017;39:164-169. Mok JM, Guillaume TJ, Talu U, et al. Clinical outcome of deep wound infec-
- tion after instrumented posterior spinal fusion: a matched cohort analysis. Spine (Phila Pa 1976). 2009;34(6):578-583. Ho C, Skaggs DL, Weiss JM, Tolo VT. Management of infection after instru-
- mented posterior spine fusion in pediatric scoliosis. Spine (Phila Pa 1976).
- Carreon IV Clinical outcomes after
- Petilon JM, Glassman SD, Dimar JR, Carreon LY. Clinical outcomes after lumbar fusion complicated by deep wound infection: a case-control study. Spine (Phila Pa 1976). 2012;37(16):1370-1374. Richards BR, Emara KM. Delayed infections after posterior TSRH spinal
- instrumentation for idiopathic scoliosis: revisited. Spine (Phila Pa 1976). 2001;26(18):1990-1996
- Hahn F, Zbinden R, Min K. Late implant infections caused by Propionibacterium acnes in scoliosis surgery. Eur Spine J. 2005;14(8):783.–788. Hedequist D, Haugen A, Hresko T, Emans J. Failure of attempted implant
- retention in spinal deformity delayed surgical site infections. Spine (Phila Pa 1976). 2009;34(1):60-64. Muschik M, Luck W, Schlenzka D. Implant removal for late-developing
- infection after instrumented posterior spinal fusion for scoliosis: reinstrumentation reduces loss of correction. A retrospective analysis of 45 cases. Eur Spine J. 2004;13(7):645–651. Tsiodras S, Falagas ME. Clinical assessment and medical treatment of spine
- infections. Clin Orthop Relat Res. 2006;444:38-50.
- Bose B. Delayed infection after instrumented spine surgery: case reports and review of the literature. Spine J. 2003;3(5):394–399. Kim JI, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infec-
- [16] tion after instrumented spinal fusion. J Spinal Disord Tech. 2010;23(4):258-

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QUESTION 2: When should patients with suspected infections of the spine be referred to an infectious disease department?

RECOMMENDATION: There is no data on the timing or need for a referral to an infectious disease department. We support a multidisciplinary approach to managing clinical spine infections.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Only one paper has addressed the collaboration with an infectious disease-specialized team in order to improve outcomes for patients with spinal surgical site infections (SSIs). The paper is a retrospective study reporting on 40 patients, none of whom needed implant removal [1]. The paper didn't report on the exact timing when collaboration started, but reported three main advantages related with this collaboration:

- 1. Efficient detection of auxiliary bacteria (reached 88%)
- 2. Early treatment with antibiotics
- 3. Appropriate duration of administration of antibiotics

There were no other papers which discussed this issue, and all subsequent searches on related articles yielded no more information on the matter.

From a theoretical point of view, referral, or at least counselling by an infectious diseases specialist, might have some advantages. Antibiotic treatments are more complex today and only specialists are adequately up-to-date on the issue. The appropriate treatment choice might be difficult in patients with allergies, multi-resistant smears or simply a low tolerance for the medication. Adjusting the choice of antibiotic, taking into account side effects and tolerance, will very likely improve compliance, which is paramount in reaching a successful treatment outcome.

REFERENCES

Kobayashi K, Imagama S, Kato D, Ando K, Hida T, Ito K, et al. Collaboration with an infection control team for patients with infection after spine surgery. Am J Infect Control. 2017;45:767–770. doi:10.1016/j.ajic.2017.01.013.

Authors: Dolors Rodriguez-Pardo, Gregory Schroeder

QUESTION 3: Which patients with vertebral osteomyelitis (VO) are suitable for outpatient management? Does any criteria exist to aid in this decision-making?

RECOMMENDATION: There are no studies aiming to identify which patients diagnosed with VO can be treated on an outpatient basis.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

VO, also known as spondylodiscitis, describes an infection of the vertebrae and intervertebral disc. By comparison, discitis describes infection limited to the intervertebral disc, however there are many who consider discitis and VO to be different stages of the same disease process. VO can arise from hematogenous seeding, contiguous spread from infection in adjacent soft tissues or direct inoculation during spinal surgery or procedures (i.e., epidural). Management of native vertebral osteomyelitis (NVO) depends on the location of the infection, disease progression and the patient's general condition including age and comorbidities.

Conservative treatment is reasonable in the early stages with no or minor neurologic deficits or in the case of severe comorbidities. However, in cases of doubt, surgical treatment should be considered. Both options require a concomitant antimicrobial therapy, initially applied intravenously and administered orally thereafter [1]. To date, there is no consistent data from randomized controlled

trials to guide the optimal duration and appropriate route of antibiotic therapy. Although the optimal duration of antibiotic therapy remains controversial, it should never undercut six weeks [2]. Recent Infectious Diseases Society of America (IDSA) guidelines for the diagnosis and treatment of NVO in adults include evidence and opinion-based recommendations for the management of patients with NVO treated with antimicrobial therapy, with or without surgical intervention, but do not address the issue of which patients affected by NVO can be treated on an outpatient basis [3,4]. The extent of pursuing spinal biopsies to determine etiology, antimicrobial therapy, response to treatment and preference for surgical techniques and timing all vary widely in clinical practice with heterogeneous studies limiting comparisons. Surgery, rather than conservative approaches, is being proposed as the default management choice because in carefully-selected patients it can offer faster reduction in pain scores and improved quality of life [5-9]. Due to a heterogeneous and often comorbid patient population and the wide variety of treatment options, no generally applicable guidelines for VO exist and management remains a challenge.

The goals of treatment include establishing a diagnosis and identifying the pathogen, eradicating the infection, preventing or minimizing neurologic involvement, maintaining spinal stability and providing an adequate nutritional state to combat infection. Often, this can be accomplished with non-operative approaches.

The mainstay treatment of pyogenic infections of the spine remains antibiotic therapy and immobilization with a proper orthosis. If nonsurgical treatment fails, however, surgical intervention may be required. Surgery is indicated in the following circumstances: to obtain a bacteriologic diagnosis when closed biopsy is negative or deemed unsafe, when a clinically significant abscess is present (spiking temperatures and evidence of sepsis), in cases of refractory to prolonged non-operative treatment where the sedimentation rate remains high or pain persists, in cases of spinal cord compression causing a neurologic deficit and in cases of substantial deformity or vertebral body destruction, especially in the cervical spine. Alton et al. reported that 75% of patients with an epidural abscess in the cervical spine who underwent medical management failed and that medical management failure was associated with a significantly increased risk of neurologic injury [10]. Patel et al. reported on 128 patients with an epidural abscess and found that 41% failed medical management. However, there were significant predictors of medical failure [11]. Four key predictors were identified, including diabetes mellitus, C-reactive protein (CRP) greater than 115, white blood cell count greater than 12.5 and positive blood culture. Patients with none of the aforementioned parameters only failed 8.3% of the time. Those with one parameter failed 35.4% of the time, those with two parameters failed 40.2% of the time and patients with three or more parameters failed 76.9% of the time.

Once the antibiotic is prescribed by oral route, if the patient is stable, the treatment could be administered in an outpatient setting. Several studies described a successful switch to oral antibiotics after 10 days, using oral agents with a high bio-availability and tissue penetration (i.e., fluorquinolones, rifampin, fusidic acid and clindamycin) [12–15]. A retrospective analysis of all patients diagnosed with NVO, at the University Hospital of Basel, Switzerland, concluded that switching to an oral antibiotic regimen after two weeks of intravenous treatment may be safe, if CRP has decreased compared to baseline CRP and epidural or paravertebral abscesses of significant size have been drained [16]. Importantly, these results do not extend to patients with endocarditis, surgical site infection, and/or vertebral implants. Also, positive blood cultures, neurological abnormalities and staphylococcal infections (compared with negative microbiology) are associated with longer intravenous courses [17].

Outpatient parenteral antibiotic therapy (OPAT) has become an option allowing for early discharge of hospitalized patients who have infections without a reliable oral alternative and requires lengthy antibiotic therapy. It provides numerous benefits, some of the most remarkable being that OPAT permits early discharge and reduces costs, avoids hospitalization trauma in children or immobilization syndrome in the elderly and reduces nosocomial infections by multidrug resistant organisms [17]. OPAT also allows for self-administration of antibiotics using elastomeric pumps [18,19]. Different retrospective studies and case series have reviewed the experience with OPAT in several countries [17,19–27]. β-Lactam antibiotics are commonly used in OPAT with higher treatment success among those treated with ceftriaxone and ertapenem, while oxacillin was associated with a higher rate of antimicrobial discontinuation because of antimicrobial-related complications [17,20,26]. Other alternatives are teicoplanin, telavancin or

daptomycin in the case of gram-positive infections [17,25,28]. All this data regarding OPAT confirms that infection management in an outpatient setting is safe, clinically efficacious, and acceptable for treating a wide range of infections with high levels of patient satisfaction and substantial cost savings. Therefore, OPAT could be considered an effective alternative for appropriately selected elderly patients with vertebral osteomyelitis.

REFERENCES

- Lener S, Hartmann S, Barbagallo GMV, Certo F, Thomé C, Tschugg A. Management of spinal infection: a review of the literature. Acta Neurochir (Wien). 2018;160:487–496. doi:10.1007/s00701-018-3467-2.
- Roblot F, Besnier JM, Juhel L, Vidal C, Ragot S, Bastides F, et al. Optimal duration of antibiotic therapy in vertebral osteomyelitis. Semin Arthritis Rheum. 2007;36:269-277. doi:10.1016/j.semarthrit.2006.09.004
- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native vertebral osteomyelitis in adults. Clin Infect Dis. 2015;61:e26-e36. doi:10.1093/cid/civ482.
- Berbari EF, Kanj SS, Kowalski TJ, Darouiche RO, Widmer AF, Schmitt SK, et al. Executive summary: 2015 Infectious Diseases Society of America (IDSA) Clinical Practice Guidelines for the diagnosis and treatment of native verte bral osteomyelitis in adults. Clin Infect Dis. 2015;61:859–863. doi:10.1093/cid/ civ633
- Doutchi M, Seng P, Menard A, Meddeb L, Adetchessi T, Fuentes S, et al. Changing trends in the epidemiology of vertebral osteomyelitis in Marseille, France. New Microbes New Infect. 2015;7:1–7. doi:10.1016/j. nmni.2015.04.008.
- Cheung WY, Luk KDK. Pyogenic spondylitis. Int Orthop. 2012;36:397–404. doi:10.1007/s00264-011-1384-6.
- Gupta A, Kowalski TJ, Osmon DR, Enzler M, Steckelberg JM, Huddleston PM, et al. Long-term outcome of pyogenic vertebral osteomyelitis: a cohort study of 260 patients. Open Forum Infect Dis. 2014;1:0fu107. doi:10.1093/ofid/ ofu107.
- Kehrer M, Pedersen C, Jensen TG, Hallas J, Lassen AT. Increased shortand long-term mortality among patients with infectious spondylodiscitis compared with a reference population. Spine J. 2015;15:1233-1240.
- doi:10.1016/j.spinee.2015.02.021. Sobottke R, Röllinghoff M, Zarghooni K, Zarghooni K, Schlüter-Brust K, Delank KS, et al. Spondylodiscitis in the elderly patient: clinical mid-term results and quality of life. Arch Orthop Trauma Surg. 2010;130:1083-1091. doi:10.1007/s00402-009-0972-z
- Alton TB, Patel AR, Bransford RJ, Bellabarba C, Lee MJ, Chapman JR. Is there a difference in neurologic outcome in medical versus early operative management of cervical epidural abscesses? Spine J. 2015;15:10-17. doi:10.1016/j.spinee.2014.06.010
- Patel AR, Alton TB, Bransford RJ, Lee MJ, Bellabarba CB, Chapman JR. Spinal epidural abscesses: risk factors, medical versus surgical management, a retrospective review of 128 cases. Spine J. 2014;14:326–330. doi:10.1016/j. spinee.2013.10.046.
- Legrand E, Flipo RM, Guggenbuhl P, Masson C, Maillefert JF, Soubrier M, et al. Management of nontuberculous infectious discitis. tréatments used in 110 patients admitted to 12 teaching hospitals in France. Joint Bone Spine. 2001;68:504–50
- Gouliouris T, Aliyu SH, Brown NM. Spondylodiscitis: update on diagnosis and management. J Antimicrob Chemother. 2010;65 Suppl 3:iii11-24.
- doi:10.1093/jac/dkq303. Fleege C, Wichelhaus TA, Rauschmann M. [Systemic and local antibiotic therapy of conservative and operative treatment of spondylodiscitis]. Orthopade. 2012;41:727-735. doi:10.1007/s00132-012-1920-0.
- Karamanis EM, Matthaiou DK, Moraitis LI, Falagas ME. Fluoroquinolones versus beta-lactam based regimens for the treatment of osteomyelitis: a meta-analysis of randomized controlled trials. Spine. 2008;33:E297-E304.
- doi:10.1097/BRS.ob013e31816f6c22. Babouee Flury B, Elzi L, Kolbe M, Frei R, Weisser M, Schären S, et al. Is switching to an oral antibiotic regimen safe after 2 weeks of intravenous treatment for primary bacterial vertebral osteomyelitis? BMC Infect Dis. 2014;14:226. doi:10.1186/1471-2334-14-226.
- Esposito S, Leone S, Noviello S, Ianniello F, Fiore M, Russo M, et al. Outpatient parenteral antibiotic therapy for bone and joint infections: an İtalian multicenter study. J Chemother. 2007;19:417-422. doi:10.1179/ joc.2007.19.4.417
- Saillen L, Arensdorff L, Moulin E, Voumard R, Cochet C, Boillat-Blanco N, et al. Patient satisfaction in an outpatient parenteral antimicrobial therapy (OPAT) unit practising predominantly self-administration of antibiotics with elastomeric pumps. Eur J Clin Microbiol Infect Dis. 2017;36:1387–1392. doi:10.1007/s10096-017-2944-5.
- Carrega G, Bartolacci V, Burastero G, Casalino Finocchio G, Izzo M, Ronca A, et al. [Chronic osteomyelitis due to Pseudomonas aeruginosa: treatment with elastomeric infusor in an outpatient setting]. Infez Med. 2011;19:257–
- Lee B, Tam I, Weigel B, Breeze JL, Paulus JK, Nelson J, et al. Comparative outcomes of β-lactam antibiotics in outpatient parenteral antibiotic therapy: treatment success, readmissions and antibiotic switches. J Antimicrob Chemother. 2015;70:2389–2396. doi:10.1093/jac/dkv130.

- [21] Galpérine T, Ader F, Piriou P, Judet T, Perronne C, Bernard L. [Outpatient parenteral antimicrobial therapy (OPAT) in bone and joint infections]. Med Mal Infect. 2006;36:132–137. doi:10.1016/j.medmal.2006.01.002.
- [22] Durojaiye OC, Bell H, Andrews D, Ntziora F, Cartwright K. Clinical efficacy, cost analysis and patient acceptability of outpatient parenteral antibiotic therapy (OPAT): a decade of Sheffield (UK) OPAT service. Int J Antimicrob Agents. 2018;51:26–32. doi:10.1016/j.ijantimicag.2017.03.016.
 [23] Kortajarena X, Goenaga MA, Ibarguren M, Azkune H, Bustinduy MJ, Fuertes
- [23] Kortajarena X, Goenaga MA, Ibarguren M, Azkune H, Bustinduy MJ, Fuertes A, et al. Outpatient parenteral antimicrobial therapy for infective endocarditis in patients over 80 years. Rev Esp Quimioter. 2017;30:276–279.
- [24] Madaline T, Nori P, Mowrey W, Zukowski E, Gohil S, Sarwar U, et al. Bundle in the Bronx: impact of a transition-of-care outpatient parenteral antibiotic therapy bundle on all-cause 30-day hospital readmissions. Open Forum Infect Dis. 2017;4:0fx097. doi:10.1093/ofid/ofx097.
- [25] Schroeder CP, Van Anglen LJ, Dretler RH, Adams JS, Prokesch RC, Luu Q, et al. Outpatient treatment of osteomyelitis with telavancin. Int J Antimicrob Agents. 2017;50:93-96. doi:10.1016/j.ijantimicag.2017.01.034.
- [26] Fernandes P, Milliren C, Mahoney-West HM, Schwartz L, Lachenauer CS, Nakamura MM. Safety of outpatient parenteral antimicrobial therapy in children. Pediatr Infect Dis J. 2018;37:157–163. doi:10.1097/INF.000000000001716.
- [27] Quirke M, Curran EM, O'Kelly P, Moran R, Daly E, Aylward S, et al. Risk factors for amendment in type, duration and setting of prescribed outpatient parenteral antimicrobial therapy (OPAT) for adult patients with cellulitis: a retrospective cohort study and CART analysis. Postgrad Med J. 2018;94:25–31. doi:10.1136/postgradmedj-2017-134968.
- 28] Dos Remedios E. Daptomycin for the treatment of osteomyelitis associated with a diabetic foot ulcer. Wounds. 2009;21:286–289.

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QUESTION 4: What is the optimal treatment of spinal infections caused by *Propionibacterium acnes* (*P. acnes*)?

RECOMMENDATION: When possible, patients should undergo complete removal of implants after *Cutibacterium acnes* (*C. acnes*) (formerly *P. acnes*) infection, especially in the setting of latent infection. Antibiotic regimens typically involve specific parenteral antibiotics for a period of greater than two weeks, with the most common antibiotic duration being six weeks of multiple parenteral and/or oral agents. However, the duration of antibiotic treatment is highly variable. It is unclear in which setting patients may be successfully treated with antibiotic therapy alone and instrumentation may be retained. Penicillin is currently the standard of care, but other non beta-lactam antibiotics should be considered based on the susceptibility profile.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 73%, Disagree: 7%, Abstain: 20% (Super Majority, Strong Consensus)

RATIONALE

P. acnes is an anaerobic, gram-positive bacillus existing as normal flora of the skin and sebaceous glands and was originally considered a common contaminant of blood cultures as well as an uncommon cause of brain, pulmonary and dental infections [1]. *C. acnes* infections are commonly thought to originate from patient skin approximation with surgical sites, are frequently poly-microbial, require an extended incubation period in culture media for diagnosis and form a resistant biofilm, making treatment with antibiotics alone difficult [2–4].

P. acnes infection of the spine was first reported as an etiology of spine infection by Serushan et al. in 1982 [5]. The patient presented with osteomyelitis of the cervical spine and was treated with 40 days of intravenous penicillin with resolution of his fever and neck pain. C. acnes has subsequently been implicated in vertebral osteomyelitis and discitis and may present with insidious onset of back pain, fever and/or neurologic symptoms, with treatment typically involving administration of parenteral antibiotics. Additional debridement or percutaneous drainage of abscesses occurs in rare cases [6–8]. Duration of antibiotics ranged from 2-28 weeks in one series, and typically involved multiple agents due to the frequency of co-infection with other pathogens including Staphylococcus, Lactobacillus and Enterococcus species [9].

Tsai et al. reported on successful treatment of two cases of *C. acnes* osteomyelitis of the cervical spine with anterior debridement, decompression and fusion with autograft and treatment with a combination of oral and parenteral antibiotics for 6-16 weeks [10]. Overall, the decision to treat *C. acnes* vertebral osteomyelitis and discitis with surgery, antibiotics or a combination of these approaches has been made on a case-by-case basis. No well-defined, widely-applicable treatment regimen was identified in the literature.

C. acnes also frequently presents as a delayed infection after spinal instrumentation, which has been attributed to its low virulence and slow growth rate, and is common in instrumented pediatric scoliosis surgery [4,11-17]. Viola et al. reported a series of eight patients with delayed infection, one of which had C. acnes infection and was treated with irrigation and debridement, removal of instrumentation and six weeks of cefotetan with good results and no loss of balance or alignment at midterm follow-up. Of 23 patients with delayed infections after posterior TSRH instrumentation, Richards and Emara found that the causative agent in delayed infections was C. acnes in 12 (52.1%). Patients underwent removal of instrumentation with either primary or delayed closure and parenteral antibiotics (two to five days) followed by a course of oral antibiotics for an additional two to four weeks [18]. Tribus reported on a delayed infection with Staphylococcus epidermidis and C. acnes resulting in laminar erosions seven years after TSRH instrumentation. The patient was treated with removal of instrumentation and seven weeks of intravenous vancomycin and oral rifampin with resolution of pain and infection [12]. In cases of late implant infections, successful treatment typically involved implant removal and greater than two weeks of a combination of parenteral and oral antibiotics.

In the largest single study evaluating treatment of *C. acnes* infection after Cotrel-Dubousset instrumentation, Bemer et al. conducted a retrospective study investigating various treatment regimens including complete or partial implant removal, implant replacement and maintenance of implants with irrigation and debridement, both with and without antibiotics. Patients who underwent partial removal with antibiotic monotherapy or absence of antibiotic therapy were more likely to develop a secondary infection. Ultimately, wide variation in treatment regimens prevented more mean-

ingful analysis of the results, though the authors concluded that complete removal of implants should be performed when possible and antibiotics should be tailored to the sensitivities of the specific organism and given for a duration of three to six months or less than three months when following total implant removal [19]. In another large case series of surgical site infection (SSI) after spine surgery, Maruo and Berven listed C. acnes infection as an independent risk factor for treatment failure (p = 0.042) [4]. Though they did not comment on the specific treatment strategies utilized for patients with *C. acnes* SSI, they note that 7 of 12 patients (58%) with late infection treated with implant retention and antibiotics required subsequent implant removal.

Due to the variation in treatment strategies for *Propionibacterium* acnes infections of the spine and the lack of prospective trials evaluating optimal antibiotic regimen, the optimal treatment of spinal infections with *C. acnes* is indeterminate. However, given reports of numerous successful treatment strategies in the literature, complete removal of implants when applicable followed by an extended course of parenteral antibiotics results in overall high cure rates for *C. acnes* infections of the spine.

REFERENCES

- Noble RC, Overman SB. Propionibacterium acnes osteomyelitis: case report and review of the literature. J Clin Microbiol. 1987;25:251–254.

 Shiono Y, Watanabe K, Hosogane N, Tsuji T, Ishii K, Nakamura M, et al. Sterility of posterior elements of the spine in posterior correction surgery.
- Spine. 2012;37:523–526. doi:10.1097/BRS.0b013e318224d7b2.
 Savage JW, Weatherford BM, Sugrue PA, Nolden MT, Liu JC, Song JK, et al. Efficacy of surgical preparation solutions in lumbar spine surgery. J Bone
- Joint Surg Am. 2012;94:490–494. doi:10.2106/JBJS.K.00471.

 Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19:398-404. doi:10.1007/s00776-014-0545-z.

- Serushan M, Spencer DL, Yeh WL, Kaminski M, Skosey JL. Osteomyelitis of cervical spine from Propionibacterium acnes. Arthritis Rheum. 1982;25:346-
- Saeed MU, Mariani P, Martin C, Smego RA, Potti A, Tight R, et al. Anaer-
- obic spondylodiscitis: case series and systematic review. South Med J. 2005;98:144-148. doi:10.1097/01.SMJ.0000129928.03804.2A.
 Harris AE, Hennicke C, Byers K, Welch WC. Postoperative discitis due to Propionibacterium acnes: a case report and review of the literature. Surg Neurol. 2005;63:538–541; discussion 541. doi:10.1016/j.surneu.2004.06.012.
- Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Osmon DR. Propionibacterium acnes vertebral osteomyelitis: seek and ye shall find? Clin Orthop Relat Res. 2007;461:25–30. doi:10.1097/BLO.ob013e318073c25d
- Uçkay I, Dinh A, Vauthey L, Asseray N, Passuti N, Rottman M, et al. Spondylodiscitis due to Propionibacterium acnes: report of twenty-nine cases and a review of the literature. Clin Microbiol Infect. 2010;16:353–358. doi:10.1111/ j.1469-0691.2009.02801.x.
- Tsai CE, Lee FT, Chang MC, Yu WK, Wang ST, Liu CL. Primary cervical osteomyelitis. J Chin Med Assoc. 2013;76:640-647. doi:10.1016/jj.jcma.2013.07.011. Heggeness MH, Esses SI, Errico T, Yuan HA. Late infection of spinal instru-
- mentation by hematogenous seeding. Spine. 1993;18:492-496.
- Tribus CB, Garvey KE. Full-thickness thoracic laminar erosion after posterior spinal fusion associated with late-presenting infection. Spine. 2003;28:E194-E197. doi:10.1097/01.BRS.0000062005.15715.C2.
 Do TT, Strub WM, Witte D. Subacute Propionibacterium acnes osteomy-
- elitis of the spine in an adolescent. J Pediatr Orthop B. 2003;12:284-287. doi:10.1097/01.bpb.0000049566.52224.b3.
- Hahn F, Zbinden R, Min K. Late implant infection caused by Propionibacterium acnes in scoliosis surgery. Eur Spine J. 2005;14:783-788. doi:10.1007/ soo586-004-0854-6. Rihn JA, Lee JY, Ward WT. Infection after the surgical treatment of adolescent
- idiopathic scoliosis: evaluation of the diagnosis, treatment, and impact on clinical outcomes. Spine. 2008;33:289–294. doi:10.1097/BRS.ob013e318162016e.
- Di Silvestre M, Bakaloudis G, Lolli F, Giacomini S. Late-developing infection following posterior fusion for adolescent idiopathic scoliosis. Eur Spine J. 2011;20 Suppl 1:S121-S127. doi:10.1007/s00586-011-1754-1.
- Zeng Y, Chen Z, Guo Z, Qi Q, Li W, Sun C. Complications of correction for focal kyphosis after posterior osteotomy and the corresponding management. J Spinal Disord Tech. 2012;26:1. doi:10.1097/BSD.ob013e3182499237.
- Richards BR, Emara KM. Delayed infections after posterior TSRH spinal instrumentation for idiopathic scoliosis: revisited. Spine. 2001;26:1990–
- Bémer P, Corvec S, Tariel S, Asseray N, Boutoille D, Langlois C, et al. Significance of Propionibacterium acnes-positive samples in spinal instrumentation. Spine. 2008;33:E971-E976. doi:10.1097/BRS.obo13e31818e28dc.

3.2. TREATMENT: ANTIBIOTICS

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QUESTION 1: Is there a role for oral antibiotics in the treatment of early postoperative spine infections?

RECOMMENDATION: There may be a role for highly bioavailable oral antibiotics in the treatment of early postoperative spine infection in select circumstances.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Broad-spectrum intravenous (IV) antibiotics may be indicated prior to identification of the infecting organism in patients with early postoperative infections while waiting for surgical intervention, or for patients who are medically unstable and cannot undergo surgery [1]. Other than the latter cases, there is no role for oral antibiotics alone in the treatment of patients with acute postoperative spine infections. Patients with established postoperative infections of the spine require surgical intervention.

The administration of antibiotics may potentially adversely affect the outcome of treatment of these patients by interfering with

isolation of the infecting organism. Antibiotic therapy should be withheld in patients with suspected spine infection, as the yield for biopsy to isolate the infecting organism is reduced when the antibiotic is administered. In a study by Cornett et al., the yield for biopsy culture dropped from 80% for those who did not receive antibiotics to 48% for those who did [1]. Another study of 87 patients, however, demonstrated that the yield for biopsy of spondylodiscitis did not significantly decrease with prior treatment of antibiotics [2]. Despite this, it is still recommended that antibiotics be withheld when possible. If antibiotics are to be administered, biopsy is still indi-

cated to isolate the infecting organism and allow for optimal treatment of the patient.

In a large case series of 1,980 patients, 74 infections were diagnosed [3]. The treatment algorithm consisted of six weeks of IV antibiotics if the patient was not fused. If the patient was fused, Staphylococcus aureus and gram-negative infections were treated with six weeks of IV antibiotics followed by six weeks of oral antibiotics with implant removal. In patients with propionibacteria and coagulasenegative Staphylococcus, four weeks of oral antibiotics were given. Oral antibiotics were not recommended as an initial treatment. Other studies have demonstrated the benefit of oral antibiotics as suppression therapy after treatment with surgical debridement and a course of IV antibiotics [4,5].

Multiple other studies have demonstrated the benefit of surgical debridement and IV antibiotics for infection [6]. In a consecutive case series of 2,391 patients, 46 cases of wound infection were identified and all treated with surgical debridement [7]. One series of 111 patients identified eight patients with postoperative infections after posterior lumbar interbody fusion [8]. All were treated with irrigation and debridement followed by four to six weeks of intravenous antibiotics followed by another six to nine weeks of oral antibiotics.

Multiple case series and expert opinion studies recommend avoiding oral antibiotics in suspected postoperative infection until culture samples are taken for better diagnosis and accurate treatment of these patients [9]. The majority of patients with established postoperative infection require surgical debridement.

REFERENCES

- Cornett CA, Vincent SA, Crow J, Hewlett A. Bacterial spine infections in adults: evaluation and management. J Am Acad Orthop Surg. 2016;24(1):11-
- Robino, 5435 JAAOS-D-13-00102.

 Foreman SC, Schwaiger BJ, Gempt J, et al. MR and CT imaging to optimize CT-guided biopsies in suspected Spondylodiscitis. World Neurosurg. 2017;99:726-734.e7. doi:10.1016/j.wneu.2016.11.017. Collins I, Wilson-MacDonald J, Chami G, et al. The diagnosis and manage-
- ment of infection following instrumented spinal fusion. Eur Spine J. 2008;17(3):445-450. doi:10.1007/s00586-007-0559-8. Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44(7):913– 920. doi:10.1086/512194.
- Levi a D, Dickman Ca, Sonntag VK. Management of postoperative infections after spinal instrumentation. J Neurosurg. 1997;86(6):975-980. doi:10.3171/ jns.1997.86.6.097
- Viola RW, King HA, Adler SM, Wilson CB. Delayed infection after elective spinal instrumentation and fusion: a retrospective analysis of eight cases. Spine (Phila Pa 1976). 1997;22(20):2444–2451. doi:10.1097/00007632-199710150-00023.
- Weinstein MA, McCabe JP, Cammisa J. Postoperative spinal wound infection: a review of 2,391 consecutive index procedures. J Spinal Disord. 2000;13(5):422–426. doi:10.1097/00002517-200010000-00009
- Mirovsky Y, Floman Y, Smorgick Y, et al. Management of deep wound infection after posterior lumbar interbody fusion with cages. J Spinal Disord
- Tech. 2007;20(2):127-131. doi:10.1097/01.bsd.0000211266.66615.es. Chahoud J, Kanafani Z, Kanj SS. Surgical site infections following spine surgery: eliminating the controversies in the diagnosis. Front Med (Lausanne). 2014;1:7. doi:10.3389/fmed.2014.00007.

Author: Yvonne Achermann

QUESTION 2: Is there a role for the use of oral antibiotic in treatment of acute and chronic spinal infections?

RECOMMENDATION: There may be a role for highly bioavailable oral antibiotics in the treatment of acute and chronic spine infection in select circumstances.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Vertebral osteomyelitis

In vertebral osteomyelitis (spondylodiscitis) without an implant, experts recommend a treatment duration of 6 to 12 weeks [1]. However, a retrospective study over 10 years by Roblot et al. found no difference in relapse rate comparing 6 and 12 weeks of treatment [2]. An open label, non-inferiority, randomized, controlled trial by Bernard et al. firstly showed that 6 weeks was not inferior to 12 weeks. In both groups, intravenous treatment was only given for a median time of 14 to 15 days and was followed by an oral fluoroquinolone and rifampin combination or aminopenicillin (both regimens with high oral bioavailability) [3]. The authors could not see a difference in the proportion of treatment failure between patients given intravenous treatment for more than one week and those for less than one week.

Postsurgical infection with an implant

There are many studies in this field regarding optimal treatment duration and agents in spinal implant-associated infections, but they are all retrospective with low levels of evidence. There are no up-to-date prospective and/or randomized studies published investigating the optimal duration of antibiotic treatment and the role of oral antibiotics in implant-associated spinal infections.

Most studies demonstrated successful treatment of spinal implant-associated infections with a total duration of six weeks [4–6]. If implants are not removed, reported durations of treatment are up to 12 weeks with intravenous treatment for 6 weeks, followed by oral antibiotic treatment for another 6 weeks [7,8].

Yet, regarding duration of intravenous treatment, there are no clear recommendations. Some studies treat intravenously for a prolonged time for up to four [8–10] or six weeks [4,11–13]. But there are also retrospective studies in which intravenous treatment was given for two weeks or less followed by oral antibiotics with good oral bioavailability [14]. Billieres et al. did a multivariate analysis on risk factors for relapse of infection and did not find an association with duration of total or intravenous antibiotic treatment [14]. Another study by Kowalsky et al. also concluded that duration of intravenous treatment is no risk factor for neither acute nor chronic infections [15].

REFERENCES

- Zimmerli W. Clinical practice. Vertebral osteomyelitis. N Engl J Med. 2010;362:1022-1029. doi:10.1056/NEJMcp0910753.
 Roblot F, Besnier JM, Juhel L, Vidal C, Ragot S, Bastides F, et al. Optimal
- [2] Roblot F, Besnier JM, Juhel L, Vidal C, Ragot S, Bastides F, et al. Optimal duration of antibiotic therapy in vertebral osteomyelitis. Semin Arthritis Rheum. 2007;36:269-277. doi:10.1016/j.semarthrit.2006.09.004.
 [3] Bernard L, Legout L, Zürcher-Pfund L, Stern R, Rohner P, Peter R, et al. Six
- Bernard L, Legout L, Zürcher-Pfund L, Stern R, Rohner P, Peter R, et al. Six weeks of antibiotic treatment is sufficient following surgery for septic arthroplasty. J Infect. 2010;61:125–132. doi:10.1016/j.jinf.2010.05.005.
 Roehrborn AA, Hansbrough JF, Gualdoni B, Kim S. Lipid-based slow-release
- [4] Roehrborn AA, Hansbrough JF, Gualdoni B, Kim S. Lipid-based slow-release formulation of amikacin sulfate reduces foreign body-associated infections in mice. Antimicrob Agents Chemother. 1995;39:1752–1755.
- [5] Viola RW, King HA, Adler SM, Wilson CB. Delayed infection after elective spinal instrumentation and fusion. A retrospective analysis of eight cases. Spine. 1997;22:2444-2450; discussion 2450-2451.
- Spine. 1993;22:2444–2450; discussion 2450–2451.

 [6] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913–920. doi:10.1086/512194
- [7] Mirovsky Y, Floman Y, Smorgick Y, Ashkenazi E, Anekstein Y, Millgram MA, et al. Management of deep wound infection after posterior lumbar inter-

- body fusion with cages. J Spinal Disord Tech. 2007;20:127–131. doi:10.1097/01. bsd.0000211266.66615.e5.
- [8] Collins I, Wilson-MacDonald J, Chami G, Burgoyne W, Vineyakam P, Berendt T, et al. The diagnosis and management of infection following instrumented spinal fusion. Eur Spine J. 2008;17:445–450. doi:10.1007/s00586-007-0559-8.
- spinal fusion. Eur Spine J. 2008;17:445–450. doi:10.1007/s00586-007-0559-8.
 [9] Falavigna A, Righesso O, Traynelis VC, Teles AR, da Silva PG. Effect of deep wound infection following lumbar arthrodesis for degenerative disc disease on long-term outcome: a prospective study: clinical article. J Neurosurg Spine. 2011;15:399–403. doi:10.3171/2011.5.SPINE10825.
- [10] Sponseller PD, LaPorte DM, Hungerford MW, Eck K, Bridwell KH, Lenke LG. Deep wound infections after neuromuscular scoliosis surgery: a multicenter study of risk factors and treatment outcomes. Spine. 2000;25:2461-2466
- 2466.
 [11] Chen SH, Lee CH, Huang KC, Hsieh PH, Tsai SY. Postoperative wound infection after posterior spinal instrumentation: analysis of long-term treatment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014-3636-9.
 [12] Chaichana KL, Bydon M, Santiago-Dieppa DR, Hwang L, McLoughlin G,
- [12] Chaichana KL, Bydon M, Santiago-Dieppa DR, Hwang L, McLoughlin G, Sciubba DM, et al. Risk of infection following posterior instrumented lumbar fusion for degenerative spine disease in 817 consecutive cases. J Neurosurg Spine. 2014;20:45–52. doi:10.3171/2013.10.SPINE1364.
- [13] Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19:398–404. doi:10.1007/s00776-014-0545-z.
 [14] Billières J, Uçkay I, Faundez A, Douissard J, Kuczma P, Suvà D, et al. Variables
- [14] Billières J, Uçkay I, Faundez A, Douissard J, Kuczma P, Suvà D, et al. Variables associated with remission in spinal surgical site infections. J Spine Surg. 2016;2:128–134. doi:10.21037/jss.2016.06.06.
- [15] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Osmon DR. Propionibacterium acnes vertebral osteomyelitis: seek and ye shall find? Clin Orthop Relat Res. 2007;461:25–30. doi:10.1097/BLO.ob013e318073c25d.

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QUESTION 3: Is there a role for chronic antibiotic suppression after treating patients with retained infected spinal hardware?

RECOMMENDATION: The use of chronic antibiotic suppression (CAS) has not been clearly investigated until now. However, it can be an option for patients whose implants cannot be removed or who refuse further surgeries because of comorbidities.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Only one study has compared patients receiving CAS [1]. They found that 5 out of 22 patients with CAS had treatment failure, compared with 5 out of 6 in the control group. The definition they used for treatment failure was described as the need for an unanticipated debridement or a clinician's decision to give a second course of antibiotics. Suppressive antibiotics were given for a median time of 303 days (IQR 147 to 672) to patients with early onset infection and 410 days (IQR 61 to 667) to patients with late onset infection. Data on treatment failure was reported only for early onset infection patients. It could be argued that patients already under CAS would not have been eligible for a second course of antibiotic treatment and this could partly increase the rates of treatment failure on the group without CAS, biasing the study results.

Other studies reporting on antibiotic treatments show large variations in the duration of treatment. Miyazaki et al. reported a mean duration of oral treatment of 336 days, ranging from 89 to 1,673 days [2]. Their study focused on multi-resistant surgical site infection treated with implant retention. Maruo et al. reported an average duration of antibiotic treatment of 255.8 days with a standard devia-

tion of 283.4 days [3]. All these reports show a huge variation in the length of antibiotic treatment, with a select group of patients in each study receiving CAS. Decision for prolonged CAS was made at the clinician's discretion and based on the patient's symptoms, so there is no particular setting in which it would be possible to offer a sound recommendation. Besides the mentioned paper by Kowalski, there are no reports comparing CAS with other treatment regimes.

REFERENCES

- [1] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44(7):913–920.
- [2] Miyazaki S, Kakutani K, Maeno K, Takada T, Yurube T, Kurosaka M, Nishida K. Surgical debridement with retention of spinal instrumentation and long-term antimicrobial therapy for multidrug-resistant surgical site infections after spinal surgery: a case series. Int Orthop. 2016;40(6):1171-1177.
- [3] Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19(3):398–404.

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QUESTION 4: Is there a role for combination antibiotics (i.e., dual or triple) in treating patients with surgical site infection (SSI) following spinal surgery?

RECOMMENDATION: There is insufficient evidence to recommend the routine use of combination antibiotics in the setting of postoperative spine infections. However, there may be a role for combination antibiotics in certain circumstances related to specific pathogens.

LEVEL OF EVIDENCE: Consensus

Spine

DELEGATE VOTE: Agree: 87%, Disagree: 13%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

The incidence of postoperative spine infection has been reported as between 0.7 and 16%, with higher rates noted in procedures with hardware implantation [1,2]. The most common organisms isolated are Staphylococcus aureus, Staphylococcus epidermidis, methicillinresistant S. aureus and Enterococcus. Up to 20 to 30% of infections are noted to be poly-microbial [3,4].

Antibiotic treatment is directed at the isolated micro-organism/s and usually only a single anti-microbial agent is used. There are a few reports of dual antibiotic therapy with rifampin, the most common additive agent [3,5]. Rifampin is chosen due to its ability to penetrate biofilms associated with implant-related infections [6]. Evidence from a mouse model has shown that the addition of rifampin to vancomycin led to an increase in bacterial death, but no change in the final outcome from the SSI [7]. There are no clinical studies comparing the use of single to multi-agent antibiotic therapy for postoperative spine infections.

REFERENCES

- Fang A, Hu SS, Endres N, Bradford DS. Risk factors for infection after spinal surgery. Spine. 2005;30:1460-1465.
- Parchi PD, Evangelisti G, Andreani L, Girardi F, Darren L, Sama A, et al. Postoperative Spine Infections. Orthop Rev (Pavia). 2015;7:5900. doi:10.4081/ ог.2015.5900
- Billières J, Uçkay I, Faundez A, Douissard J, Kuczma P, Suvà D, et al. Variables associated with remission in spinal surgical site infections. J Spine Surg. 2016;2:128-134. doi:10.21037/jss.2016.06.06.
 Weinstein MA, McCabe JP, Cammisa FP. Postoperative spinal wound
- infection: a review of 2,391 consecutive index procedures. J Spinal Disord.
- Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913-920.
- doi:10.1086/512194.
 Zheng Z, Stewart PS. Penetration of rifampin through Staphylococcus epidermidis biofilms. Antimicrob Agents Chemother. 2002;46:900-903
- Hu Y, Hegde V, Johansen D, Loftin AH, Dworsky E, Zoller SD, et al. Combinatory antibiotic therapy increases rate of bacterial kill but not final outcome in a novel mouse model of Staphylococcus aureus spinal implant infection. PLoS ONE. 2017;12:e0173019. doi:10.1371/journal.pone.0173019.

Author: Yvonne Achermann

QUESTION 5: How long should antibiotics be administered after surgical debridement for an acute postsurgical spinal infection?

RECOMMENDATION: For vertebral osteomyelitis: Initial intravenous treatment for one to two weeks, followed by an oral treatment of four to five weeks to reach a total treatment duration of six weeks.

For deep surgical site infections: There is limited knowledge about the ideal duration of antibiotic treatment and which intravenous and/or oral agents should be given. As extrapolated from studies in periprosthetic joint infections (PJIs) and retrospective studies in spine infections, 12 weeks of antibiotic treatment can be recommended in cases with early infection and implant retention, six weeks if the implant is removed and prolonged suppressive treatment in delayed infections without removal of the implant.

LEVEL OF EVIDENCE: Moderate for vertebral osteomyelitis. Limited for surgical site infections after spine surgery

DELEGATE VOTE: Agree: 80%, Disagree: 13%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Vertebral Osteomyelitis

In vertebral osteomyelitis (spondylodiscitis) without an implant, experts recommend a treatment duration of 6 to 12 weeks [1]. However, a retrospective study over 10 years by Roblot et al. [2] found no difference in relapse rate between 6 and 12 weeks of treatment [2]. An open label, non-inferiority, randomized, controlled trial by Bernard et al. first showed that 6 weeks was not inferior to 12 weeks. In both groups, intravenous treatment was only given for a median time of 14 to 15 days followed by an oral fluoroquinolone and rifampin combination or aminopenicillin (both regimens with high oral bioavailability) [3]. The authors could not see a difference in the proportion of treatment failure between patients given intravenous treatment for more than one week and those for less than one week.

Postsurgical infection with an implant

There are many studies in this field regarding optimal treatment duration and agents in spinal implant-associated infections, but they are all retrospective with low levels of evidence. There are no up-to-date prospective and/or randomized studies published investigating the optimal duration of antibiotic treatment and the role of oral antibiotics in implant-associated spinal infections.

Most studies demonstrated successful treatment of spinal implant-associated infections with a total duration of six weeks [4–6]. If implants are not removed, reported durations of treatment are up to 12 weeks with intravenous treatment for six weeks, followed by oral antibiotic treatment for another six weeks [7,8].

Yet, regarding duration of intravenous treatment, there are no clear recommendations. Some studies treat intravenously for a prolonged time for up to four [8-10] or six weeks [4,11-13]. But there are also retrospective studies in which intravenous treatment was given for two weeks or less followed by oral antibiotics with good oral bioavailability [14]. Billieres et al. did a multivariate analysis on risk factors for relapse of infection and did not find an association with duration of total or intravenous antibiotic treatment [14]. Another study by Kowalsky et al. also concluded that duration of intravenous treatment is not a risk factor for acute chronic infections [15].

REFERENCES

- Zimmerli W. Clinical practice. Vertebral osteomyelitis. N Engl J Med. 2010;362(11): 1022-1029
- Roblot F, Besnier JM, Juhel L, et al. Optimal duration of antibiotic therapy in vertebral osteomyelitis. Semin Arthritis Rheum. 2007;36(5):269–277.

- Bernard L, Legout L, Zurcher-Pfund L, et al. Six weeks of antibiotic treatment is sufficient following surgery for septic arthroplasty. J Infect. 2010;61(2):125-
- Roehrborn AA, Hansbrough JF, Gualdoni B, Kim S. Lipid-based slow-release formulation of amikacin sulfate reduces foreign body-associated infections in mice. Antimicrobi Agents Chemother. 1995;39(8):1752–1755. Viola RW, King HA, Adler SM, Wilson CB. Delayed infection after elective
- spinal instrumentation and fusion. A retrospective analysis of eight cases.
- Spine. 1997;22(20):2444-2450; discussion 50-51. Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44(7):913-
- Mirovsky Y, Floman Y, Smorgick Y, et al. Management of deep wound infection after posterior lumbar interbody fusion with cages. J Spinal Disord Tech. 2007;20(2):127-131.
- Collins I, Wilson-MacDonald J, Chami G, et al. The diagnosis and management of infection following instrumented spinal fusion. Eur Spine J.
- 2007;17(3):445-450. Falavigna A, Righesso O, Traynelis VC, Teles AR, da Silva PG. Effect of deep wound infection following lumbar arthrodesis for degenerative disc disease on long-term outcome: a prospective study: clinical article. J Neuro-
- surg Spine. 2011;15(4):399–403. Sponseller PD, LaPorte DM, Hungerford MW, Eck K, Bridwell KH, Lenke LG. Deep wound infections after neuromuscular scoliosis surgery: a multicenter study of risk factors and treatment outcomes. Spine. 2000;25(19):2461–2466. Chen SH, Lee CH, Huang KC, Hsieh PH, Tsai SY. Postoperative wound infec-
- tion after posterior spinal instrumentation: analysis of long-term treat-
- ment outcomes. Eur Spine J. 2014;24(3):561–570. Chaichana KL, Bydon M, Santiago-Dieppa DR, et al. Risk of infection following posterior instrumented lumbar fusion for degenerative spine disease in 817 consecutive cases. | Neurosurg Spine. 2013;20(1):45–52.
- Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J
- Orthop Sci. 2014;19(3):398-404. Billieres J, Uckay I, Faundez A, et al. Variables associated with remission in
- spinal surgical site infections. J Spine Surg. 2016;2(2):128–134. Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Osmon DR. *Propionibacterium acnes* vertebral osteomyelitis: seek and ye shall find? Clin Orthop Relat Res. 2007;461:25-30.

Authors: Gregory Schroeder, Mayan Lendner

QUESTION 6: How long should antibiotics be continued when spinal wounds are left to heal by secondary intention?

RECOMMENDATION: Only standard perioperative antibiotic prophylaxis is recommended.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Following spine surgery, surgical wounds are normally closed via primary intention where all tissue is fastened closed with sutures, staples, glue or some other form of closure material. In rare cases, however, wounds are left to close naturally via secondary intention. Normally, this is done in cases where the risk of persistence of infection is high or when a large gap in soft tissue exists as a result of tissue loss.

Antibiotic prophylaxis has been shown to be useful in preventing infection following spine surgery. However, no specific agent or schedule has been identified as superior over any other [1].

In a randomized, blinded, controlled study, Gupta et al. found that topical antibiotics, specifically sucralfate, increased wound healing in patients at four weeks following hemorrhoidectomy left to heal via secondary intention when compared to placebo (78% compared to 52%) [2]. In contrast, Doung et al. found that the use of trimethoprim-sulfamethoxazole in pediatric skin abscess treatment compared to placebo did not significantly affect the recurrence of new lesions in the long term [3].

A systematic review by Norman et al. found that no robust evidence exists on the relative effectiveness of any antibiotic preparation in cases where surgical wounds have been left to heal by secondary intention [4]. There is no high-level evidence directly related to spine surgery for this topic. In general, if there is hardware present, patients often should receive at least six weeks of intravenous antibiotics and continued suppressive antibiotics until the wound heals.

REFERENCES

Watters WC, Baisden J, Bono CM, Heggeness MH, Resnick DK, Shaffer WO, Toton JF. Antibiotic prophylaxis in spine surgery: an evidence-based clinical guideline for the use of prophylactic antibiotics in spine surgery. Spine J. 2009;9(2):142–146. doi:10.1016/j.spinee.2008.05.008.

- [2] Gupta PJ, Heda PS, Kalaskar S, Tamaskar VP. Topical sucralfate decreases pain after hemorrhoidectomy and improves healing: a randomized, blinded, controlled study. Dis Colon Rectum. 2008;51(2):231–234. doi:10.1007/s10350-007-0092-4.
- [3] Duong M, Markwell S, Peter J, Barenkamp S. Randomized, controlled trial of antibiotics in the management of community-acquired skin abscesses
- in the pediatric patient. Ann Emerg Med. 2010;55(5):401–407. doi:10.1016/j. annemergmed.2009.03.014
- [4] Norman G, Dumville JC, Mohapatra DP, Owens GL, Crosbie EJ. Antibiotics and antiseptics for surgical wounds healing by secondary intention. Cochrane Database of Syst Rev. 2016;3. doi:10.1002/14651858.cdo11712. pub2.

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QUESTION 7: What is the optimal duration of antibiotic treatment following spine infection in patients within whom hardware is retained? Is the antibiotic treatment different for those with spine infection without hardware?

RECOMMENDATION: There are no case-control studies allowing for an evidence-based recommendation on the optimal length of antibiotic treatment following spine infections in the presence of retained hardware. The most commonly implemented antibiotic regime is three months. However, duration of treatment was highly variable among all studies. Patients with non-instrumented surgeries did well with a shorter course of antibiotics.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

After searching PubMed, CINAHL and Embase (with MeSH terms "surgical site infection," "spine" and "antibiotic") and reviewing 381 abstracts, a final 14 studies included treatment of spinal surgical site infection (SSI) with retained implants (including data on antibiotic treatment regimens) [1-14]. There were no studies analyzing or comparing different antibiotic regimes. Most of these studies were retrospective in nature, however one study was a prospective observational study. There were no studies comparing different antibiotic treatment regimens. There was also a wide variation in the duration of treatment among the studies ranging from 42 to 597 days in 1 study, and ranging between 89 and 1,673 days in a separate study [9,11]. These variations were usually related to treatment failure or poor control of the infection. Of 14 studies, 7 reported mean antibiotic treatments of 12 weeks or 3 months [3–6,10,13,14]. All but three studies reported on time of intravenous (IV) and oral antibiotics. The most reported mean time for IV antibiotic administration was an average of four to eight weeks in eight studies. One study reported on 81 SSIs, of which 39 were treated with suppressive antimicrobial therapy [2]. At final two-year follow-up, seven patients were still under antibiotic treat-

Three studies reported data on patients with early and late infection [2,5,10]. Also, there were significant variations regarding the onset of infection. Some studies only reported ranges and gave no mean or median values. Of the nine studies with available mean data, mean time to onset of infection was 103.2 days. Removing an outlier with 778 days for late infection, mean time to onset of infection was 18.98 days (range of mean values was 2.9 to 54)

There was only one retrospective study analyzing the antibiotic treatment regimen in a series of 74 patients, all with implant removal (IR) [15]. Patients had a median duration of IV antibiotics of four weeks and an additional five weeks of oral antibiotic treatment. There were no comparative studies regarding different antibiotic regimen.

Regarding IR, there were two very different settings in which implants had to be removed. Of 729 SSI cases recorded in the 15 studies, implants were removed in 195 patients (26.74%). In 114 cases (15.6%), IR was performed as part of SSI treatment during the

first debridement procedure. In the remaining 81 cases (11.11%), IR was performed because of treatment failure after several debridement procedures. The fact that IR can be split into two differentiated groups makes it more difficult to compare treatment regimes. Usually, when IR was performed as the initial treatment, antibiotic regimens tended to be shorter [15]. On the other hand, when IR was performed because of treatment failure, antibiotic treatments were longer.

With regards to non-instrumented spine surgeries, Maruo et al. compared 59 non-instrumented infections with 166 instrumented cases [8]. They reported longer antibiotic treatment for instrumented cases (mean 40 days IV vs. 25.4 in non-instrumented and mean 255 days oral vs. 42). Only 10% of the non-instrumented cases needed more than one debridement compared to 28% for instrumented spine procedures. Of the non-instrumented spine surgeries, 20% were successfully treated without surgical debridement compared to only 6% of instrumented spine procedures.

REFERENCES

- [1] Falavigna A, Righesso Neto O, Fonseca GP, Nervo M. [Management of deep wound infections in spinal lumbar fusions]. Arq Neuropsiquiatr. 2006;64:1001–1004.
- [2] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913–920. doi:10.1086/512194.
- [3] Mirovsky Y, Floman Y, Smorgick Y, Ashkenazi E, Anekstein Y, Millgram MA, et al. Management of deep wound infection after posterior lumbar interbody fusion with cages. J Spinal Disord Tech. 2007;20:127–131. doi:10.1097/01. bsd.0000211266.66615.e5.
- [4] Hong HS, Chang MC, Liu CL, Chen TH. Is aggressive surgery necessary for acute postoperative deep spinal wound infection? Spine. 2008;33:2473–2478. doi:10.1097/BRS.0b013e3181894ffo.
- [5] Sierra-Hoffman M, Jinadatha C, Carpenter JL, Rahm M. Postoperative instrumented spine infections: a retrospective review. South Med J. 2010;103:25–30. doi:10.1097/SMJ.0b013e3181c4e00b.
- [6] Dubée V, Lenoir T, Leflon-Guibout V, Briere-Bellier C, Guigui P, Fantin B. Three-month antibiotic therapy for early-onset postoperative spinal implant infections. Clin Infect Dis. 2012;55:1481-1487. doi:10.1093/cid/cis769.
- [7] Ahmed R, Greenlee JDW, Traynelis VC. Preservation of spinal instrumentation after development of postoperative bacterial infections in patients undergoing spinal arthrodesis. J Spinal Disord Tech. 2012;25:299–302. doi:10.1097/BSD.0b013e31821fbf72.

- Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19:398–404. doi:10.1007/s00776-014-0545-z. Messina AF, Berman DM, Ghazarian SR, Patel R, Neustadt J, Hahn G, et al.
- The management and outcome of spinal implant-related infections in pediatric patients: a retrospective review. Pediatr Infect Dis J. 2014;33:720–723. doi:10.1097/INF.000000000000264. Chen SH, Lee CH, Huang KC, Hsieh H, Tsai SY. Postoperative wound infec-
- tion after posterior spinal instrumentation: analysis of long-term treatment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014:3636-9. Miyazaki S, Kakutani K, Maeno K, Takada T, Yurube T, Kurosaka M, et al.
- Surgical debridement with retention of spinal instrumentation and longterm antimicrobial therapy for multidrug-resistant surgical site infections after spinal surgery: a case series. Int Orthop. 2016;40:1171-1177. doi:10.1007/ soo264-015-3073-3.
- Billières J, Uçkay I, Faundez A, Douissard J, Kuczma P, Suvà D, et al. Variables associated with remission in spinal surgical site infections. J Spine Surg. 2016;2:128-134. doi:10.21037/jss.2016.06.06.
- Wille H, Dauchy FA, Desclaux A, Dutronc H, Vareil MO, Dubois V, et al. Efficacy of debridement, antibiotic therapy and implant retention within
- three months during postoperative instrumented spine infections. Infect Dis (Lond). 2017;49:261–267. doi:10.1080/23744235.2016.1255351.

 Takizawa T, Tsutsumimoto T, Yui M, Misawa H. Surgical site infections caused by methicillin-resistant Staphylococcus epidermidis after spinal instrumentation surgery. Spine. 2017;42:525-530. doi:10.1097/ BRS.0000000000001792.
- Collins I, Wilson-MacDonald J, Chami G, Burgoyne W, Vineyakam P, Berendt T, et al. The diagnosis and management of infection following instrumented spinal fusion. Eur Spine J. 2008;17:445–450. doi:10.1007/s00586-007-0559-8.

Author: Maja Babic

QUESTION 8: What tests should be used to monitor response to antibiotic treatment in patients with spine infection?

RECOMMENDATION: Serum C-reactive protein (CRP) levels are closely related to clinical response in spine infections and are therefore the preferred marker in monitoring the therapeutic course.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

In two large retrospective studies including 363 patients, criteria for discontinuation of treatment included CRP normalization in addition to resolution of clinical symptoms [1,2]. A weekly decrease of CRP by 50% has been suggested as a therapeutic response in the retrospective study population [3].

Lack of normalization of serum CRP levels is a predictor of treatment failure and warrants additional evaluation, as demonstrated both by a retrospective cohort including 79 patients and a prospective study including 21 patients followed for postsurgical wound infections of the spine [4-5].

Moreover, in a retrospective analysis of 61 patients treated for bacterial spondylodiscitis, the only predictor for de-escalating intravenous therapy to highly bioavailable oral agents was a CRP decrease by week 2 of therapy [6].

REFERENCES

- Legrand E, Flipo RM, Guggenbuhl P, Masson C, Maillefert JF, Soubrier M, et al. Management of nontuberculous infectious discitis. Treatments used in 110 patients admitted to 12 teaching hospitals in France. Joint Bone Spine.
- 2001;68:504–509.
 McHenry MC, Easley KA, Locker GA. Vertebral osteomyelitis: long-term outcome for 253 patients from 7 Cleveland-area hospitals. Clin Infect Dis.
- Legrand E, Massin P, Levasseur R, Hoppé E, Chappard D, Audran M. Stratégie diagnostique et principes thérapeutiques au cours des spondylodiscites infectieuses bactériennes. Revue Du Rhumatisme. 2006;73:373-379.
- doi:10.1016/j.rhum.2006.01.005. Khan MH, Smith PN, Rao N, Donaldson WF. Serum C-reactive protein levels correlate with clinical response in patients treated with antibiotics for wound infections after spinal surgery. Spine J. 2006;6:311–315. doi:10.1016/j. spinee.2005.07.006.
- Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Osmon DR. Do follow-up imaging examinations provide useful prognostic informa-tion in patients with spine infection? Clin Infect Dis. 2006;43:172-179. doi:10.1086/505118.
- Babouee Flury B, Elzi L, Kolbe M, Frei R, Weisser M, Schären S, et al. Is switching to an oral antibiotic regimen safe after 2 weeks of intravenous treatment for primary bacterial vertebral osteomyelitis? BMC Infect Dis. 2014;14:226. doi:10.1186/1471-2334-14-226.

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QUESTION 9: Which is the best alternative antimicrobial therapy for fluoroguinolone-resistant gram-negative acute post-surgical infection in spinal surgery?

RECOMMENDATION: The choice of antimicrobial therapy should be based on the pathogen and the susceptibility profile.

LEVEL OF EVIDENCE: Moderate

DELEGATE VOTE: Agree: 93%, Disagree: 7%, Abstain: 0% (Super Majority, Strong Consensus)

RATIONALE

Currently, over 30% of all spinal surgical site infections (SSIs) are secondary to gram-negative bacteria (GNB). Focusing on acute postsurgical infection of spinal surgery, there is no published experience regarding the best therapeutic strategies in case infection by GNB resistant to quinolones. Thus, the treatment criteria used in these cases are the same as those used in the case of fluoroquinolone-resistant GNB periprosthetic join infections (PJIs). The importance of using fluoroquinolones in acute PJIs due to gram-negative bacilli has been demonstrated, but limited antimicrobial agents are available in the case of implant-associated infections caused by fluoroquinolone-resistant GNB [1–3].

The most commonly used antibiotics in the event of fluoroquinolone resistance are β -lactams and carbapenems with or without anti-pseudomonal activity [4]. Grossi et al. described the outcome of 76 GNB-PJIs managed with a curative intent and in their experience, intravenous β-lactams throughout treatment duration (median 90 days) results in an effective alternative to fluoroquinolones [5].

Therapeutic alternatives to β -lactams have been poorly assessed. Cotrimoxazole, which can be switched to oral therapy, has been successfully used in some of these cases [1–6]. Other possible alternatives are the "recovery" of the use of less conventional antibiotics, such as colistin and fosfomycin [7–9]. Colistin shows good spread in bacterial biofilm and a synergistic effect when combined with other antibiotics, especially β-lactams, and has been demonstrated to be effective in vitro against P. aeruginosa and enterobacteria [7]. Corvec et al. compared the activities of fosfomycin, tigecycline, colistin and gentamicin (alone and in combination), against a CTX-M15-producing strain of Escherichia coli in vitro and in a foreign-body infection model [10]. Fosfomycin was the only single agent, which was able to eradicate *E. coli* biofilms (cure rate, 17% of implanted, infected cages). In combination, colistin plus tigecycline (50%) and fosfomycin plus gentamicin (42%) cured significantly more infected cages than colistin plus gentamicin (33%) or fosfomycin plus tigecycline (25%) (p < 0.05). The combination of fosfomycin plus colistin showed the highest cure rate (67%), which was significantly better than that of fosfomycin alone (p < 0.05). Therefore, the authors conclude that the combination of fosfomycin plus colistin is a promising treatment option for implant-associated infections caused by fluoroquinolone-resistant GNB, but the effectiveness of this combination should be assessed in vivo.

Other potential therapeutic alternatives are combinations that include tigecycline or rifampin for their demonstrated in vitro synergism with several drugs. Tigecycline has been used for carbapenemase-producing gram-negative PJIs, although bone concentrations of the drug are usually lower than the minimum inhibitory concentrations of these bacteria [11]. Drapeau et al. recently described a literature review of 19 clinical studies on the use of rifampin in treatments for multidrug resistant gramnegative (MDRGN) bacterial infection [12]. Nonetheless, the real clinical benefit of using rifampin-containing therapies for MDRGN bacteria in terms of clinical outcome and survival rates remains to be defined.

The development of new agents (ceftazidime/avibactam, aztreonam/avibactam, cefiderocol, ceftolozane/tazobactam) with activity against MDRGN bacteria will provide important therapeutic options for clinicians, but definitive data showing clinical efficacy is currently lacking [13].

The efficacy of intrawound tobramycin powder in terms of eradicating a known bacterial contamination in an Escherichia coliinfected rabbit spinal implantation model was assessed, with the researchers concluding that intrawound tobramycin eliminated Escherichia coli surgical site contamination [14].

REFERENCES

- Rodríguez-Pardo D, Pigrau C, Lora-Tamayo J, Soriano A, del Toro MD, Cobo J, et al. Gram-negative prosthetic joint infection: outcome of a debridement, antibiotics and implant retention approach. A large multicentre study. Clin Microbiol Infect. 2014;20:0911-0919. doi:10.1111/1469-0691.12649.
- Jaén N, Martínez-Pastor JC, Muñoz-Mahamud E, García-Ramiro S, Bosch J, Mensa J, et al. Long-term outcome of acute prosthetic joint infections due to gram-negative bacilli treated with retention of prosthesis. Rev Esp
- Quimioter. 2012;25:194–198. Lee CY, Wu MH, Cheng CC, Huang TJ, Huang TY, Lee CY, et al. Comparison of gram-negative and gram-positive hematogenous pyogenic spondylodiscitis: clinical characteristics and outcomes of treatment. BMC Infect Dis. 2016;16:735. doi:10.1186/s12879-016-2071-4
- Widmer AF, Frei R, Rajacic Z, Zimmerli W. Correlation between in vivo and in vitro efficacy of antimicrobial agents against foreign body infections. J Infect Dis. 1990;162:96–102.
- Grossi O, Asseray N, Bourigault C, Corvec S, Valette M, Navas D, et al. Gramnegative prosthetic joint infections managed according to a multidisci-plinary standardized approach: risk factors for failure and outcome with and without fluoroquinolones. J Antimicrob Chemother. 2016;71:2593–2597. doi:10.1093/jac/dkw202.
- Martínez-Pastor JC, Muñoz-Mahamud E, Vilchez F, García-Ramiro S, Bori G, Sierra J, et al. Oútcome of acute prosthetic joint infections due to gramnegative bacilli treated with open debridement and retention of the prosthesis. Antimicrob Agents Chemother. 2009;53:4772-4777. doi:10.1128/ AAC.00188-00.
- Lora-Tamayo J, Murillo O, Bergen PJ, Nation RL, Poudyal A, Luo X, et al. Activity of colistin combined with doripenem at clinically relevant concentrations against multidrug-resistant Pseudomonas aeruginosa in an in vitro dynamic biofilm model. J Antimicrob Chemother. 2014;69:2434-2442. doi:10.1093/jac/dku151.
- Falagas ME, Kastoris AC, Kapaskelis AM, Karageorgopoulos DE. Fosfomycin for the treatment of multidrug-resistant, including extended-spectrum beta-lactamase producing, Enterobacteriaceae infections: a systematic review. Lancet Infect Dis. 2010;10:43–50. doi:10.1016/S1473-3099(09)70325-1.
- Falagas ME, Kastoris AC, Karageorgopoulos DE, Rafailidis PI. Fosfomycin for the treatment of infections caused by multidrug-resistant non-fermenting gram-negative bacilli: a systematic review of microbiological, animal and clinical studies. Int J Antimicrob Agents. 2009;34:111–120. doi:10.1016/j.ijantimicag.2009.03.009
- Corvec S, Furustrand Tafin U, Betrisey B, Borens O, Trampuz A. Activities of fosfomycin, tigecycline, colistin, and gentamicin against extendedspectrum-β-lactamase-producing Escherichia coli in a foreign-body infection model. Antimicrob Agents Chemother. 2013;57:1421-1427. doi:10.1128/ AAC.01718-12.
- de Sanctis J, Teixeira L, van Duin D, Odio C, Hall G, Tomford JW, et al. Complex prosthetic joint infections due to carbapenemase-producing Klebsiella pneumoniae: a unique challenge in the era of untreatable infections. Int J Infect Dis. 2014;25:73–78. doi:10.1016/j.ijid.2014.01.028.
- Drapeau CMJ, Grilli E, Petrosillo N. Rifampicin combined regimens for gram-negative infections: data from the literature. Int J Antimicrob Agents.
- 2010;35:39–44. doi:10.1016/j.ijantimicag.2009.08.011. Wright H, Bonomo RA, Paterson DL. New agents for the treatment of infections with gram-negative bacteria: Restoring the miracle or false dawn? Clin Microbiol Infect. 2017;23:704–712. doi:10.1016/j.cmi.2017.09.001. Laratta JL, Shillingford JN, Hardy N, Lombardi JM, Saifi C, Romanov A, et
- al. Intrawound tobramycin powder eradicates surgical wound contamination: an in vivo rabbit study. Spine. 2017;42:E1393-E1397. doi:10.1097/ BRS.0000000000002187.

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QUESTION 10: Is there a difference in the efficacy of vancomycin beads versus vancomycin powder for spinal implant infections?

RECOMMENDATION: It is unclear whether there is a difference in the efficacy of vancomycin beads versus vancomycin powder for spinal implants infections.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Currently, there are no studies comparing or individually evaluating the efficacy of vancomycin powder and vancomycin beads for the

treatment of infections following spinal instrumentation.

3.3. TREATMENT: IMPLANTS

Authors: Pouya Alijanipour, Caroline Granger

QUESTION 1: Should a cage be removed in patients with postoperative spine infection?

RECOMMENDATION: No. The interbody cage can be maintained in the absence of clinical and radiographic signs of loosening or displacement of the cage or compression on neural and vascular structures. However, the cage should be removed if the infection persists despite salvage attempts consisting of irrigation and debridement procedures combined with intravenous antibiotic treatment.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 73%, Disagree: 0%, Abstain: 27% (Super Majority, Strong Consensus)

RATIONALE

The incidence of surgical site infection in the presence of an interbody cage depends on various factors including the type of approach (anterior, posterior or lateral) and whether the cage is stand-alone or associated with posterolateral instrumentation fusion. Series with stand-alone posterior lumbar interbody fusion (PLIF) or anterior lumbar interbody fusion (ALIF) have lower infection rates (up to 3%) compared to those with long constructs in degenerative adult scoliosis (up to 11%) [1]. On the other hand, adding interbody fusion to posterolateral spinal fusion can be a risk factor for infection and a series of posterolateral fusion with interbody fusion reported higher incidence of surgical site infection compared to those without interbody fusion, most probably due to prolonged surgical procedure, increased blood loss and tissue damage associated with interbody fusion (0.3% versus 1.4%) [2].

Spondylodiscitis at the site of an interbody fusion can present with or without signs of superficial wound infection. If superficial infection does not exist, deep infection can be underestimated or ignored initially due to late presentation. In one report, the average time to diagnosis for spondylitis in patients with PLIF was 164.5 days (range 10-410 days) and time to diagnosis longer than three months was the only predictive factor of failure of intravenous antibiotic treatment and need for implant removal [3]. Moreover, the intervertebral disc tissue is a naturally avascular tissue, limiting the efficiency of immune response as well as efficiency of antibiotics for eradication of infection. Delayed treatment of cage infection can be associated with the risk of extension of infection to the neural elements as well as to the vital retroperitoneal organs and major vessels with disastrous consequences [4].

Cage removal is associated with a risk of interbody space collapse, foraminal narrowing, loss of alignment, progression of deformity, loss of fixation, instability and pseudoarthrosis [5]. On the other hand, inappropriate cage retention can establish bacterial colonization and biofilm formation on the surface of the implants, and diminishes the efficacy of antibiotic treatment [6]. Time of presentation (early versus late postoperative infection), chronicity and severity of symptoms are other considerable factors [7,8].

According to the published case series, in most cases of interbody cage infection, the cage can successfully be retained with an initial salvage attempt consisting of irrigation and debridement procedures combined with antibiotic treatment [1,9–15]. Although, there is no agreed definition criteria for failure of salvage treatment, the following conditions have been considered as indication of cage removal: presence of discitis, osteomyelitis, signs of cage loosening, epidural abscess, extension of infection to soft tissues and presence of bone loss [1,4,8]. Most of these criteria are based on the findings of advanced imaging such as computed tomography and magnetic resonance imaging. One study presented 10 cases with uncontrolled infection of interbody cage, all of which were placed via posterior approaches. In 9 out of 10 cases, solid bone fusion was achieved via an anterior procedure consisting of cage removal and the use of autogenous iliac bone graft to fill the interbody space [16]. An anterior approach for removal of a posteriorly-placed interbody cage prevents complications associated with epidural scar tissue and fibrosis due to the inflammatory response to the original surgery and infection process [16].

REFERENCES

- Pappou IP, Papadopoulos EC, Sama AA, Girardi FP, Cammisa FP. Postoperative infections in interbody fusion for degenerative spinal disease. Clin Orthop Relat Res. 2006;444:120-128. doi:10.1097/01.blo.0000203446.06028.b5.
- Ahn DK, Park HS, Choi DJ, Kim TW, Chun TH, Yang JH, et al. The difference of surgical site infection according to the methods of lumbar fusion surgery. J
- Spinal Disord Tech. 2012;25:E230–E234. doi:10.1097/BSD.0b013e31825c6f7b. Lee JS, Ahn DK, Chang BK, Lee JI. Treatment of surgical site infection in posterior lumbar interbody fusion. Asian Spine J. 2015;9:841-848. doi:10.4184/ asj.2015.9.6.841.
- Carmouche JJ, Molinari RW. Epidural abscess and discitis complicating instrumented posterior lumbar interbody fusion: a case report. Spine.
- Kim JI, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infection after instrumented spinal fusion. J Spinal Disord Tech. 2010;23:258–265. doi:10.1097/BSD.obo13e3181a9452c. Arnold WV, Shirtliff ME, Stoodley P. Bacterial biofilms and periprosthetic
- infections. J Bone Joint Surg Am. 2013;95:2223-2229.

- Hedequist D, Haugen A, Hresko T, Emans J. Failure of attempted implant retention in spinal deformity delayed surgical site infections. Spine. 2009;34:60–64. doi:10.1097/BRS.obo13e31818ed75e
- Kanayama M, Hashimoto T, Shigenobu K, Oha F, Iwata A, Tanaka M. MRIbased decision making of implant removal in deep wound infection after instrumented lumbar fusion. Clin Spine Surg. 2017;30:E99–E103. doi:10.1097/ BSD.obo13e3182aa4c72
- Mirovsky Y, Floman Y, Smorgick Y, Ashkenazi E, Anekstein Y, Millgram MA, et al. Management of deep wound infection after posterior lumbar interbody fusion with cages. J Spinal Disord Tech. 2007;20:127–131. doi:10.1097/01. bsd.0000211266.66615.e5
- Okuyama K, Abe E, Suzuki T, Tamura Y, Chiba M, Sato K. Posterior lumbar interbody fusion: a retrospective study of complications after facet joint excision and pedicle screw fixation in 148 cases. Acta Orthop Scand.
- 1999;70:329–334. Ray CD. Threaded titanium cages for lumbar interbody fusions. Spine. 1997;22:667-679; discussion 679-680.
- Mehbod AA, Ogilvie JW, Pinto MR, Schwender JD, Transfeldt EE, Wood KB, et al. Postoperative deep wound infections in adults after spinal fusion: management with vacuum-assisted wound closure. J Spinal Disord Tech.
- Elias WJ, Simmons NE, Kaptain GJ, Chadduck JB, Whitehill R. Complica-tions of posterior lumbar interbody fusion when using a titanium threaded cage device. J Neurosurg. 2000;93:45–52. Rosenberg WS, Mummaneni PV. Transforaminal lumbar interbody fusion:
- technique, complications, and early results. Neurosurgery. 2001;48:569–574;
- discussion 574-575. Hee HT, Castro FP, Majd ME, Holt RT, Myers L. Anterior/posterior lumbar fusion versus transforaminal lumbar interbody fusion: analysis of compli-cations and predictive factors. J Spinal Disord. 2001;14:533–540.
- Ha KY, Kim YH. Postoperative spondylitis after posterior lumbar interbody fusion using cages. Eur Spine J. 2004;13:419-424. doi:10.1007/s00586-003-0584-1.

Authors: Christopher Kepler, Barrett Boody

QUESTION 2: Is there a length of time of infection beyond which instrumentation should be removed?

RECOMMENDATION: The data suggests that early infection can commonly be treated with implant retention and debridement followed by intravenous (IV) antibiotics and common oral antibiotic treatment. If the patient has achieved spinal fusion, the implants can be safely removed. In the setting of pseudarthrosis, thought should be given to removal of implants to eradicate infection followed by re-instrumentation.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

The primary goals of treating postoperative spinal surgical site infections (SSIs) are to eradicate the infection, maintain stability and achieve fusion (when warranted). While the decision to retain existing instrumentation in the setting of an acute infection may be necessary for maintaining stability or promoting fusion, this may jeopardize the surgeon's ability to completely eradicate the SSI. The preponderance of available evidence suggests the ability to both retain hardware and successfully eradicate the infection depends on the acuity of the presentation, with early diagnoses of SSI (within 30 to 90 days after index procedure) having higher rates of successful retention after debridement and IV antibiotics, while deep infections over one year commonly require removal.

Several studies have demonstrated successful eradication of infection with debridement and hardware retention for earlyonset SSI. Patel et al. reviewed surgical debridement and retention of instrumentation in 17 patients with SSI after spinal arthrodesis ranging from 1 to 6 weeks after the index procedure, noting eradication of infection in all patients and successful fusion in 15 of 17 (88.2%) [1]. Sierra-Hoffman et al. reported successful instrumentation retention with early onset (< 30 days) SSIs with debridement and longterm antibiotics alone, noting eradication of infection in 17 out of 19 (89.5%) patients. However, six of the seven late infections (> 30 days) ultimately required instrumentation removal for eradication of the

Pull ter Gunne et al. noted that their management of SSI involved aggressive debridement (89.3%) with hardware retention (if stable) and revision of hardware (if unstable), followed by an average of 40 days of antibiotics. With this protocol, 76% of their deep infections were eradicated with a single debridement, although no comment was made about the chronicity of the SSI prior to reoperation [3]. Kowalski et al. reported on 30 acute SSIs (<30 days) with 80% successfully retaining implants with surgical debridement and IV antibiotics followed by oral suppressive antibiotics [4]. Tominaga et al. reviewed risk factors for unavoidable

removal of instrumentation after SSI < 90 days, finding that 12 of 16 cases successfully retained implants after debridement and IV antibiotics, but noted that 3 of 4 failures grew methicillin-resistant Staphylococcus aureus (MRSA) on operative cultures, compared with only 1 of 12 successfully-treated cases diagnosed with MRSA [5]. Nunez-Pereira et al. reported 43 patients with acute SSI after posterior spinal fusion requiring debridement and IV antibiotics for at least 8 weeks, finding 90.7% survival (survival to follow-up timepoint with avoidance of implant removal) at 6 months, 85.4% at 12 months, and 73.2% out to 4 years [6]. Multivariate analysis revealed a significant risk of treatment failure in patients who developed sepsis (hazard ratio 12.5 [95% confidence interval 2.6 to 59.9]; p < 0.001) or who had more than three fused segments (hazard ratio 4.5 [95% confidence interval 1.25 to 24.05]; p = 0.03) [1].

Accurately predicting the number of required debridements to eradicate the SSI can be challenging. Thalgott et al. identified that initial debridement culture results and the patient's comorbidities, including systemic disease, immunocompromise and malnourishment, are prognostic for the number of debridements required. Healthy patients with less virulent bacteria commonly required a single debridement, while immunocompromised hosts, multiple and/or more virulent organisms, and polymicrobial infections often require multiple debridements [7]. DiPaola et al. evaluated risk factors predicting multiple debridements, identifying MRSA and distant site infection as the strongest predictors, and diabetes mellitis, the presence of instrumentation, use of allograft and posterior lumbar spine location also displaying significant associations [8].

Conversely, delayed diagnoses of SSI commonly require implant removal for successful infection eradication. Hedequist et al. found all 26 cases with SSIs presenting greater than 3 months postoperatively required implant removal to definitively clear the infection [9]. Similarly, Kowalski et al. reported 7 out of 13 late diagnoses of SSI (> 30 days) failed debridement and initial implant retention, requiring secondary surgery for implant removal [4]. Tsubouchi et al. noted that although 29 out of 43 patients successfully retained spinal implants for SSI < 30 days postoperatively, only 4 of 12 patients diagnosed later than 30 days and 0 of 4 patients diagnosed later than 90 days successfully retained implants [10]. Garg et al. reported on 42 patients with deep infection more than 1 year postoperatively after spinal fusion, noting that 41 required implant removal and retention attempted in 1 patient failed. Additionally, 27 of the 42 patients showed *C. acnes* on intraoperative cultures [11].

Ho et al. reviewed their experience with pediatric SSI after instrumented fusion for scoliosis, noting that 43 out of 53 (81%) patients had retained implants at their first irrigation and debridement. They found a significant increase in secondary debridement required with implant retention (47%) in comparison to implant removal at the first irrigation and debridement (20%). However, implant removal was associated with a 10-degree or greater curve progression in 60% of patients [12]. Balancing the need for spinal stability and prevention of deformity progression or pseudarthrosis against a more complete eradication of infection remains a case-by-case decision guided by surgeon experience.

Mok et al. reviewed the functional impact of infection after posterior spinal fusion with 12 early (< 90 days) and 4 late (> 90 days) SSIs undergoing debridement with retention of instrumentation, and reported no significant difference in long-term SF-36 outcomes compared with non-infected controls at an average follow-up of 56.7

months [13]. Kuhns et al. similarly compared quality of life (QQL) scores between infected posterior cervical fusions requiring reoperation to noninfected matched controls. While the total projected costs were increased (\$21,778 vs. \$9,159) and 6-month QQLs were significantly lower for the infected cohort, no significant differences were found in QQL outcomes at the 12-month follow-up [14].

Recent literature has questioned the significance of time-based decision-making for implant removal following SSI and instead has turned to advanced imaging to understand the causes of implant retention failures. Kanavama et al. evaluated preoperative magnetic resonance imaging (MRIs) in SSIs, noting that once vertebral osteomyelitis and/or intervertebral abscess were evident in MR images, all the hardware should be removed [15]. Six of seven patients without osteomyelitis or intervertebral abscess successfully retained implants, while 9 of 13 patients with osteomyelitis or intervertebral abscess ultimately required implant removal and three of four patients who retained implants resulted in loss of fixation stability [15].

REFERENCES

- Patel H, Khoury H, Girgenti D, Welner S, Yu H. Burden of surgical site infections associated with select spine operations and involvement of Staphylococcus aureus. Surg Infect (Larchmt). 2017;18:461–473. doi:10.1089/ sur.2016.186.
- [2] Sierra-Hoffman M, Jinadatha C, Carpenter JL, Rahm M. Postoperative instrumented spine infections: a retrospective review. South Med J. 2010;103:25–30. doi:10.1097/SMJ.ob013e3181c4e00b.
- [3] Pull ter Gunné AF, Cohen DB. Incidence, prevalence, and analysis of risk factors for surgical site infection following adult spinal surgery. Spine. 2009;34:1422–1428. doi:10.1097/BRS.ob013e3181a03013.
- 2009;34:1422-1428. doi:10.1097/BRS.ob013e3181a03013.
 [4] Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913–920. doi:10.1086/512194.
- [5] Tominaga H, Setoguchi T, Kawamura H, Kawamura I, Nagano S, Abematsu M, et al. Risk factors for unavoidable removal of instrumentation after surgical site infection of spine surgery: A retrospective case-control study. Medicine (Baltimore). 2016;95:e5118. doi:10.1097/MD.000000000005118.
- [6] Núñez-Pereira S, Pellisé F, Rodríguez-Pardo D, Pigrau C, Bagó J, Villanueva C, et al. Implant survival after deep infection of an instrumented spinal fusion. Bone Joint J. 2013;95-B:1121-1126. doi:10.1302/0301-620X.95B8.30784.
- Thalgott JS, Cotler HB, Sasso RC, LaRocca H, Gardner V. Postoperative infections in spinal implants. Classification and analysis—a multicenter study. Spine. 1991;16:981–984.
- [8] DiPaola CP, Saravanja DD, Boriani L, Zhang H, Boyd MC, Kwon BK, et al. Post-operative infection treatment score for the spine (PITSS): construction and validation of a predictive model to define need for single versus multiple irrigation and debridement for spinal surgical site infection. Spine J. 2012;12:218–230. doi:10.1016/j.spinee.2012.02.004.
- [9] Hedequist D, Haugen A, Hresko T, Emans J. Failure of attempted implant retention in spinal deformity delayed surgical site infections. Spine. 2009;34:60–64. doi:10.1097/BRS.0b013e31818ed75e.
- [10] Tsubouchi N, Fujibayashi S, Otsuki B, Izeki M, Kimura H, Ota M, et al. Risk factors for implant removal after spinal surgical site infection. Eur Spine J. 2017. doi:10.1007/s00586-017-5294-1.
 [11] Garg S, LaGreca J, Hotchkiss M, Erickson M. Management of late (>1 y) deep
- [11] Garg S, LaGreca J, Hotchkiss M, Erickson M. Management of late (>1 y) deep infection after spinal fusion: a retrospective cohort study. J Pediatr Orthop. 2015;35:266–270. doi:10.1097/BPO.000000000000252.
- [12] Ho C, Skaggs DL, Weiss JM, Tolo VT. Management of infection after instrumented posterior spine fusion in pediatric scoliosis. Spine. 2007;32:2739–2744. doi:10.1097/BRS.obo13e31815a5a86.
- [13] Mok JM, Guillaume TJ, Talu Ū, Berven SH, Deviren V, Kroeber M, et al. Clinical outcome of deep wound infection after instrumented posterior spinal fusion: a matched cohort analysis. Spine. 2009;34:578–583. doi:10.1097/BRS.0b0192318194827c.
- [14] Kuhns BD, Lubelski D, Alvin MD, Taub JS, McGirt MJ, Benzel EC, et al. Cost and quality of life outcome analysis of postoperative infections after subaxial dorsal cervical fusions. J Neurosurg Spine. 2015;22:381–386. doi:10.3171/2014.10.SPINE14228.
- [15] Kanayama M, Hashimoto T, Shigenobu K, Oha F, Iwata A, Tanaka M. MRI-based decision making of implant removal in deep wound infection after instrumented lumbar fusion. Clin Spine Surg. 2017;30:E99–E103. doi:10.1097/BSD.ob013e3182aa4c72.

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QUESTION 3: Should bone graft be removed in patients with postoperative spine infection? If yes, should a distinction be made between allograft and autograft?

RECOMMENDATION: Bone graft need not be routinely removed following irrigation and debridement, especially if partially incorporated. However, loose or purulent graft should be considered for removal. Retained allograft may increase the risk for requiring repeat debridement compared to autograft.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 87%, Disagree: 0%, Abstain: 13% (Super Majority, Strong Consensus)

RATIONALE

No literature could be found that directly stratified patients who had bone graft retained versus removed. Weinstein et al. studied 46 postoperative infections in 2,391 patents [1]. In their regimen, bone graft material that appeared viable was left in place and instrumentation was retained as well. After six weeks of antibiotics, all of the wounds healed. Massie et al. similarly reported that bone graft may be retained and rarely is it necessary to remove all bone graft [2]. Ahmed et al. also showed in their retrospective review that debridement and antibiotics with implant and bone graft retention (allograft and autograft) can result in complete eradication of infection [3].

Nonetheless, bone graft loosened by irrigation may be removed. It seems rational that unincorporated bone graft and loose, dead bone serves as a continued nidus for infection and as such should be removed [4]. Multiple authors thus recommend thorough irrigation and debridement with removal of nonviable, purulent and loose graft material. However, this appears largely based upon intuition and not strict evidence.

There is limited evidence that perhaps autograft is better tolerated in the setting of an infection. Dipola et al. created a predictive model to differentiate patients requiring one versus multiple debridements [5]. The use of bone graft rather than autograft was shown to be predictive of requiring multiple debridements. Perhaps, therefore, closer attention ought to be given to the viability and infection burden in patients with allograft. However, no specific recommendations can be given and this should be considered on a case-by-case basis, with considerations of host status, infectious organism and infection burden.

REFERENCES

- Weinstein MA, McCabe JP, Cammisa FP. Postoperative spinal wound infection: a review of 2,391 consecutive index procedures. J Spinal Disord.
- Massie JB, Heller JG, Abitbol JJ, McPherson D, Garfin SR. Postoperative posterior spinal wound infections. Clin Orthop Relat Res. 1992:99–108.
- Ahmed R, Greenlee JDW, Traynelis VC. Preservation of spinal instrumentation after development of postoperative bacterial infections in patients undergoing spinal arthrodesis. J Spinal Disord Tech. 2012;25:299-302. doi:10.1097/BSD.obo13e31821fbf72.
- Hegde V, Meredith DS, Kepler CK, Huang RC. Management of postoperative spinal infections. World J Orthop. 2012;3:182–189. doi:10.5312/wjo.v3.i11.182. Dipaola CP, Saravanja DD, Boriani L, Zhang H, Boyd MC, Kwon BK, et al. Post-
- operative infection treatment score for the spine (PITSS): construction and validation of a predictive model to define need for single versus multiple irrigation and debridement for spinal surgical site infection. Spine J. 2012;12:218-230. doi:10.1016/j.spinee.2012.02.004.

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QUESTION 4: What are the indications for implant retention or removal of hardware in spinal infections?

RECOMMENDATION: In early or acute infections, debridement with retention of the implant might be possible and should always be favored, as removal of the implant carries a great risk for non-fusion despite the risk of chronic low-grade infections with possible implant loosening. In late infections, removal is recommended if feasible.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 87%, Disagree: 7%, Abstain: 6% (Super Majority, Strong Consensus)

RATIONALE

Similar to periprosthetic joint infections (PJI), several authors recommend that in early spinal implant-associated infections (within one month after surgical treatment or symptom duration less than three weeks), a debridement with retention of the implant constitutes a sufficient treatment strategy [1-5]. However, their recommendation is based on a retrospective, small case series of patients. There are also reports describing continuous irrigation in early infections [6,7], but no controlled studies with non-continuous irrigation are

In chronic infections, which are often caused by low-grade pathogens, such as coagulase-negative staphylococci or Cutibacterium acnes, removal of implants is regarded as the treatment of choice [3,8–10]. Infections with low-grade pathogens often present in a delayed fashion so that the implant-associated biofilm is mature and bacteria in the biofilm cannot be killed by antibiotics only or debridement with retention of the implant. In addition, patients with chronic infections often present with pseudarthrosis [11]. Hedequist et al. retrospectively reported on 26 chronic infections in which curing was only achieved after removal of the implants with prior unsuccessful treatment attempts with implant retention [12]. In six patients, hardware reimplantation was needed due to progression of the underlying deformity (curve progression). Implant removal carries the risk of disc collapse, lack of fusion, loss of normal lordosis and pseudarthrosis [3,13], which have to be considered.

There are no recommendations as to whether only the dorsal instrumentation or the interdiscal cage should be removed as well for successful treatment. In addition, no prospective clinical trials comparing removal versus retention of the implant in chronic infections exist. Lall et al. nicely summarized treatment regimens of deep wound infections after spinal instrumentation [14].

REFERENCES

- Viola RW, King HA, Adler SM, Wilson CB. Delayed infection after elective spinal instrumentation and fusion. A retrospective analysis of eight cases. Spine. 1997;22:2444–2450; discussion 2450-2451.
- Spine. 1997;22:2444–2450; discussion 2450-2451.

 [2] Falavigna A, Righesso O, Traynelis VC, Teles AR, da Silva PG. Effect of deep wound infection following lumbar arthrodesis for degenerative disc disease on long-term outcome: a prospective study: clinical article. J Neurosurg Spine 2011;15:200–403. doi:10.3171/2011.5 SPINE10825
- surg Spine. 2011;15:399–403. doi:10.3171/2011.5.SPINE10825.

 [3] Chen SH, Lee CH, Huang KC, Hsieh PH, Tsai SY. Postoperative wound infection after posterior spinal instrumentation: analysis of long-term treatment outcomes. Eur Spine J. 2015;24:561–570. doi:10.1007/s00586-014-3636-9.

- [4] Chaichana KL, Bydon M, Santiago-Dieppa DR, Hwang L, McLoughlin G, Sciubba DM, et al. Risk of infection following posterior instrumented lumbar fusion for degenerative spine disease in 817 consecutive cases. J Neurosurg Spine. 2014;20:45–52. doi:10.3171/2013.10.SPINE1364.
- Neurosurg Spine. 2014;20:45-52. doi:10.3171/2013.10.SPINE1364.

 [5] Picada R, Winter RB, Lonstein JE, Denis F, Pinto MR, Smith MD, et al. Postoperative deep wound infection in adults after posterior lumbosacral spine fusion with instrumentation: incidence and management. J Spinal Disord. 2000:13:42-45.
- 2000;13:42-45.

 Chikawa T, Sakai T, Bhatia NN, Sairyo K, Utunomiya R, Nakamura M, et al. Retrospective study of deep surgical site infections following spinal surgery and the effectiveness of continuous irrigation. Br J Neurosurg. 2011;25:621-624. doi:10.3109/02688697.2010.546902.
- [7] Lian XF, Xu JG, Zeng BF, Liu X-K, Li H, Qiu M, et al. Continuous irrigation and drainage for early postoperative deep wound infection after posterior instrumented spinal fusion. J Spinal Disord Tech. 2014;27:E315–E317. doi:10.1097/BSD.000000000000122.
- [8] Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19:398–404. doi:10.1007/s00776-014-0545-z.
- [9] Hahn F, Zbinden R, Min K. Late implant infections caused by Propionibacterium acnes in scoliosis surgery. Eur Spine J. 2005;14:783–788. doi:10.1007/s00586-004-0854-6.
- [10] Richards BR, Emara KM. Delayed infections after posterior TSRH spinal instrumentation for idiopathic scoliosis: revisited. Spine. 2001;26:1990-1996.
- [11] Pull ter Gunne AF, Mohamed AS, Skolasky RL, van Laarhoven CJHM, Cohen DB. The presentation, incidence, etiology, and treatment of surgical site infections after spinal surgery. Spine. 2010;35:1323-1328. doi:10.1097/ BRS.0b013e3181bcde61.
- [12] Hedequist D, Haugen A, Hresko T, Emans J. Failure of attempted implant retention in spinal deformity delayed surgical site infections. Spine. 2009;34:60–64. doi:10.1097/BRS.ob013e31818ed75e.
- [13] Kim JJ, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infection after instrumented spinal fusion. J Spinal Disord Tech. 2010;23:258–265. doi:10.1097/BSD.0b013e3181a9452c.
- [14] Lall RR, Wong AP, Lall RR, Lawton CD, Smith ZA, Dahdaleh NS. Evidence-based management of deep wound infection after spinal instrumentation. J Clin Neurosci. 2015;22:238–242. doi:10.1016/j.jocn.2014.07.010.

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Authors: Barrett Woods, Maja Babic

QUESTION 5: Is there a role for one-stage exchange of hardware in the presence of spinal infections?

RECOMMENDATION: There is insufficient data on one-stage exchange of hardware in the presence of spine infection.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

RATIONALE

Evidence supports debridement and implant retention in early implant-associated infections. In delayed implant-associated spine infections, evidence favors hardware removal followed by a course of antibiotics. Even if solid fusion is present, significant loss of correction can occur, posing the question of whether one-stage exchange of hardware would be adequate [1]. It is established that placing spinal instrumentation into an infected spine is safe when necessary for spinal stability and eradication infection, with low recurrence and reoperation rates [2]. Data on hardware one-stage exchange in deep infections with instrumentation is lacking.

Infection following instrumented spinal fusion can result in significant morbidity to the patient, resulting in prolonged hospitalization, chronic pain and need for revision surgery. In addition to the morbidity, the economic impact of this type of infection to the healthcare system and patient cannot be overstated. Several risk factors associated with the development of surgical site infection (SSI) following instrumented spinal fusion have been identified

[2–4]. Management of superficial infection typically consists of oral or intravenous (IV) antibiotics, with surgical intervention reserved for failure of medical management, symptomatic deep infections or draining wounds with soft tissue compromise. Treatment of deep infections surgically is complicated by the presence of spinal instrumentation. Eradication of infection is the primary goal of surgery, however premature removal of instrumentation can result in pain, pseudoarthrosis and deformity [5–7].

Several series have been published illustrating successful treatment of deep wound infection with irrigation debridement and retention of original instrumentation [8–14]. Picada et al. published on a series of 26 patients with infection following instrumented spinal procedures, with 24 (92.3%) successfully treated with surgical debridement, intravenous antibiotics, nutrition optimization and primary or delayed secondary closure [13].

Kowalski et al. retrospectively reviewed the management of 81 patients with infections following spinal instrumentation. The

cohorts were defined by early and late onset infection [9]. Of the patients with early onset infection, 28 of 30 were treated with irrigated debridement and retention of hardware with predicted probability of treatment success at two years being 71%, while patients with late onset infections required removal of hardware to achieve an 84% probability of treatment success at two years. Maruo et al. retrospectively reviewed a series of 225 consecutive patients with SSIs following spinal surgery [10]. Of those, 126 or 76% were successfully treated with surgical debridement, IV antibiotic therapy and retention of hardware. Failure of this treatment strategy was associated with late infection, long constructs with pelvic fixation, *Propionibac*terium acnes speciation and poly-microbial infection.

Nunez-Pereira et al. published on a series of 43 consecutive patients with SSI treated with surgical debridement and targeted antibiotic therapy with retention of original instrumentation [11]. At a 26-month follow-up, 10 patients (23,3%) failed, requiring removal of hardware, or died. Multivariate analysis found treatment failure associated with sepsis and long constructs (> three levels fused). Tominaga et al. published a retrospective series of 16 consecutive patients who developed SSI following spine instrumentation over an eight-year span [15]. Twelve of the 16 cases (75%) were successfully treated with retention of hardware, with failure associated with long instrumented constructs, previous spinal surgery, low preoperative hemoglobin, high preoperative creatinine and methicillin-resistant Staphylococcus aureus (MRSA) speciation. DiPaola et al. developed a predictive model determining the need for single versus multiple irrigation and debridement procedures to successfully eradicate postsurgical spinal infection [8]. The authors identified MRSA-positive cultures, bacteremia, non-autogenous bone graft and diabetics as predictive for requiring multiple debridement procedures. Vacuum-assisted closure (VAC) can be used to help facilitate wound healing following irrigation and debridement with hardware retention for spinal infection [16].

There are several studies illustrating the successful management of SSI following spinal instrumentation with surgical debridement, IV antibiotic therapy and primary or delayed secondary closure. Factors consistently associated with treatment failure included late infection, long constructs with pelvic fixation, C. acnes/MRSA speciation and bacteremia. Patients with these characteristics should likely have removal of hardware in addition to surgical debridement. Multiple debridement procedures may be required to successfully treat the infection, which can be assisted by the use of a wound VAC.

REFERENCES

- Lall RR, Wong AP, Lall RR, Lawton CD, Smith ZA, Dahdaleh NS. Evidencebased management of deep wound infection after spinal instrumentation. J Clin Neurosci. 2015;22:238–242. doi:10.1016/j.jocn.2014.07.010.
- Pull ter Gunne AF, Cohen DB. Incidence, prevalence, and analysis of risk factors for surgical site infection following adult spinal surgery. Spine. 2009;34:1422-1428. doi:10.1097/BRS.obo13e3181a03013.
- Veeravagu A, Patil CG, Lad SP, Boakye M. Risk factors for postoperative spinal wound infections after spinal decompression and fusion surgeries. Spine. 2009;34:1869–1872. doi:10.1097/BRS.obo13e3181adc989.
- Xing D, Ma JX, Ma XL, Song DH, Wang J, Chen Y, et al. A methodological, systematic review of evidence-based independent risk factors for surgical site infections after spinal surgery. Eur Spine J. 2013;22:605-615. doi:10.1007/ s00586-012-2514-6.
- Weiss LE, Vaccaro AR, Scuderi G, McGuire M, Garfin SR. Pseudarthrosis after postoperative wound infection in the lumbar spine. J Spinal Disord.
- Alpert HW, Farley FA, Caird MS, Hensinger RN, Li Y, Vanderhave KL. Outcomes following removal of instrumentation after posterior spinal fusion. J Pediatr Orthop. 2014;34:613-617. doi:10.1097/BPO.0000000000000145. Kim JI, Suh KT, Kim SJ, Lee JS. Implant removal for the management of infec-
- tion after instrumented spinal fusion. J Spinal Disord Tech. 2010;23:258-265. doi:10.1097/BSD.obo13e3181a9452c.
- DiPaola CP, Saravanja DD, Boriani L, Zhang H, Boyd MC, Kwon BK, et al. Postoperative infection treatment score for the spine (PITSS): construction and validation of a predictive model to define need for single versus multiple irrigation and debridement for spinal surgical site infection. Spine J. 2012;12:218–230. doi:10.1016/j.spinee.2012.02.004.
- Kowalski TJ, Berbari EF, Huddleston PM, Steckelberg JM, Mandrekar JN, Osmon DR. The management and outcome of spinal implant infections: contemporary retrospective cohort study. Clin Infect Dis. 2007;44:913-920. doi:10.1086/512194.
- Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. J Orthop Sci. 2014;19:398–404. doi:10.1007/s00776-014-0545-z.
- Lee MC, Wang MY, Fessler RG, Liauw J, Kim DH. Instrumentation in patients with spinal infection. Neurosurg Focus. 2004;17:E7.
- Núñez-Pereira S, Pellisé F, Rodríguez-Pardo D, Pigrau C, Bagó J, Villanueva C, et al. Implant survival after deep infection of an instrumented spinal fusion. Bone Joint J. 2013;95-B:1121-1126. doi:10.1302/0301-620X.95B8.30784
- Wimmer C, Gluch H. Management of postoperative wound infection in posterior spinal fusion with instrumentation. J Spinal Disord. 1996;9:505-
- Picada R, Winter RB, Lonstein JE, Denis F, Pinto MR, Smith MD, et al. Postoperative deep wound infection in adults after posterior lumbosacral spine fusion with instrumentation: incidence and management. J Spinal Disord. 2000;13:42-45
- Tominaga H, Setoguchi T, Kawamura H, Kawamura I, Nagano S, Abematsu M, et al. Risk factors for unavoidable removal of instrumentation after surgical site infection of spine surgery: a retrospective case-control study. Medicine (Baltimore). 2016;95:e5118. doi:10.1097/MD.00000000000118.
- Canavese F, Gupta S, Krajbich JI, Emara KM. Vacuum-assisted closure for deep infection after spinal instrumentation for scoliosis. J Bone Joint Surg Br. 2008;90:377-381. doi:10.1302/0301-620X.90B3.19890.

3.4. TREATMENT: WOUND CARE

Authors: Carles Pigrau, Gregory Schroeder

QUESTION 1: Should infected wounds undergo primary closure or a two-stage closure?

RECOMMENDATION: The current recommended practice for spine wounds remains primary closure in the majority of postoperative infections. However, there may be circumstances when primary closure of the wound may not be possible or preferred. This may include patients with grossly contaminated traumatic wounds, patients with persistent wound drainage when attempts to address drainage have failed or patients with severe soft tissue loss when primary closure is not possible.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Following surgery, wounds are typically closed in a primary fashion. Alternative methods of wound closure include secondary closure and delayed primary closure. Secondary closure is when wounds are left to close naturally on their own. Delayed primary closure (DPC), a combination of secondary and primary closure, is when a wound is cleaned and left open until infection is controlled, followed by surgical closure of the wound. Delayed primary closure is only used on occasion, typically involving contaminated traumatic injuries.

In their prospective randomized study, Singh et al. found that patients undergoing delayed primary closure of contaminated abdominal wounds related to hollow viscus perforation had lower infection rates (17.5%) and shorter hospital stays (18.1 days) when compared to patients undergoing primary closure (42.5% infection and 20.7 days) [1]. Chiang et al. found a similar result for treatment of perforated appendicitis. Patients randomized to primary closure had an infection rate of 38.9% and an 8.4-day length of stay, while patients randomized to delayed primary closure had an infection rate of 2.9% and a 6.3-day length of stay [2].

DPC has also been shown to result in no long-term issues and not be associated with a higher incidence of complications in pediatric liver transplant recipients [3]. Orthopaedic surgeons are familiar with DPC in the context of fasciotomy wounds in patients with compartment syndrome when delayed primary closure is utilized [4,5].

There are, however, no high-level studies related to the role of DPC in spine surgery. In the absence of concrete evidence, and in borrowing from general surgery and other fields of orthopaedics, we feel that primary closure of a wound is the most preferred method of dealing with wound issues in spine patients. However, there may be circumstances when primary closure of the wound may not be possible or preferred. This may include patients with grossly contaminated traumatic wounds, patients with persistent wound drainage when attempts to address drainage have failed and in patients with severe soft tissue loss when primary closure is not possible.

REFERENCES

- [1] Singh PK, Saxena N, Poddar D, Gohil RK, Patel G. Comparative study of wound healing in primary versus delayed primary closure in contaminated abdominal surgery. Hellenic J Surg. 2016;88(5):314–320. doi:10.1007/s13126-016-0340-8.
- [2] Chiang R, Chen S, Tsai Y. Delayed primary closure versus primary closure for wound management in perforated appendicitis: a prospective randomized controlled trial. J Chin Med Assoc. 2012;75(4):156–159. doi:10.1016/j.jcma.2012.02.013.
- [3] Ziaziaris WA, Darani A, Holland AJ, Alexander A, Karpelowsky J, Shun A, Thomas G. Delayed primary closure and the incidence of surgical complications in pediatric liver transplant recipients. J Pediatr Surg. 2015;50(12):2137– 2140. doi:10.1016/j.jpedsurg.2015.08.045.
- 2140. doi:10.1016/j.jpedsurg.2015.08.045.
 [4] Rijal L, Nepal P, Adhikari A, Regmi S. Luggage tag tie technique for delayed primary closure of fasciotomy wounds. Eur J Orthop Surg Traumatol. 2011;21(6):249–452. doi:10.1007/s00590-010-0729-V.
- 2011;21(6):449–452. doi:10.1007/s00590-010-0729-y.

 [5] Barnea Y, Gur E, Amir A, Leshem D, Zaretski A, Miller E, Weiss J. Delayed primary closure of fasciotomy wounds with Wisebands®, a skin- and soft tissue-stretch device. Injury. 2006;37(6):561–566. doi:10.1016/j. injury.2006.02.056.

Author: Wesley Bronson

QUESTION 2: What is the indication for muscle advancement flaps in patients with spinal infections?

RECOMMENDATION: Muscle advancement flaps are useful to help close wounds with exposed hardware as well as those which fail local treatment/vacuum-assisted closure (VAC) therapy and to help improve infection eradication.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 93%, Disagree: 0%, Abstain: 7% (Super Majority, Strong Consensus)

RATIONALE

Multiple risk factors exist for wound complications following spinal surgery, including diabetes, chronic obstructive pulmonary disease, resection of neoplasm with excision of significant soft tissue and prior radiation. Additionally, infection is often complicated by loss of soft tissue and poor tissue viability, which leads to an inability to close the wound overall, resulting in exposed hardware [1,2].

Even if the wound is able to be closed primarily or following VAC therapy, it is important to recognize that the same factors that led to the infection and wound breakdown in the first place still exist [3]. To that end, local or vascularized muscle flaps provide multiple advantages over simple wound closure or delayed primary closure. Muscle flaps have been shown to increase blood flow and oxygen delivery, and decrease bacterial load [4-6].

It seems rational that wounds that are completely unable to be closed due to large soft tissue defects with exposed hardware or wounds that fail to close following VAC therapy are reasonable indications for flap coverage. But, the absolute indication for flap coverage following wound debridement in an otherwise closeable wound remains unclear. Multiple authors argue that it remains a reasonable option versus irrigation and debridement with immediate or delayed primary closure.

Dumanian et al. reviewed their experience with flap coverage for spinal wounds [7]. Fifteen patients in their group had postoperative wound dehiscence or infection, with 12 patients having exposed hardware. They were treated with either immediate local flap coverage or two to three days of dressing changes followed by flap coverage. Of the surviving 14 patients, 13 had healed wounds at final follow-up, and none required hardware removal. One patient on chronic steroids/immunosuppression had persistent infection treated with chronic suppressive antibiotics.

Chieng et al. performed a systematic review on the use of flaps for management of wound complications [8]. While several case reports and retrospective series present supportive data, the authors note that relying on the data is difficult as no level 1 or level

2 evidence exists. Additionally, there is a lack of comparative studies directly looking at flap coverage versus traditional wound closure techniques.

REFERENCES

- Koutsoumbelis S, Hughes AP, Girardi FP, Cammisa FP, Finerty EA, Nguyen JT, et al. Risk factors for postoperative infection following posterior lumbar instrumented arthrodesis. J Bone Joint Surg Am. 2011;93:1627–1633. doi:10.2106/JBIS.L00020.
- doi:10.2106/JBJS.J.00039.

 [2] Chang DW, Friel MT, Youssef AA. Reconstructive strategies in soft tissue reconstruction after resection of spinal neoplasms. Spine. 2007;32:1101–1106. doi:10.1097/01.brs.0000261555,72265,3f.
- [3] Mitra A, Mitra A, Harlin S. Treatment of massive thoracolumbar wounds and vertebral osteomyelitis following scoliosis surgery. Plast Reconstr Surg. 2004;113:206–213. doi:10.1097/01.PRS.0000097440.15013.5C.
- [4] Eshima I, Mathes SJ, Paty P. Comparison of the intracellular bacterial killing activity of leukocytes in musculocutaneous and random-pattern flaps. Plast Reconstr Surg. 1990;86:541–547.
- Reconstr Surg. 1990;86:541–547.

 Murphy RC, Robson MC, Heggers JP, Kadowaki M. The effect of microbial contamination on musculocutaneous and random flaps. J Surg Res. 1986;41:75–80.
- 6] Calderon W, Chang N, Mathes SJ. Comparison of the effect of bacterial inoculation in musculocutaneous and fasciocutaneous flaps. Plast Reconstr Surg. 1986;77:785–794.
- [7] Dumanian GA, Ondra SL, Liu J, Schafer MF, Chao JD. Muscle flap salvage of spine wounds with soft tissue defects or infection. Spine. 2003;28:1203–1211.
- doi:10.1097/01.BRS.0000067260.22943.48.
 [8] Chieng LO, Hubbard Z, Salgado CJ, Levi AD, Chim H. Reconstruction of open wounds as a complication of spinal surgery with flaps: a systematic review. Neurosurg Focus. 2015;39:E17. doi:10.3171/2015.7.FOCUS15245.

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Authors: Koji Yamada, Kazuhiro Kohata

QUESTION 3: What is the optimal irrigation solution (volume, type and frequency) during clean or infected spinal surgery cases?

RECOMMENDATION:

- 1. There is insufficient evidence to recommend for or against normal saline irrigation before closure for the purpose of preventing surgical site infection (SSI) in clean spinal surgery.
- There is insufficient evidence to support recommendations for optimal volume, type and frequency of irrigation to prevent SSI in clean spinal surgery.
- Consider the use of irrigation with an aqueous povidone-iodine solution before closure for the purpose of preventing SSI in clean spinal surgery.
- 4. There is insufficient evidence to recommend for or against chlorhexidine and antibiotic solution irrigation of incisional wounds for the purpose of preventing SSI in clean spinal surgery.
- 5. There is insufficient evidence to recommend a specific solution (volume, type and frequency) for irrigation in infected spinal surgery.

LEVEL OF EVIDENCE:

- 1. Consensus
- 2. Limited
- Moderate
- 4. Consensus
- 5. Consensus

DELEGATE VOTE: Agree: 73%, Disagree: 7%, Abstain: 20% (Super Majority, Strong Consensus)

RATIONALE

1: Irrigation versus no irrigation

No randomized controlled trials (RCTs) or observational studies have compared incisional wound irrigation with normal saline versus no irrigation in clean spinal surgery.

One retrospective observational study evaluating 1,831 posterior lumbar interbody fusion (PLIF) procedures demonstrated a significantly higher risk of SSI with no local bone irrigation compared to those with local bone irrigation in multivariate analysis (odds ratio (OR): 5.248, p = 0.001) [1]. Two retrospective observational studies demonstrated no significant association between interbody irrigation with SSI compared with no interbody irrigation in those undergoing PLIF and lumbar microdiscectomy [1,2].

2: Optimal volume, type and frequency of irrigation for clean spinal surgery

No RCT has compared the amount of normal saline for irrigation to prevent SSI in spinal surgery. One observational study including 223 consecutive spinal operations in a single university

hospital demonstrated a significant association with prevention of SSI (OR o.o8, 95%, confidence interval (CI) o.o1 to o.61) with sufficient amount of saline (mean > 2,000 ml per hour compared with <1,000 ml per hour) in a multivariate analysis [3].

No RCT or observational study has compared the frequency of irrigation to prevent SSI in spinal surgery.

A very low quality of evidence from two observational studies demonstrated a benefit of pulse pressure irrigation compared to bulb syringe irrigation with normal saline [4,5]. One study showed an advantage of decreasing wound contamination rate in PLIF surgical procedures (OR:6.35, p = 0.046) [4]. Another study showed significant decrease of postoperative infection by ten-fold (11% [28/261] vs. 0.7% [2/263], p < 0.001) by using pulsatile irrigation with vancomycin and ceftazidime prophylaxis for posterior spinal fusion surgeries in adolescent idiopathic scoliosis patients [5].

3 and 4: Optimal solution for clean spinal surgery

There is moderate-quality evidence from two RCTs and two observational studies that povidone iodine irrigation has a signifi-

cant benefit in reducing SSI risk in patients with primarily closed surgical incisions when compared to conventional normal saline wound irrigation [6–9]. In one RCT focusing on primary instrumented lumbosacral posterolateral fusion performed by the same surgeon, SSI was significantly lower in those who underwent 0.35% povidone-iodine irrigation compared with normal saline irrigation (0% [0/120] vs. 4.8% [6/124], p = 0.029), with no significant difference in fusion rate, wound healing, improvement of pain score, function score and ambulatory capacity [6].

In another RCT focusing on spinal surgery, SSI was significantly lower in those who underwent 0.35% povidone-iodine irrigation compared with normal saline irrigation (0% [0/208] vs. 3.4% [7/206], p = 0.0072) [7]. In one observational study comparing before and after the application of combination of 0.3% betadine irrigation with intra-wound vancomycin (VCM) powder (1 gm), the incidence of SSI significantly decreased after intervention (1.3% [15/1173] vs. 2.4% [30/1,252], p = 0.042) with a protective effect in multivariate analysis (OR 0.23, 95% CI: 0.06-0.86; p = 0.0287) [8]. In another observational study involving 950 spinal surgeries comparing before and after application of povidone-iodine and hydrogen peroxide solution irrigation, those irrigated with povidone-iodine and hydrogen peroxide solution were less likely to develop SSI compared with pre-intervention period (0% [0/490] vs. 1.5% [7/460]) [9].

No RCT or observational study has compared chlorhexidine or antibiotic solution irrigation to normal saline irrigation to prevent SSI in spinal surgery.

5: Optimal irrigation for infected spinal surgery

No RCT or observational study has compared incisional wound irrigation with no irrigation in infected spinal surgery.

REFERENCES

- [1] Kim JH, Ahn DK, Kim JW, Kim GW. Particular features of surgical site infection in posterior lumbar interbody fusion. Clin Orthop Surg. 2015;7:337–343. doi:10.4055/cios.2015.7.3.337.
- [2] Zhu RS, Ren YM, Yuan JJ, Cui ZJ, Wan J, Fan BY, et al. Does local lavage influence functional recovery during lumber discectomy of disc herniation?: One year's systematic follow-up of 410 patients. Medicine (Baltimore). 2016;95:e5022. doi:10.1097/MD.0000000000005022.
- [3] Watanabe M, Sakai D, Matsuyama D, Yamamoto Y, Sato M, Mochida J. Risk factors for surgical site infection following spine surgery: efficacy of intraoperative saline irrigation. J Neurosurg Spine. 2010;12:540–546. doi:10.3171/2009.11.SPINE09308.
- [4] Ahn DK, Lee S, Moon SH, Kim DG, Hong SW, Shin WS. Bulb syringe and pulsed irrigation: which is more effective to remove bacteria in spine surgeries? Clin Spine Surg. 2016;29:34–37. doi:10.1097/BSD.0000000000000068.
- Clin Spine Surg. 2016;29:34–37. doi:10.1097/BSD.00000000000000000068.

 [5] Myung KS, Glassman DM, Tolo VT, Skaggs DL. Simple steps to minimize spine infections in adolescent idiopathic scoliosis. J Pediatr Orthop. 2014;34:29–33. doi:10.1097/BPO.0b013g:1829b2d75.

 [6] Chang FY, Chang MC, Wang ST, Yu WK, Liu CL, Chen TH. Can povidone-
- [6] Chang FY, Chang MC, Wang ST, Yu WK, Liu CL, Chen TH. Can povidoneiodine solution be used safely in a spinal surgery? Eur Spine J. 2006;15:1005– 1014. doi:10.1007/s00586-005-0975-6.
- 1014. doi:10.1007/s00586-005-0975-6.
 [7] Cheng MT, Chang MC, Wang ST, Yu WK, Liu CL, Chen TH. Efficacy of dilute betadine solution irrigation in the prevention of postoperative infection of spinal surgery. Spine. 2005;30:1689-1693.
- [8] Tomov M, Mítsunaga L, Durbin-Johnson B, Nallur D, Roberto R. Reducing surgical site infection in spinal surgery with betadine irrigation and intrawound vancomycin powder. Spine. 2015;40:491–499. doi:10.1097/ BRS.000000000000000789.
- BRS.000000000000789.

 [9] Ulivieri S, Toninelli S, Petrini C, Giorgio A, Oliveri G. Prevention of post-operative infections in spine surgery by wound irrigation with a solution of povidone-iodine and hydrogen peroxide. Arch Orthop Trauma Surg. 2011;131:1203–1206. doi:10.1007/S00402-011-1262-0.

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Author: Carles Pigrau

QUESTION 4: Is negative pressure wound therapy (NPWT) effective in the treatment of wounds that are left to heal by secondary intention?

RECOMMENDATION: There is no evidence that NPWT is superior to conventional standard dressing changes in the treatment of wounds that are left to heal by secondary intention.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 60%, Disagree: 20%, Abstain: 20% (Super Majority, Weak Consensus)

RATIONALE

Animal studies have shown that sub-atmospheric pressure improves the local wound environment through both direct and indirect effects. Sub-atmospheric pressure accelerates healing and reduces the time to wound closure and the incidence of wound infections [1,2]. NPWT removes interstitial fluid and improves lymphatic drainage and microvascular blood flow. It increases oxygen and nutrient delivery in the wound, facilitates removal of metabolic byproducts, increases granulation tissue formation and ultimately accelerates wound healing. Moreover, by isolating the wound from the surrounding environment, NPWT may reduce the colonization of the wound by bacteria and avoid superinfections, particularly in areas with high skin contamination rates such as the perineal and lower back spine area.

Predominantly observational studies, but also small trials (low quality of evidence), have suggested that rates of surgical site infection (SSI) may be lower if NPWT is used instead of conven-

tional wound dressings [3]. In a meta-analysis of six randomized control trials including a systematic review, it was observed that the risk of SSI was reduced when NPWT was used (odds ratio 0.56, 95% CI 0.32 to 0.96) in both clean and clean-contaminated procedures. However, results were no longer significant for orthopaedic/trauma surgery [3]. In a Cochrane meta-analysis that compared NPWT with other types of wound dressing for persistently-draining wounds in skin graft patients, in orthopaedic patients undergoing arthroplasty and general/trauma surgery patients it was concluded that there is no evidence for the effectiveness of NPWT on the complete healing of wounds expected to heal by primary intention [4]. An up-to date systematic review in trauma patients concluded that, based on available observational studies, NPWT [5] was safe and showed an efficacy comparable to standard dressings [6]. The primary clinical advantages of NPWT in the trauma population are its ease of application, decreased number of dressing changes and reduction in the complexity of subsequent reconstructive procedures [7–11].

In a 2013 systematic review of NPWT for spinal wounds, no randomized clinical trials were found that addressed the use of NPWT to treat wound healing or spine SSIs, nor as prophylactic wound treatment to prevent wound breakdown and infection [12]. The duration of NPWT therapy and the number of debridement and irrigation procedures performed before the definitive wound closure operation were variable. After this review, an additional noncomparative study [12] showed the benefits of this therapy among only 6 of 317 infections after surgery for spinal stenosis. An average of 5.1 debridement and irrigation procedures were performed before the definitive wound closure operation. Vacuum-assisted closure dressings were changed at 3-day intervals and the median duration was 15 days (range 9-24).

After the revision published in 2013, only one longitudinal cohort study addressed NWPT use as a prophylactic therapy for spinal wounds. It is a well-designed, retrospective longitudinal study, which includes 160 adult patients with thoraco-lumbar spine deformity undergoing multi-level thoraco-lumbar fusion [13]. A 50% decrease in the incidence of wound dehiscence was observed in the NPWT cohort (46 cases) compared to the non-NPWT cohort (114 patients) and the incidence of postoperative SSI was significantly lower (10.6% vs 14.9%, p = 0.04).

In conclusion, prophylactic use of NWPT may significantly reduce wound dehiscence and wound infection after long-segment thoraco-lumbar spine fusion. There is no further evidence addressing the superiority of NWPT therapy compared to standard dressings. NPWT is safe in cases without dural leaks, easy to apply, and it decreases the number of dressing changes and reduces the complexity of wound closure. All these factors favor its use in selected cases.

REFERENCES

- Morykwas MJ, Argenta LC, Shelton-Brown EI, McGuirt W. Vacuum-assisted closure: a new method for wound control and treatment: animal studies and basic foundation. Ann Plast Surg. 1997;38:553–562.
 Chen SZ, Li J, Li XY, Xu LS. Effects of vacuum-assisted closure on wound
- [2] Chen SZ, Li J, Li XY, Xu LS. Effects of vacuum-assisted closure on wound microcirculation: an experimental study. Asian J Surg. 2005;28:211–217. doi:10.1016/S1015-9584(09)60346-8.
- [3] De Vries FÉE, Wallert ED, Solomkin JS, Allegranzi B, Egger M, Dellinger EP, et al. A systematic review and meta-analysis including GRADE qualification of the risk of surgical site infections after prophylactic negative pressure wound therapy compared with conventional dressings in clean and contaminated surgery. Medicine (Baltimore). 2016;95:e4673. doi:10.1097/MD.0000000000004673.
- MD.000000000004673.
 [4] Webster J, Scuffham P, Sherriff KL, Stankiewicz M, Chaboyer WP. Negative pressure wound therapy for skin grafts and surgical wounds healing by primary intention. Cochrane Database Syst Rev. 2012:CD009261. doi:10.1002/14651858.CD009261.pub2.
- [5] Gestring M. Negative pressure wound therapy. https://www.uptodate.com/
- contents/negative-pressure-wound-therapy. 2018.

 [6] Kanakaris NK, Thanasas C, Keramaris N, Kontakis G, Granick MS, Giannoudis PV. The efficacy of negative pressure wound therapy in the management of lower extremity trauma: review of clinical evidence. Injury. 2007;38

 Suppl 5:S9–S18. doi:10.1016/j.injury.2007.10.029.
- [7] Argenta LC, Morykwas MJ. Vacuum-assisted closure: a new method for wound control and treatment: clinical experience. Ann Plast Surg. 1997;38:563–576; discussion 577.
- [8] Barendse-Hofmann MG, van Doorn L, Steenvoorde P. Circumferential application of VAC on a large degloving injury on the lower extremity. J Wound Care. 2009;18:79–82. doi:10.12968/jowc.2009.18.2.38747.
- [9] DeFranzo AJ, Marks MW, Argenta LC, Genecov DG. Vacuum-assisted closure for the treatment of degloving injuries. Plast Reconstr Surg. 1999;104:2145– 2148
- [10] DeFranzo AJ, Argenta LC, Marks MW, Molnar JA, David LR, Webb LX, et al. The use of vacuum-assisted closure therapy for the treatment of lower-extremity wounds with exposed bone. Plast Reconstr Surg. 2001;108:1184-1101.
- [11] Meara JG, Guo L, Smith JD, Pribaz JJ, Breuing KH, Orgill DP. Vacuum-assisted closure in the treatment of degloving injuries. Ann Plast Surg. 1999;42:589–594.
- [12] Ousey KJ, Atkinson RA, Williamson JB, Lui S. Negative pressure wound therapy (NPWT) for spinal wounds: a systematic review. Spine J. 2013;13:1393–1405. doi:10.1016/j.spinee.2013.06.040.
- [13] Adogwa O, Fatemi P, Perez E, Moreno J, Gazcon GC, Gokaslan ZL, et al. Negative pressure wound therapy reduces incidence of postoperative wound infection and dehiscence after long-segment thoracolumbar spinal fusion: a single institutional experience. Spine J. 2014;14:2911–2917. doi:10.1016/j. spinee.2014.04.011.

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